



Lindee Climo

Clinical Pathology in Practice

Nicole Fernandez, DACVP
November 3, 2018

My Desk



Clinical Pathology in Practice

- Ask the audience
- Sample handling
- Cytology
- Hematology

Ask the Audience

- From your smart phone, tablet, or laptop



Respond at **PollEv.com/nicolefernan414**



Text **NICOLEFERNAN414** to **37607** once to join, then **A, B, or C**

What is your practice type?

Exclusively small animal

Exclusively food animal

Exclusively equine

Exclusively companion
animal (small and equine)

Exclusively large animal
(food animal and equine)

Other

Why did you decide to attend this session?

Clin path is AWESOME

I am comfortable with
cytology and hematology
but could use a review

I have forgotten all my
cytology and hematology
from vet school

Sample Handling

Resources

Cytology samples

Hematology samples



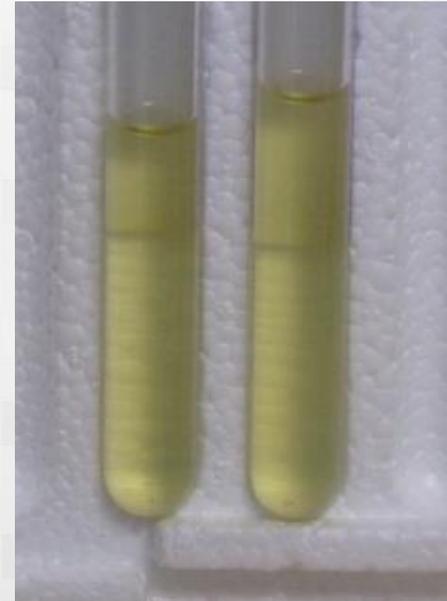
Sample Handling

- PDS Protocol Manual is online
- Search “pds protocol manual”
- <http://pdsinc.ca/Portals/0/Protocol%20Manual%202017%20with%20bookmarks.pdf>

Sample Handling

PDS lab techs recommend:

- Separate bag for each patient
- Label containers with patient name, date, sample type
 - Is this serum or urine?
- Wipe off outside of containers
- More padding than you think



Sample Handling

Cytology smears

- Air dry slides
- Adequate padding/proper containers
- Pack separately from histopath samples
 - Applies to blood smears too

Sample Handling

Fluid samples

- Submit in EDTA
- Make direct smears
- Keep cool

Sample Handling

Hematology samples

- CBC- EDTA tube
- Make blood smears

- Chemistry- red top tube
- Separate serum from cells if possible

- Keep cool

Cytology

Sample collection

Approach to the slide

A few lumps and bumps



Owen Fink

Sample Collection

- FNA and/or FNNA
 - Videos in PDS Protocol Manual
- Make smears quickly
- Air dry
- Stain with Diff-Quik

Do you collect cytology samples in-house?

Yes, all the
time

Yes,
occasionally

No

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 Text **NICOLEFERNAN414** to **37607** once to join, then **A, B, or C**

Do you read your own cytology samples?

Yes, most of the time

Yes, I look at them but usually submit them to a diagnostic lab

No, I submit them all

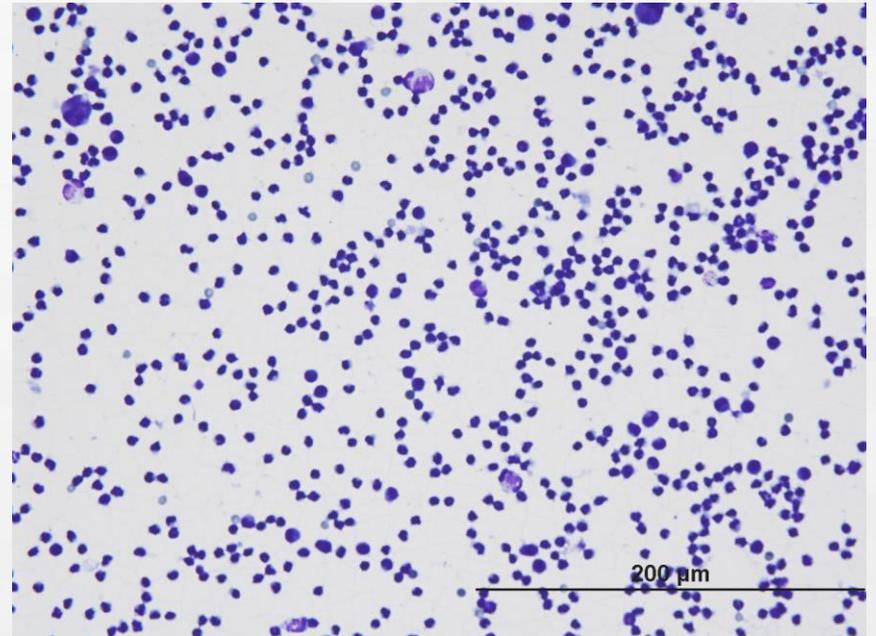
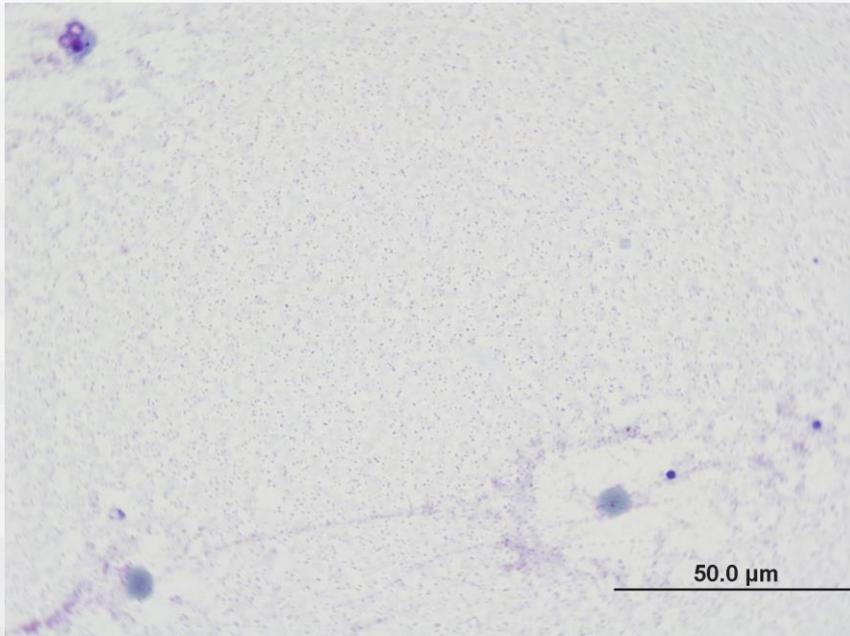
Approach to the Slide

1. Gross Examination
 - How cellular is the slide?
 - Where on the slide is the material distributed?
 - Is blood present?
2. 4x objective
 - Scan entire smear
 - Find areas to evaluate at higher magnification
 - Intact cells spread in a thin layer
3. 10x-40x objective
 - Evaluate several areas identified at 4x
 - Identify cell types present
 - Classify the lesion (see Figure 1)
4. 100x objective (oil)
 - Confirm identification of cells
 - Evaluate cellular details
 - Identify bacteria

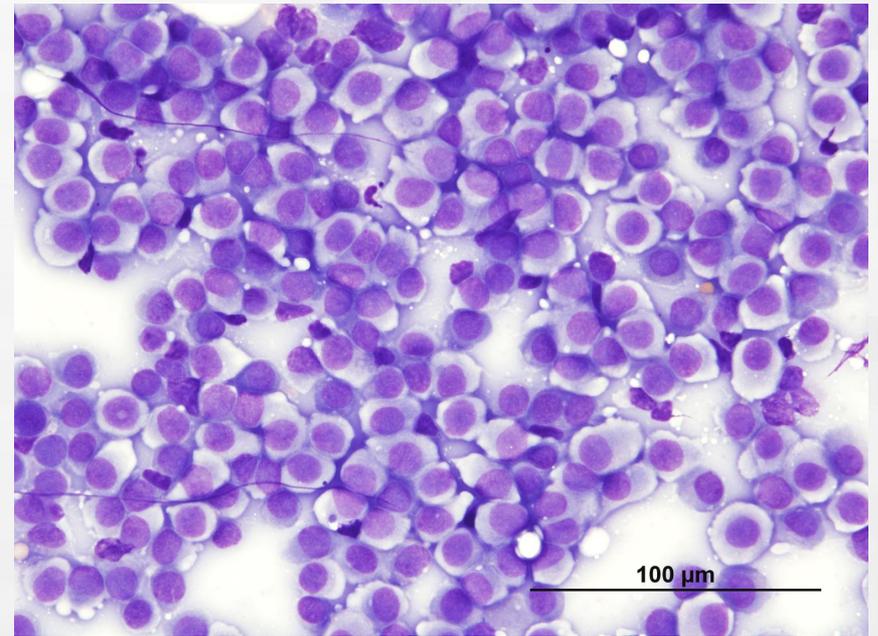
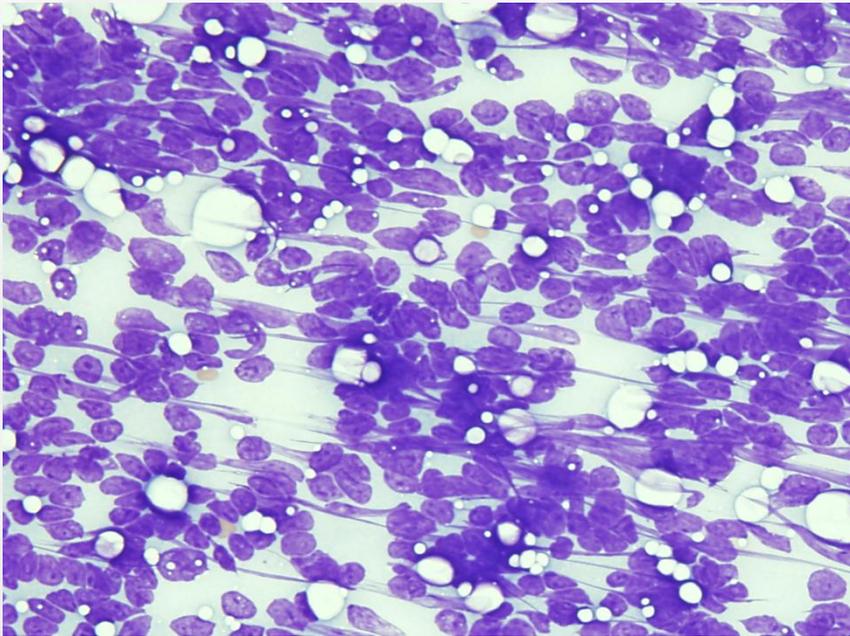
About the images

- Cases from WCVM and UCVM
- Many taken by Dr Galezowski

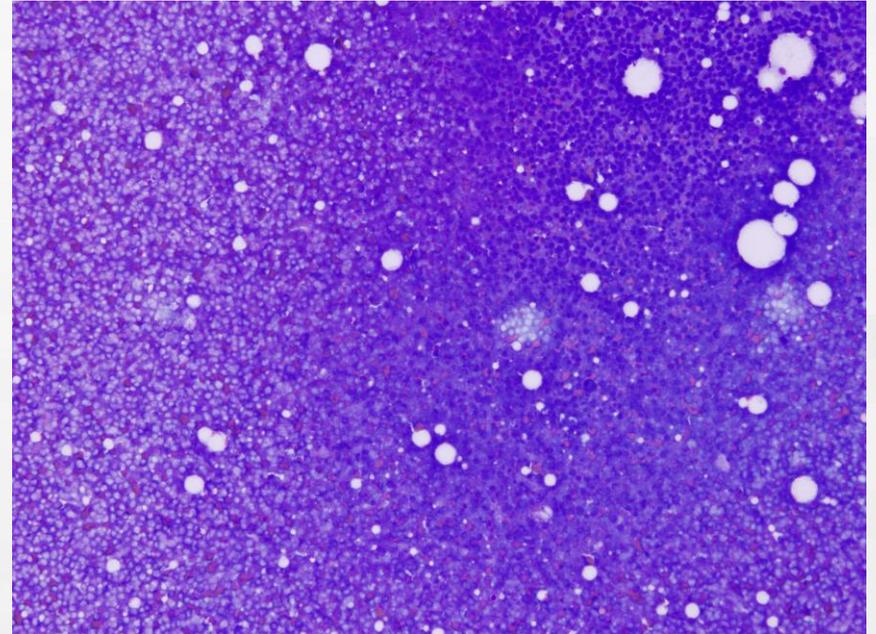
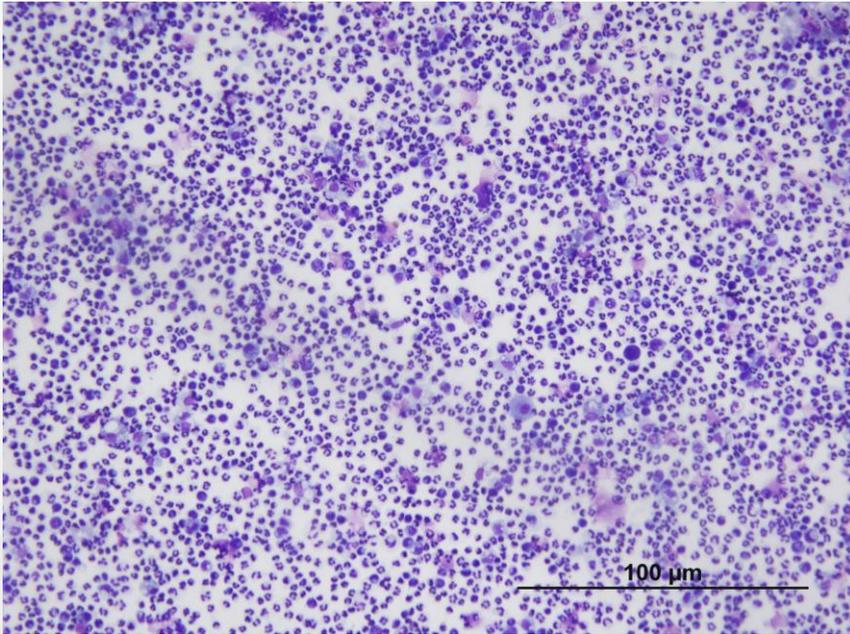
Cells needed...



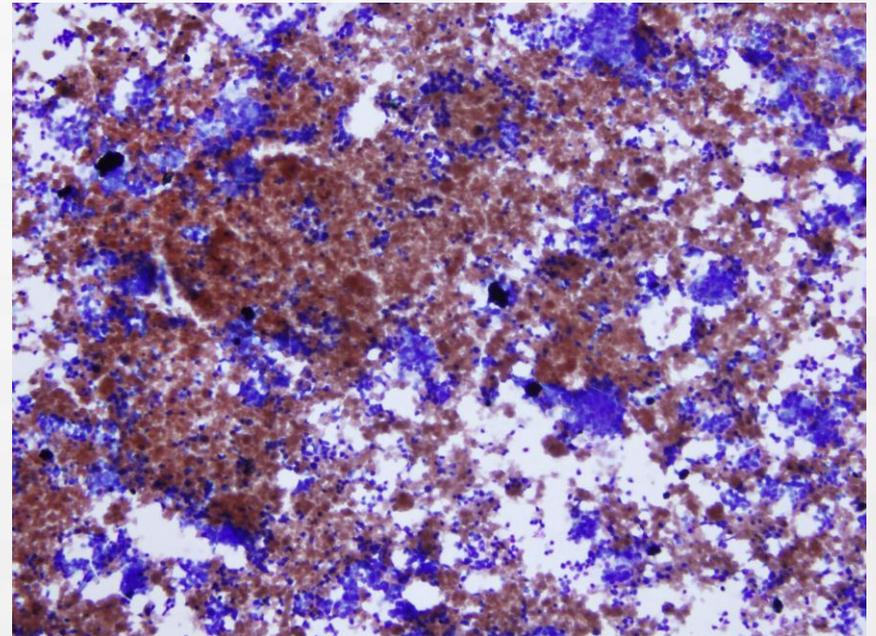
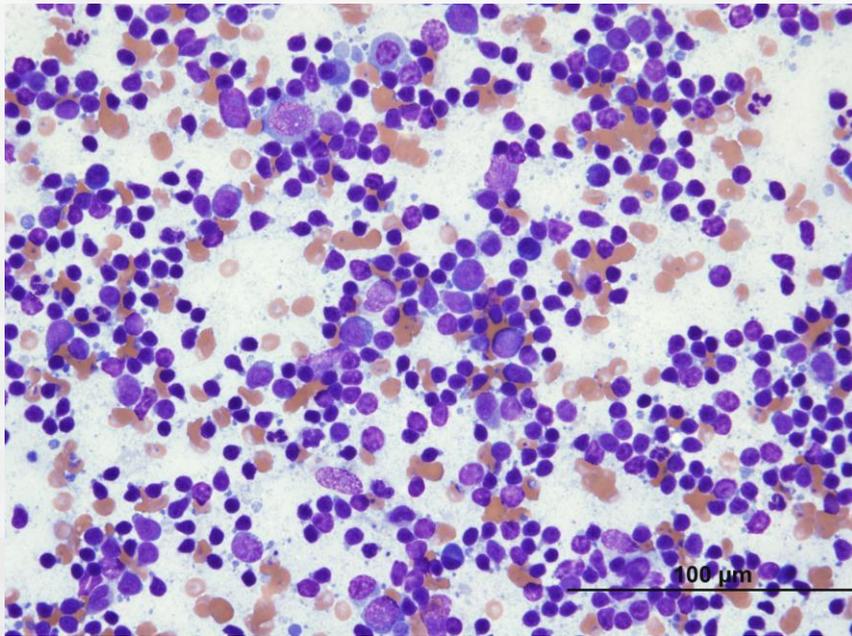
Intact cells needed...

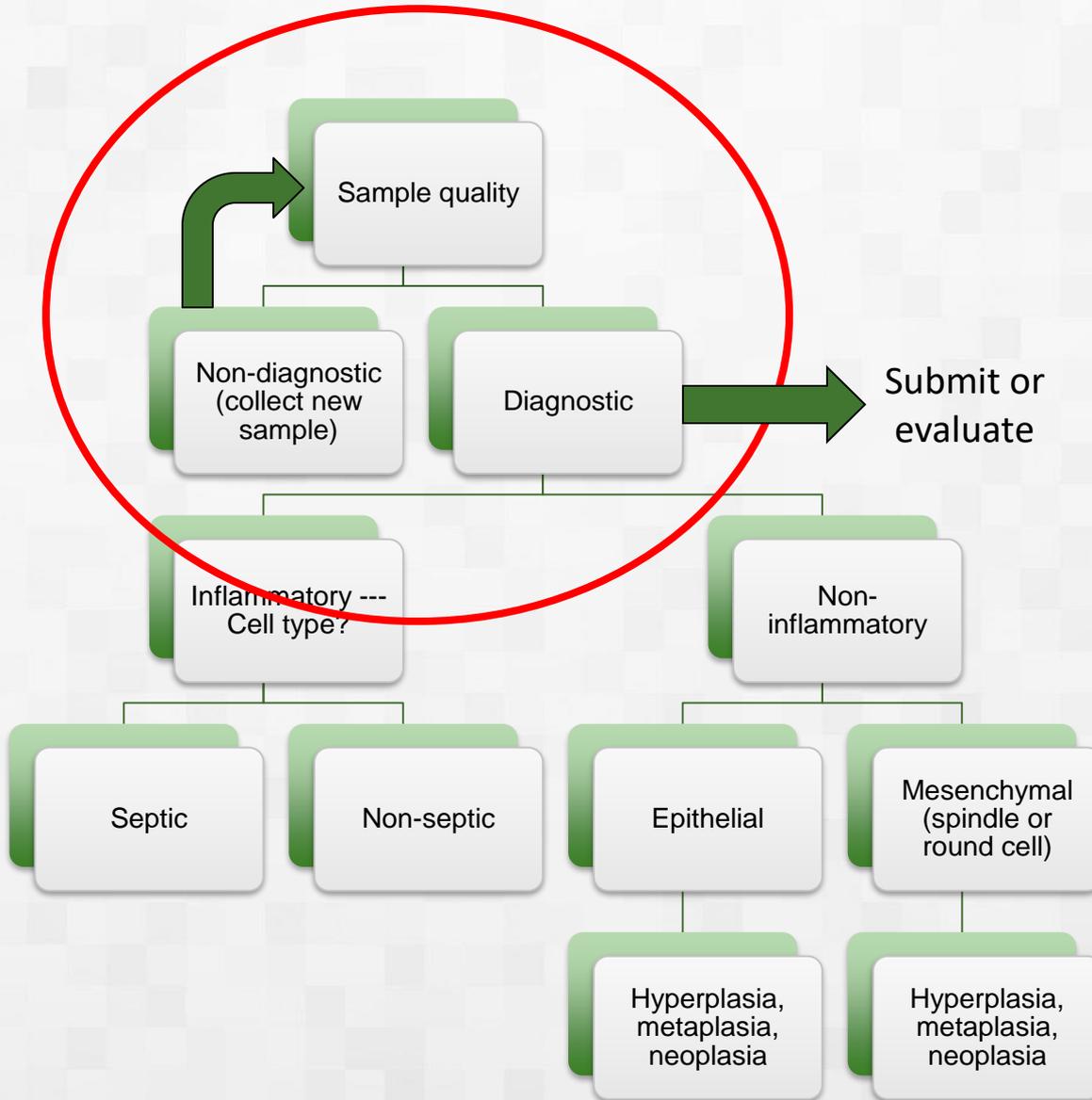


In a thin layer...



Without too much blood





In-house Cytology: The Short List

Melanoma

Mast Cell Tumor

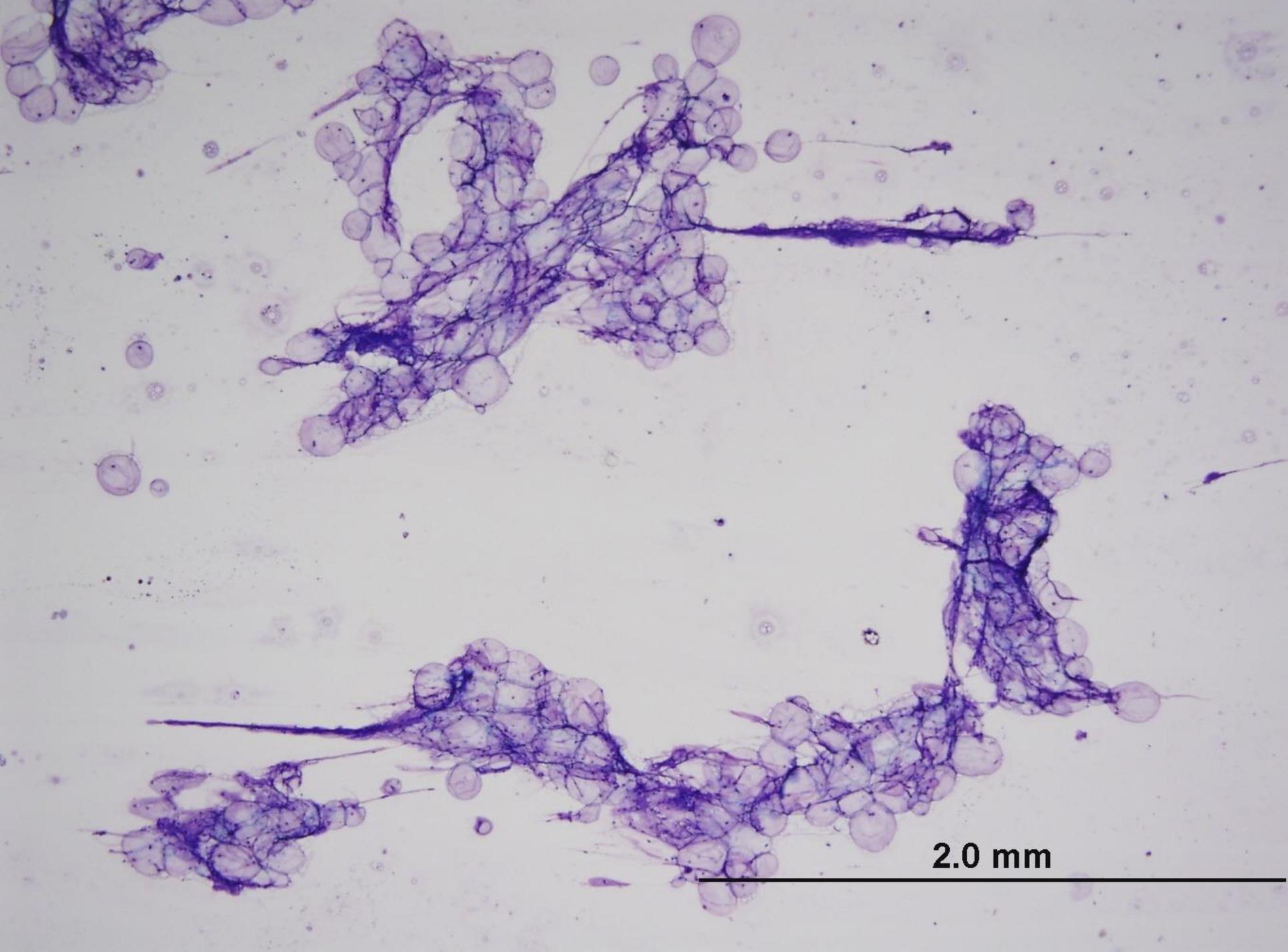
Lipoma

Sebaceous
adenoma

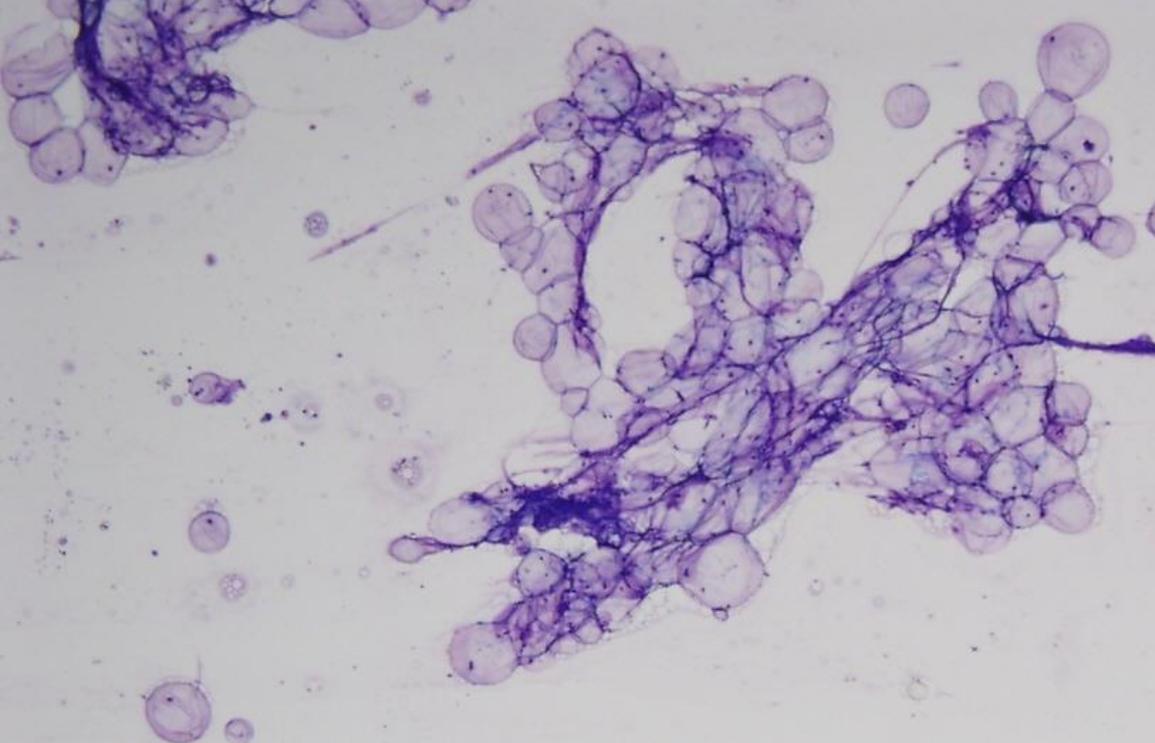
Follicular Cyst

Abscess

Septic Inflammation



2.0 mm



What is your diagnosis?

Abscess **A**

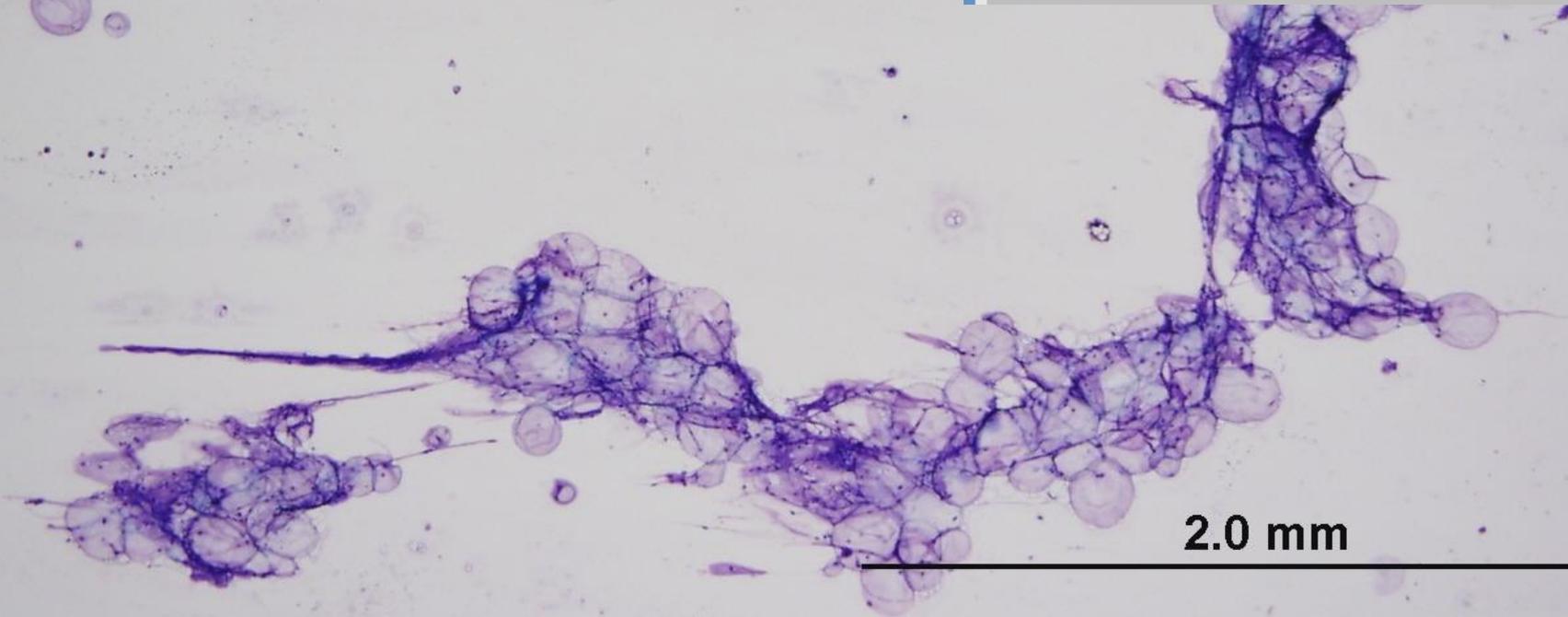
Lipoma **B**

Sebaceous
adenoma **C**

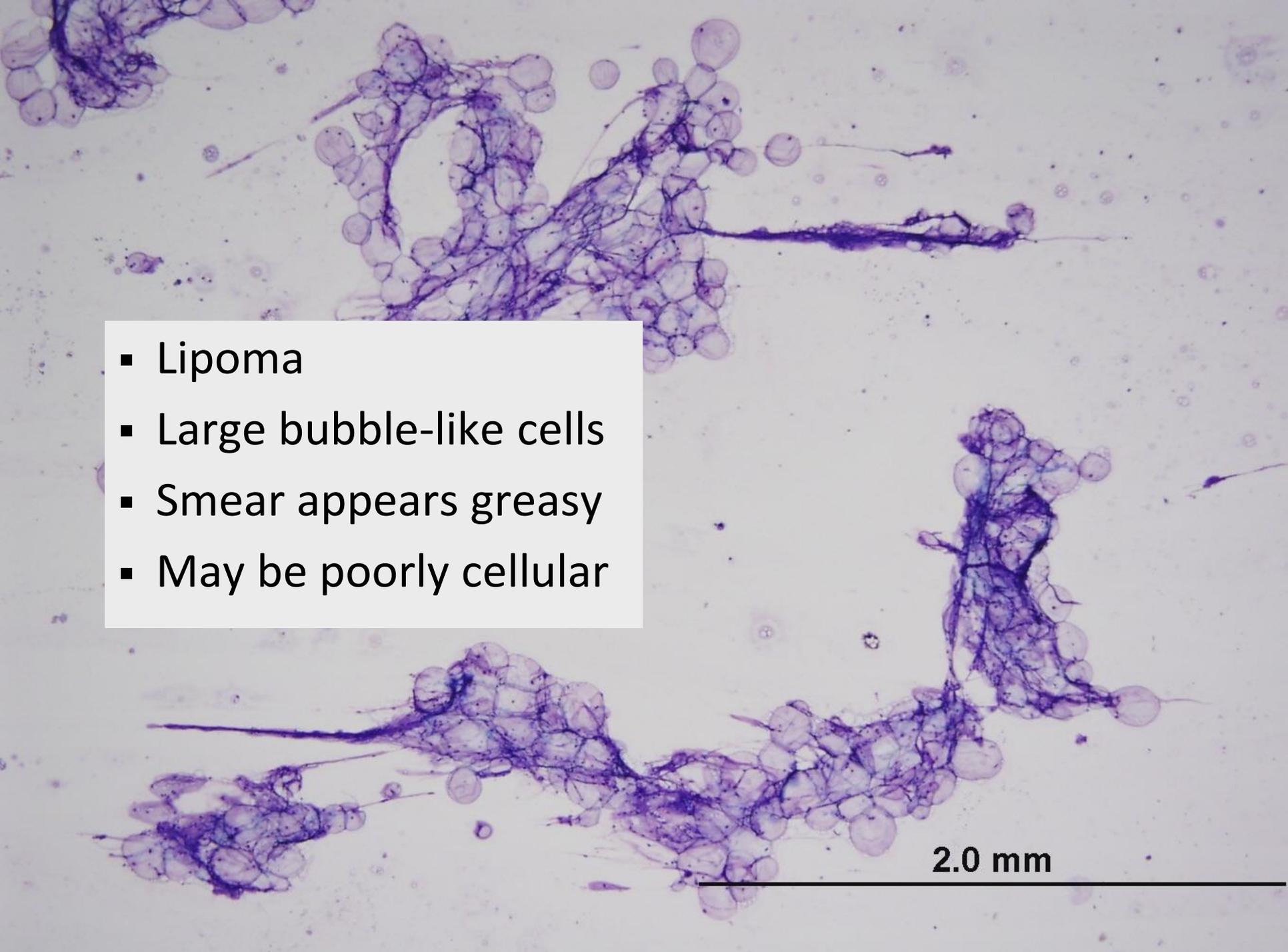
Follicular
cyst **D**

ill Everywhere

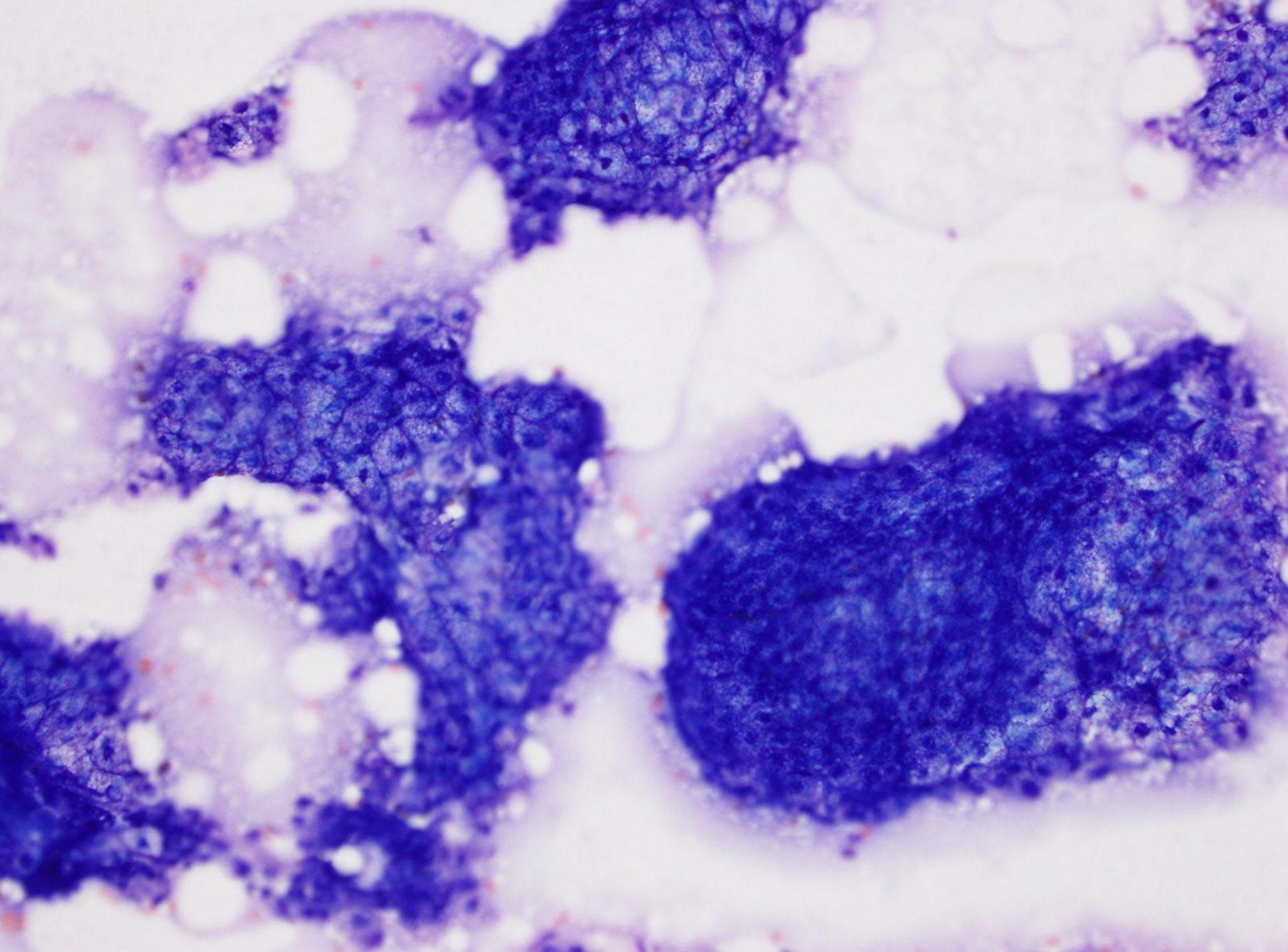
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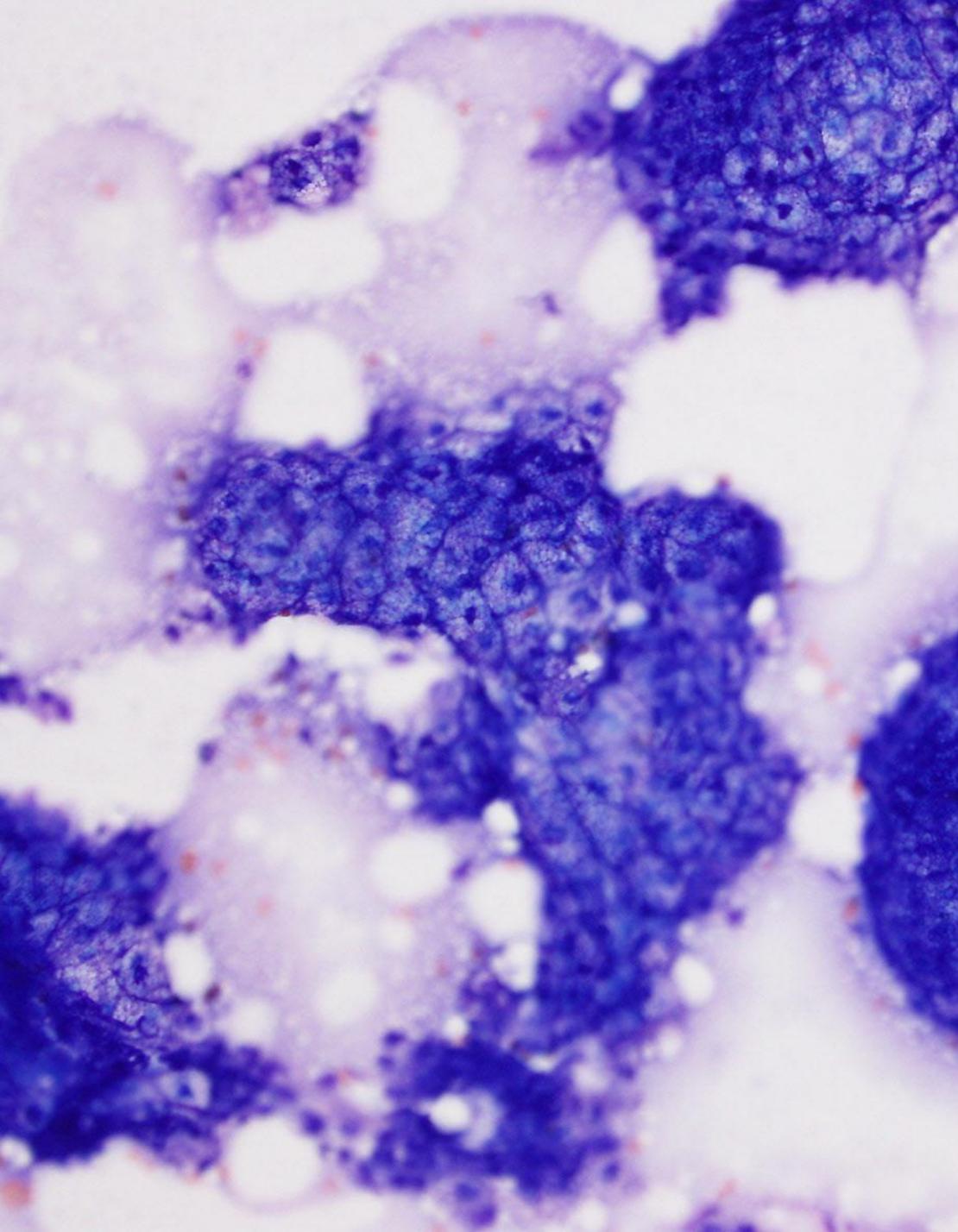


2.0 mm

- 
- A microscopic view of a lipoma smear. The image shows several clusters of large, clear, bubble-like cells with thin, purple-stained cell walls. The cells are arranged in irregular, somewhat circular or elongated groups. The background is a light, off-white color with some scattered purple-stained debris. A scale bar at the bottom right indicates 2.0 mm.
- Lipoma
 - Large bubble-like cells
 - Smear appears greasy
 - May be poorly cellular

2.0 mm





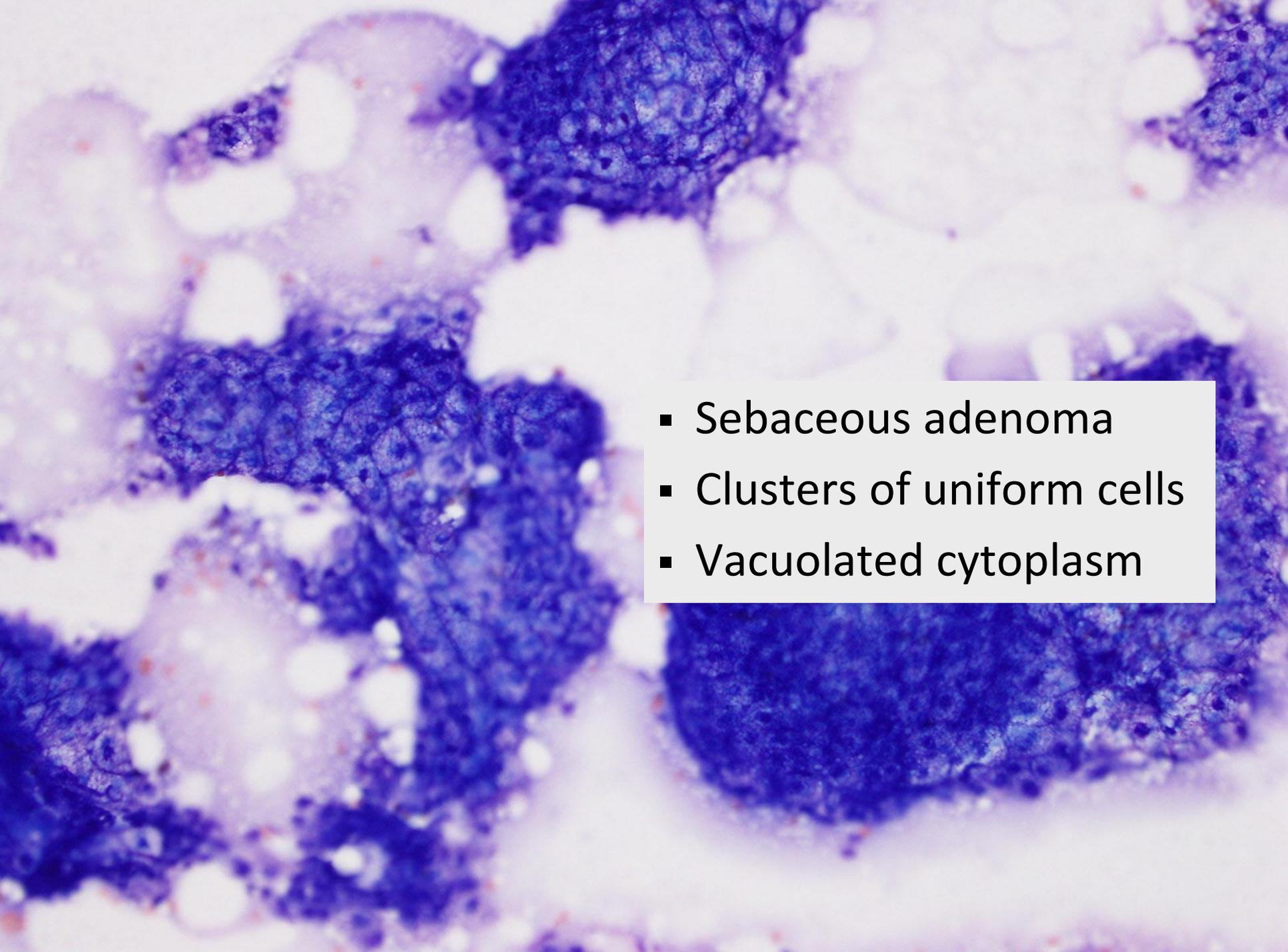
What is your diagnosis?

Lipoma

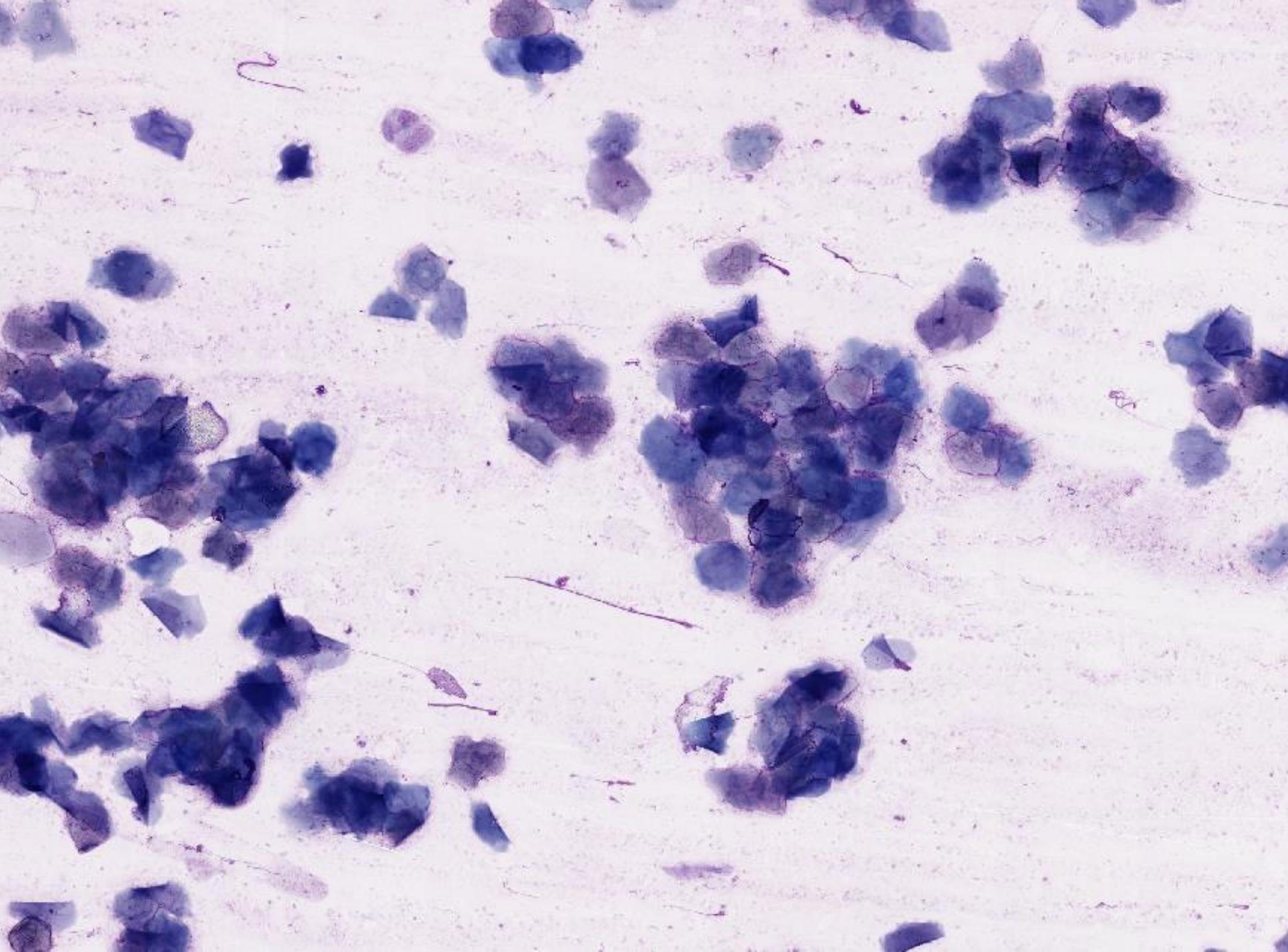
Sebaceous
adenoma

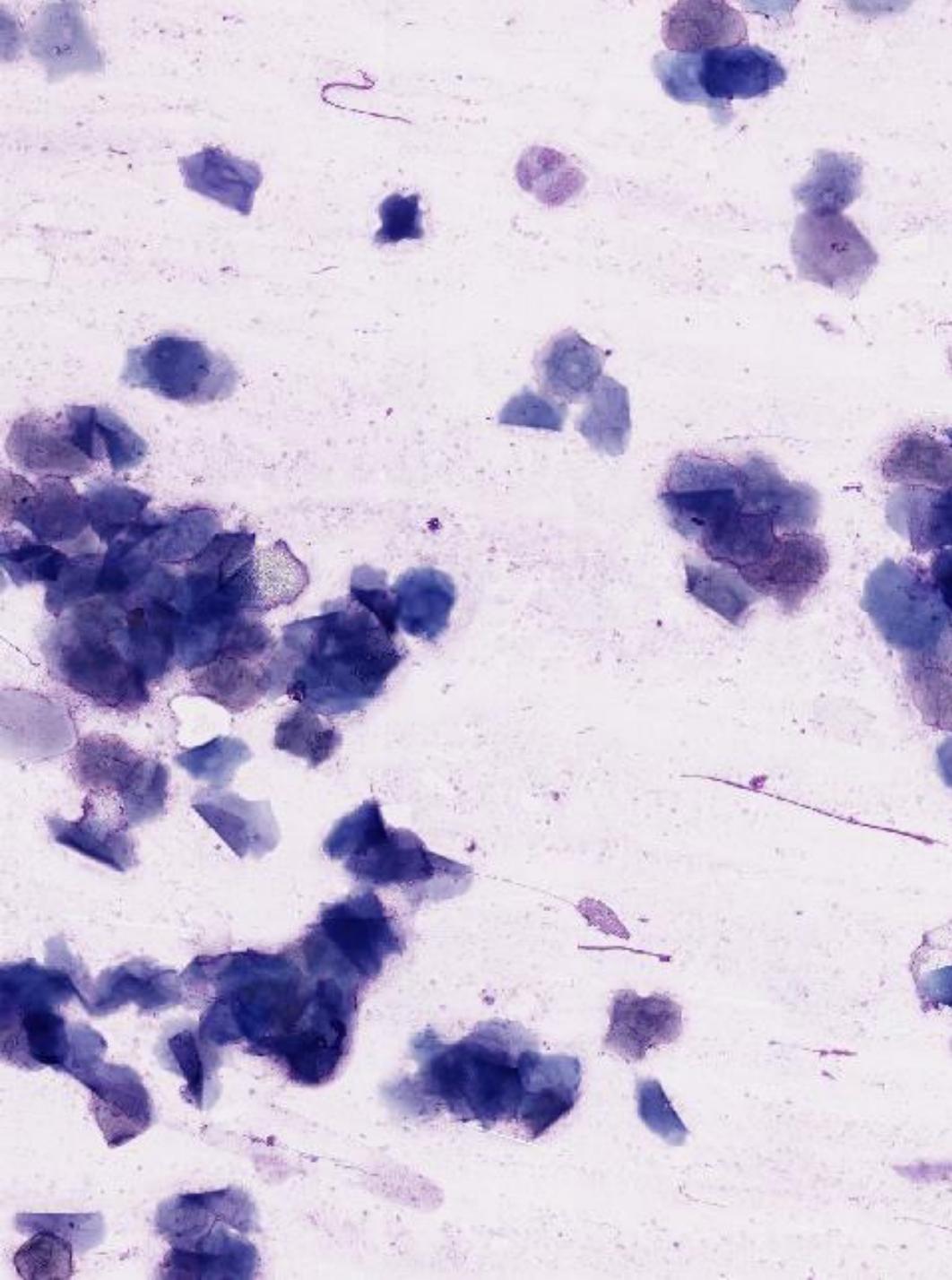
Mast cell
tumor

Follicular
cyst



- Sebaceous adenoma
- Clusters of uniform cells
- Vacuolated cytoplasm





What is your diagnosis?

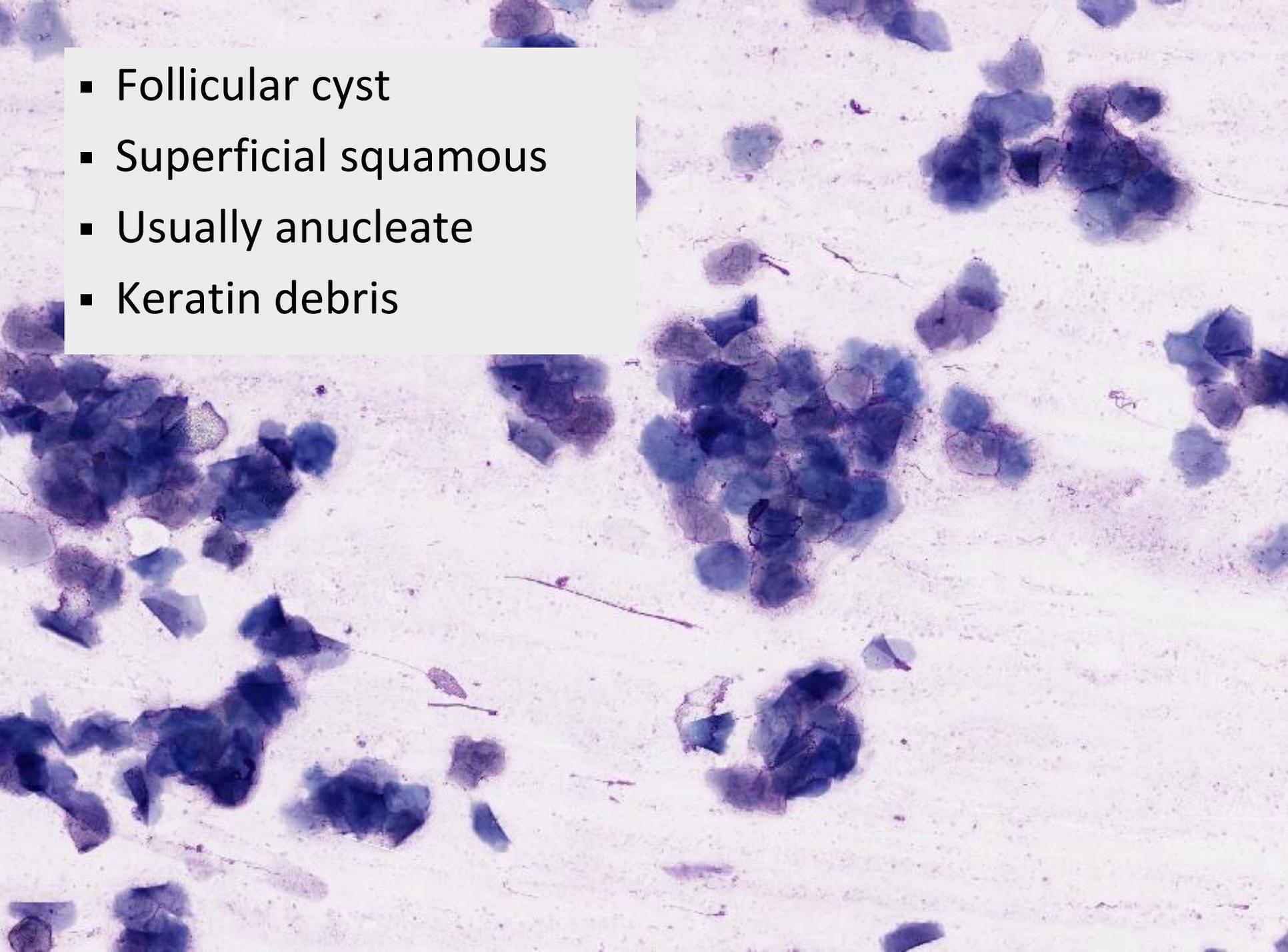
Septic
inflammation

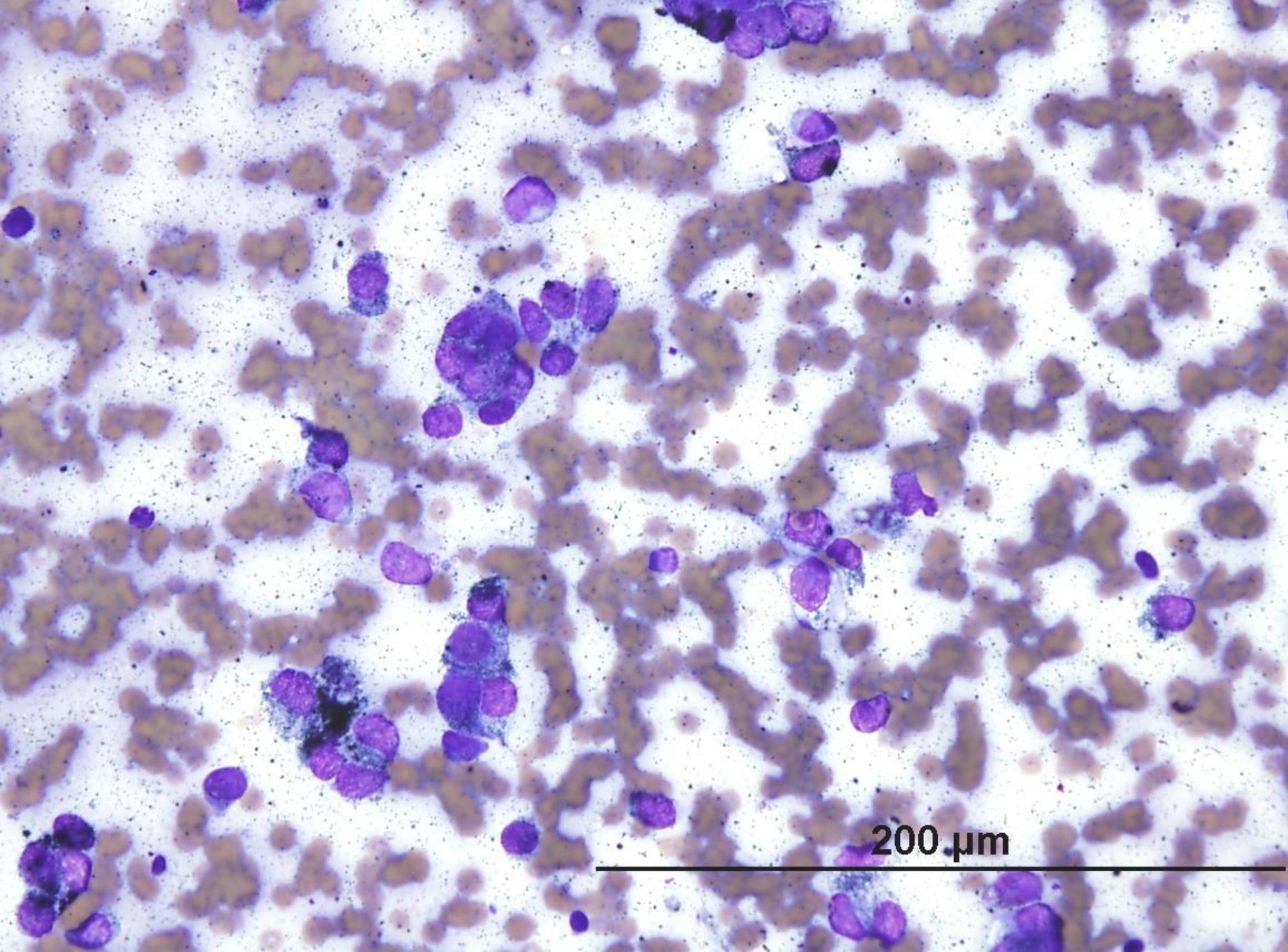
Mast cell
tumor

Follicular
cyst

Lipoma

- Follicular cyst
- Superficial squamous
- Usually anucleate
- Keratin debris





200 μm

What is your diagnosis?

Mast cell tumor

Septic
inflammation

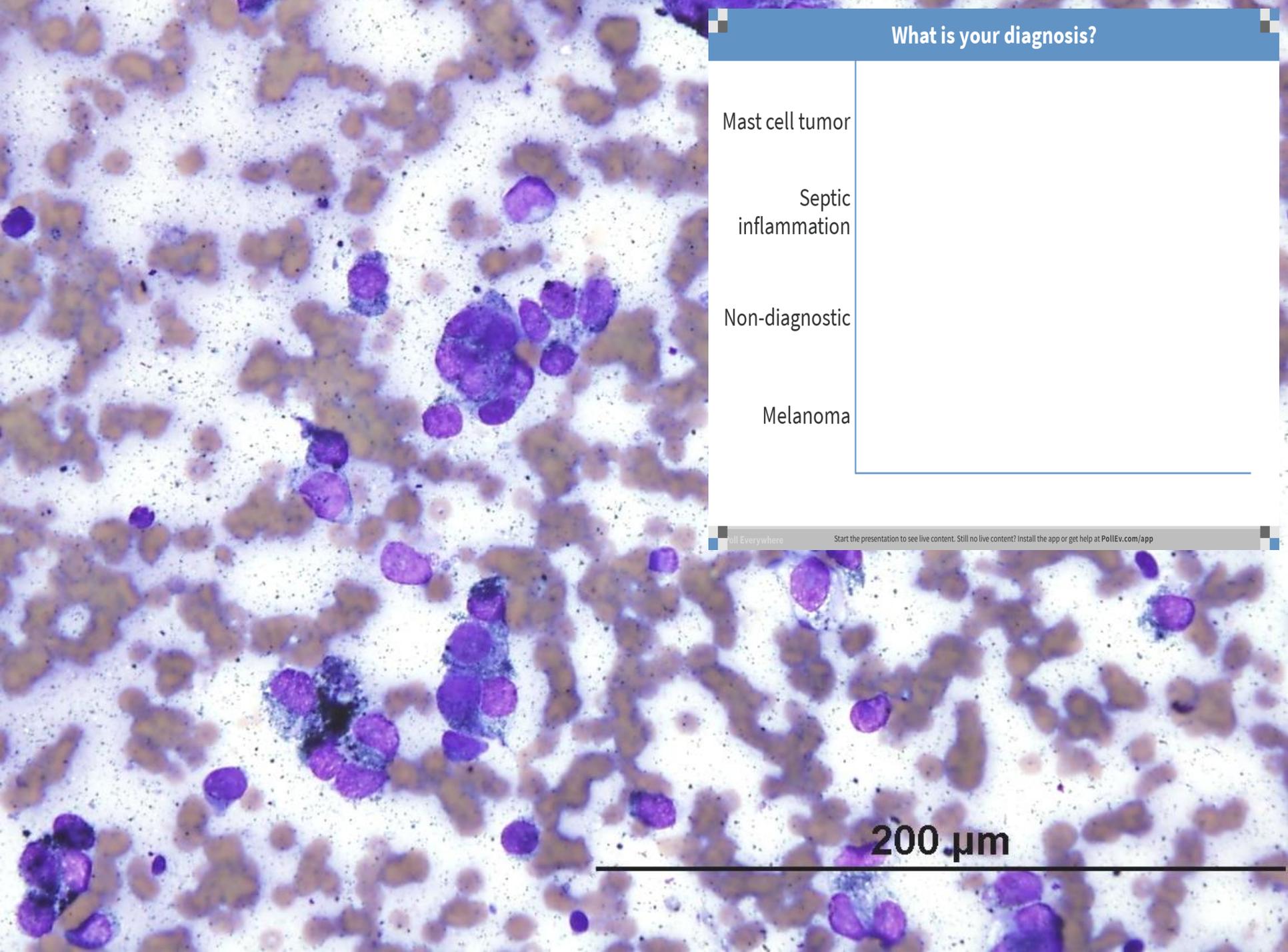
Non-diagnostic

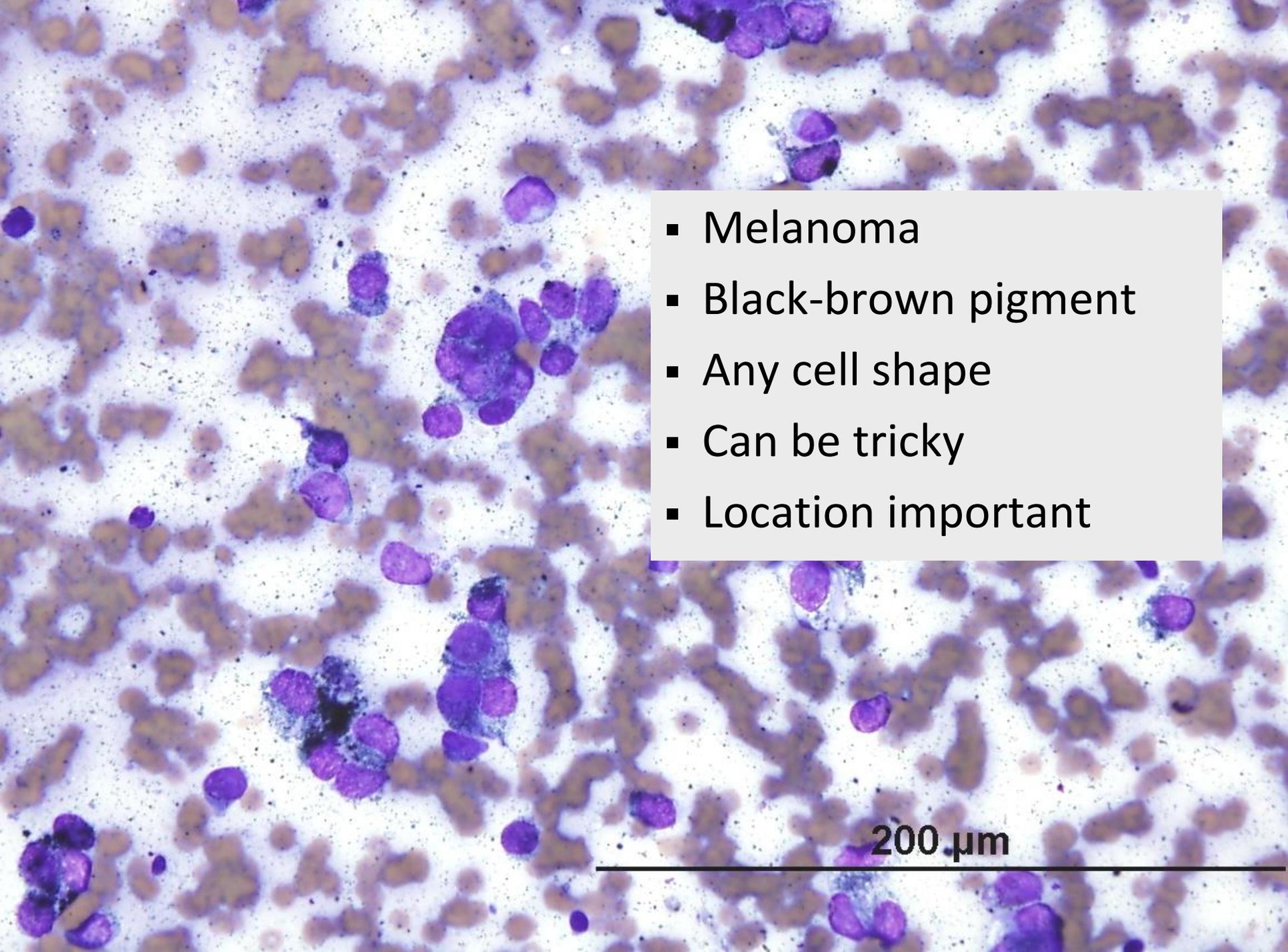
Melanoma

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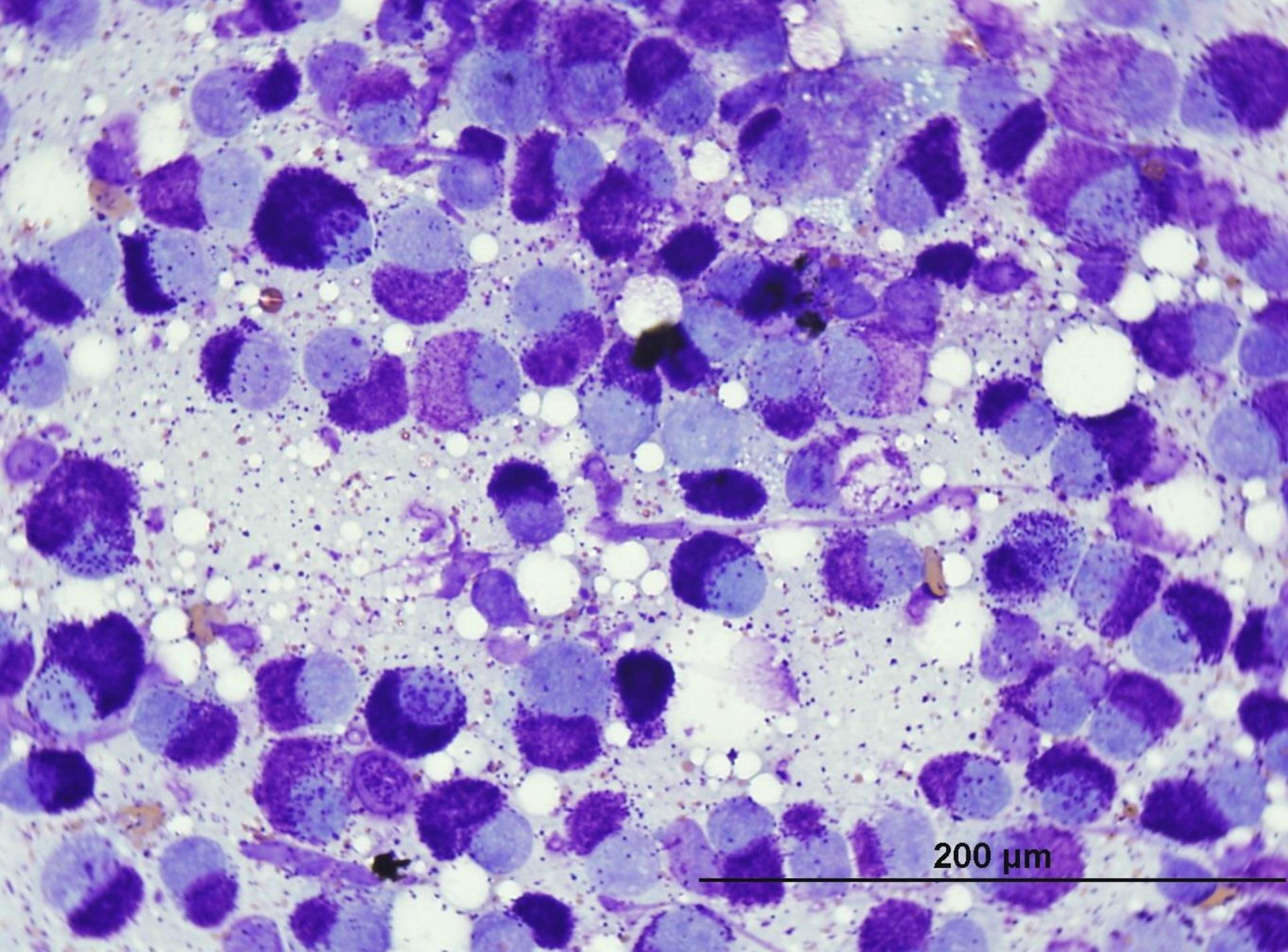
200 μ m



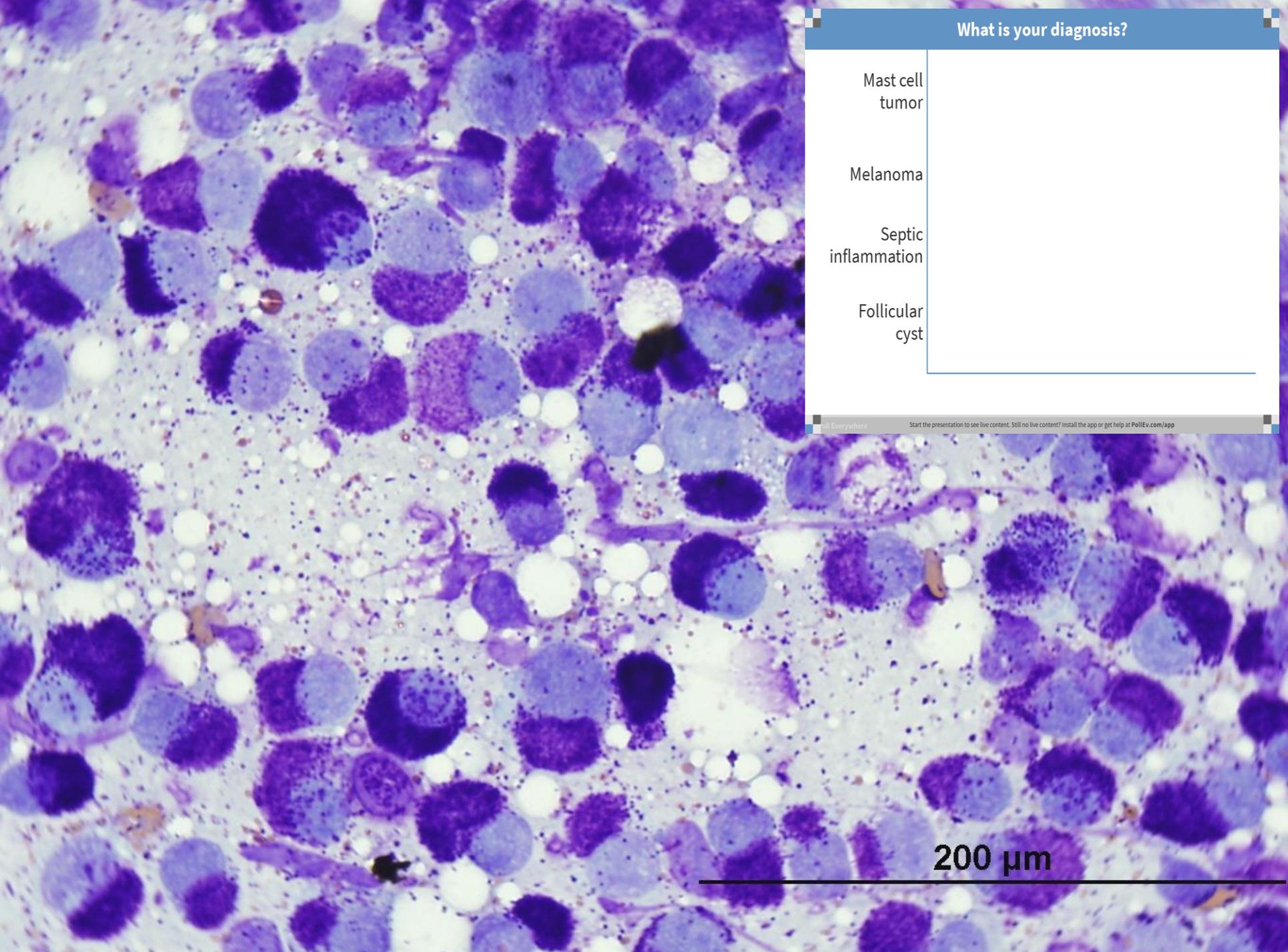


- Melanoma
- Black-brown pigment
- Any cell shape
- Can be tricky
- Location important

200 μm



200 μm



What is your diagnosis?

Mast cell tumor

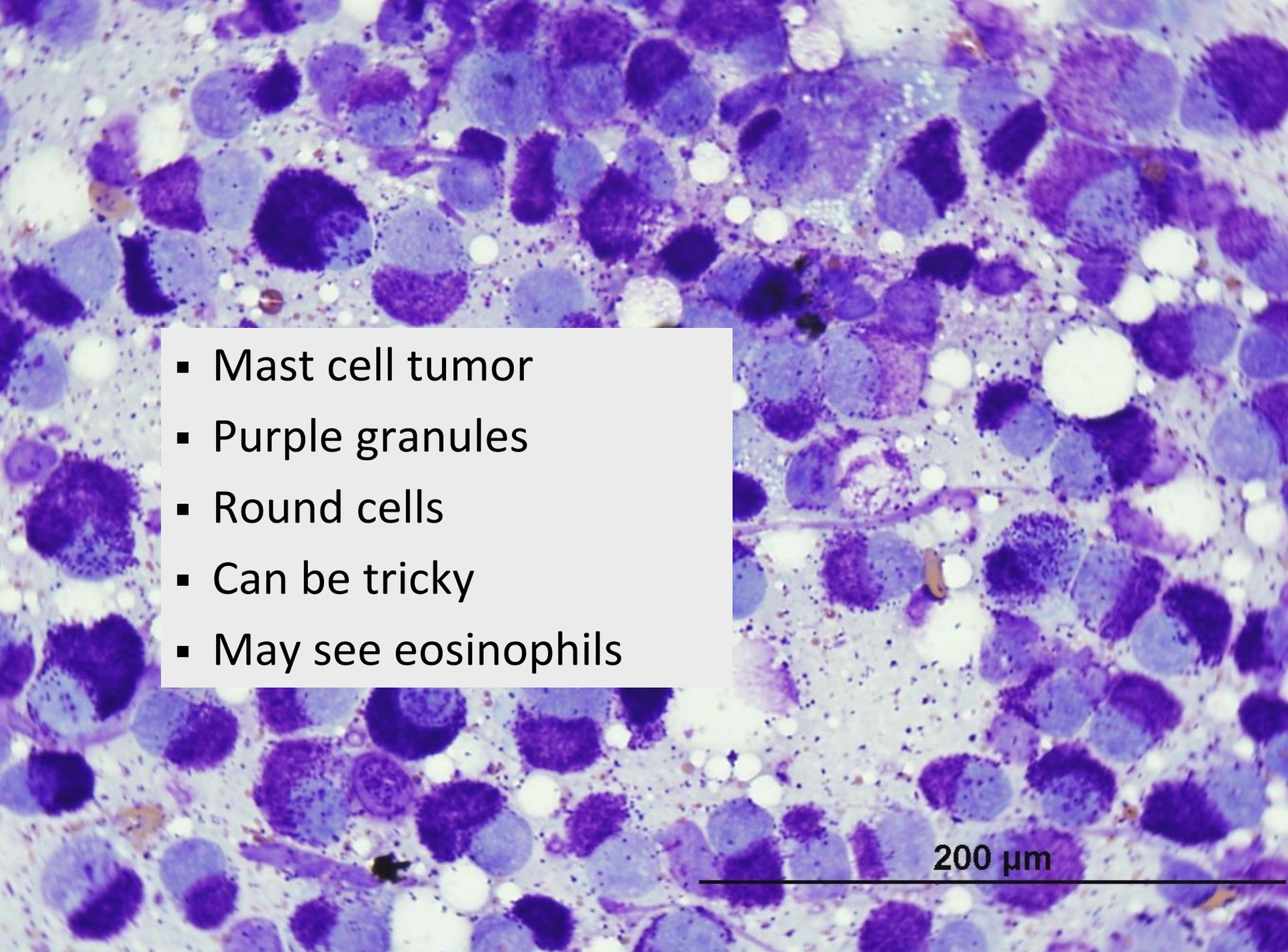
Melanoma

Septic inflammation

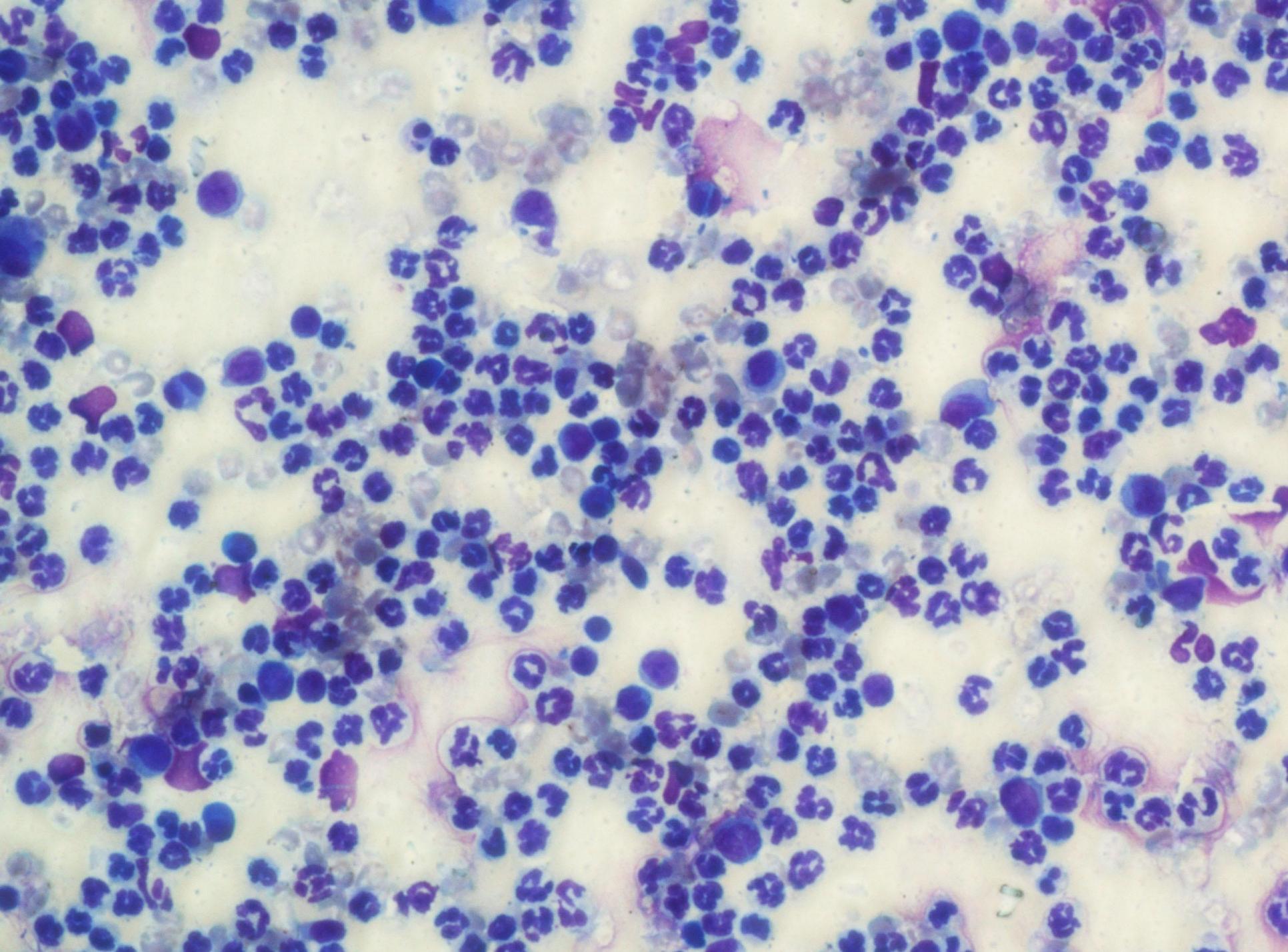
Follicular cyst

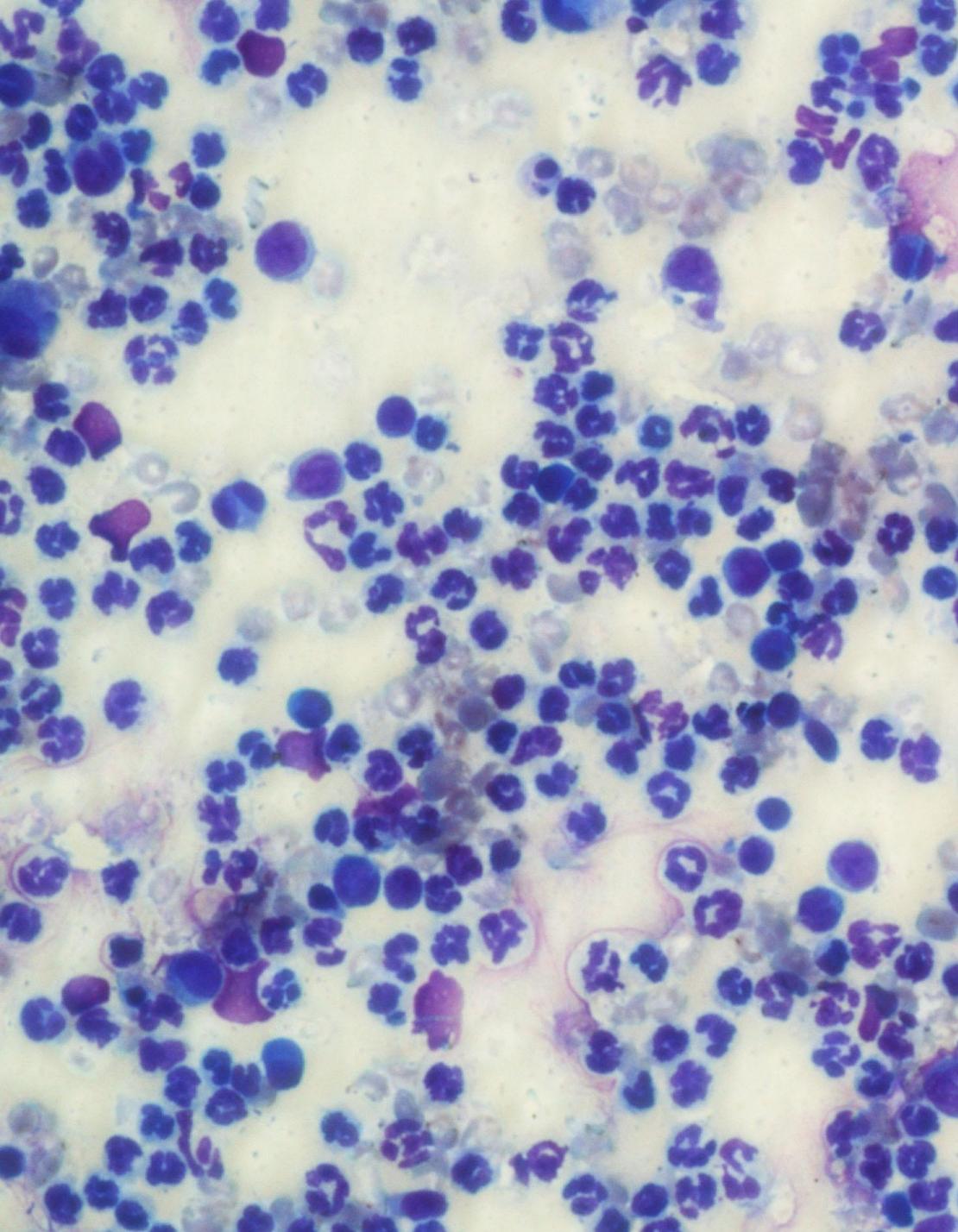
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200 μ m

- 
- A high-magnification photomicrograph of a mast cell tumor. The field is densely populated with round to oval cells. Each cell is characterized by a large, dark purple, granular cytoplasm that often obscures the nucleus. The nuclei are typically small and dark. The background is a light, pinkish-purple color, suggesting the presence of connective tissue or other cellular components. The overall appearance is that of a highly cellular neoplasm.
- Mast cell tumor
 - Purple granules
 - Round cells
 - Can be tricky
 - May see eosinophils

200 μm





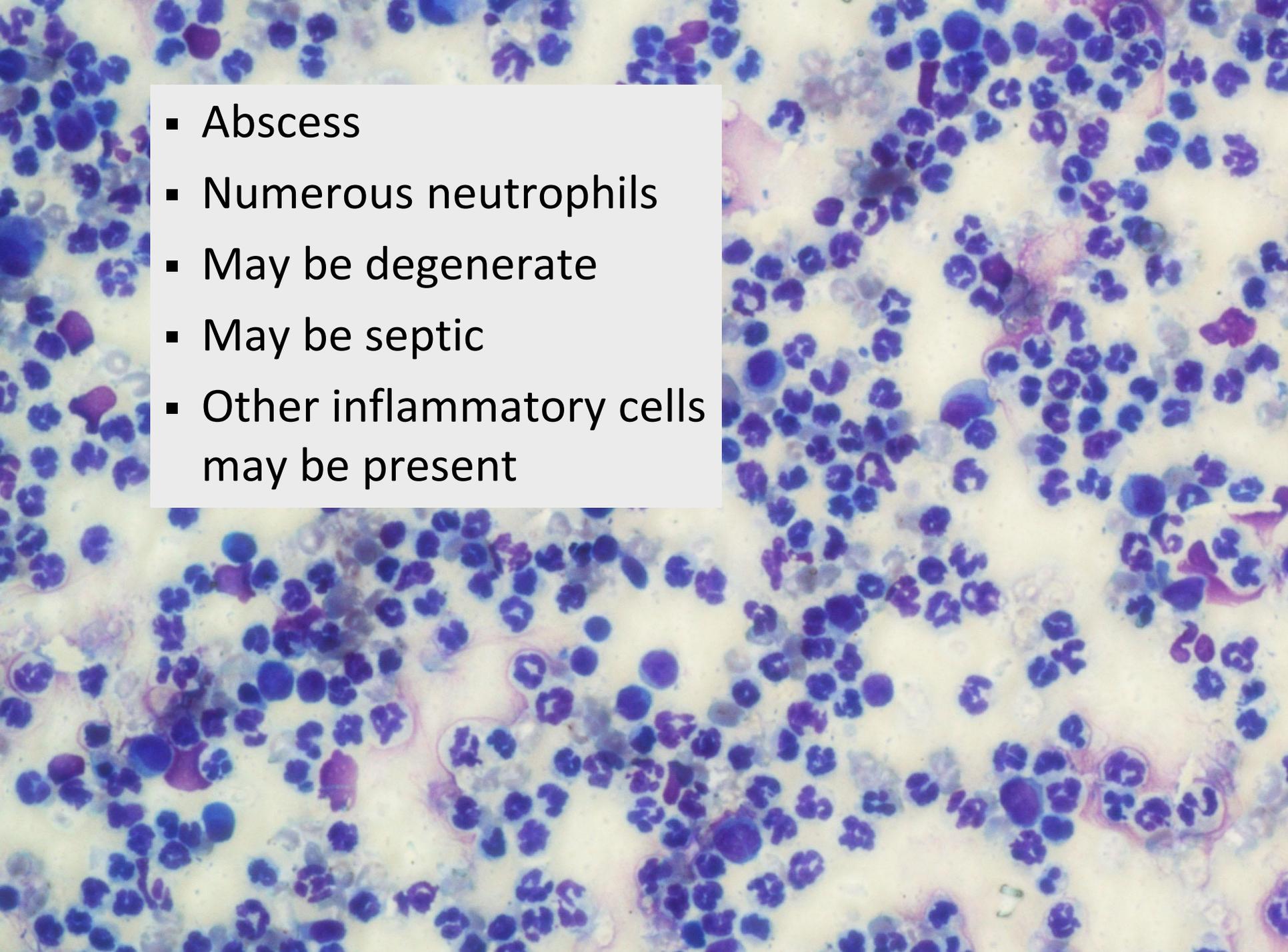
What is your diagnosis?

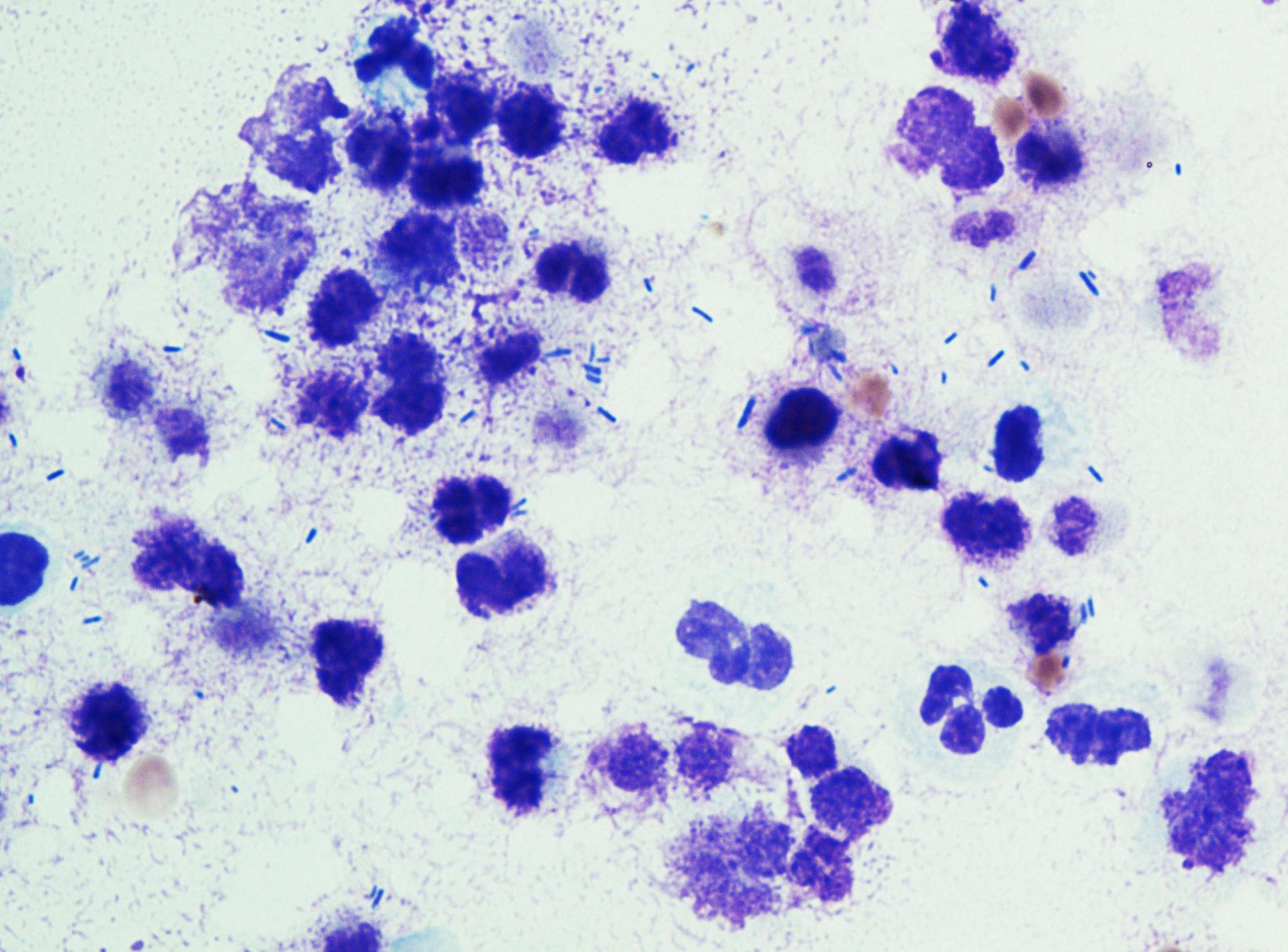
Septic inflammation

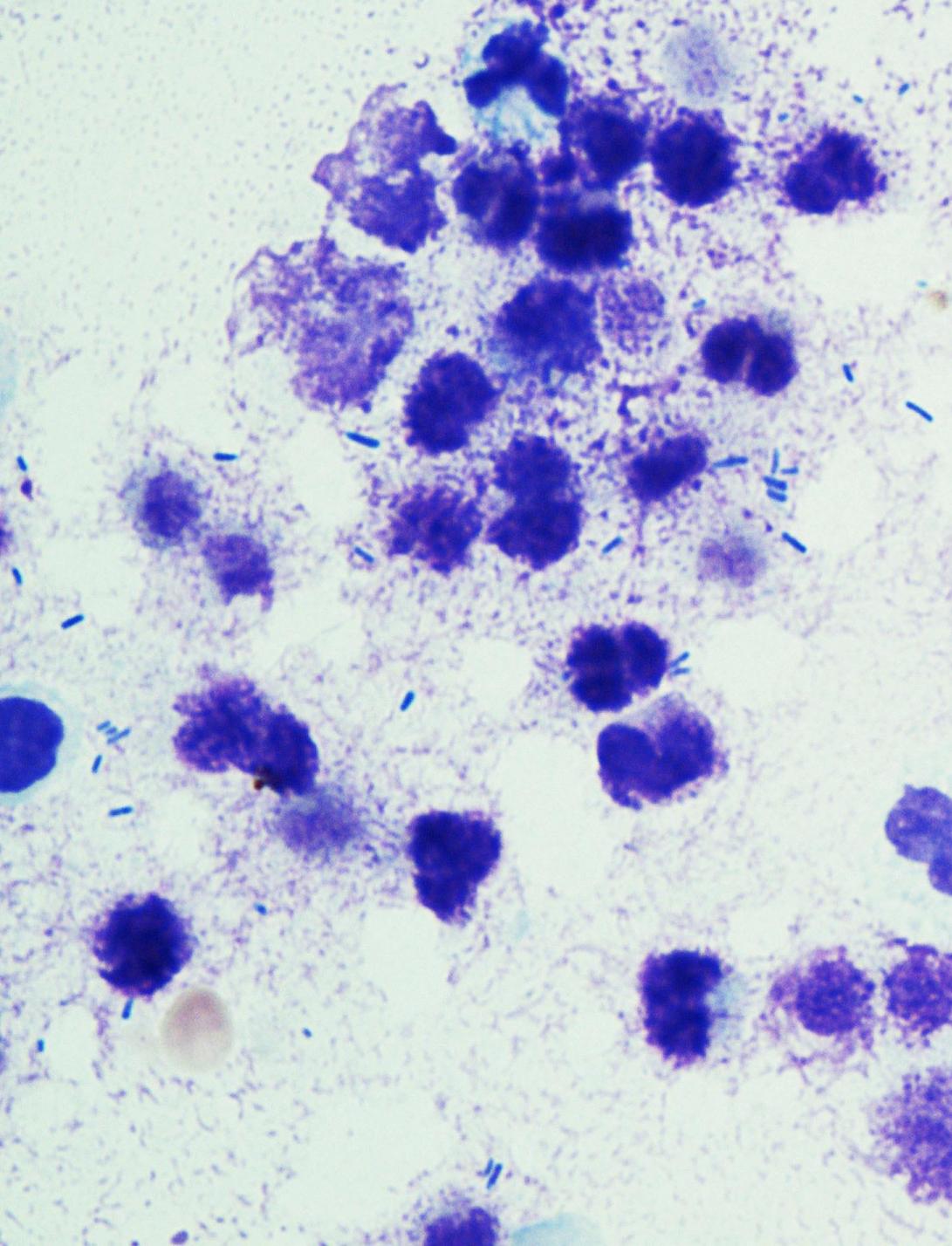
Follicular cyst

Abscess

Melanoma

- 
- A microscopic image of an abscess, showing a dense collection of neutrophils (white blood cells) with characteristic multi-lobed nuclei. The cells are stained with hematoxylin and eosin (H&E), resulting in purple nuclei and pink cytoplasm/extracellular matrix. The neutrophils are arranged in clusters, and some show signs of degeneration, such as pyknotic nuclei and cytoplasmic vacuolization. The background is a pale, eosinophilic (pink) matrix, likely fibrin or pus.
- Abscess
 - Numerous neutrophils
 - May be degenerate
 - May be septic
 - Other inflammatory cells may be present





What is your diagnosis?

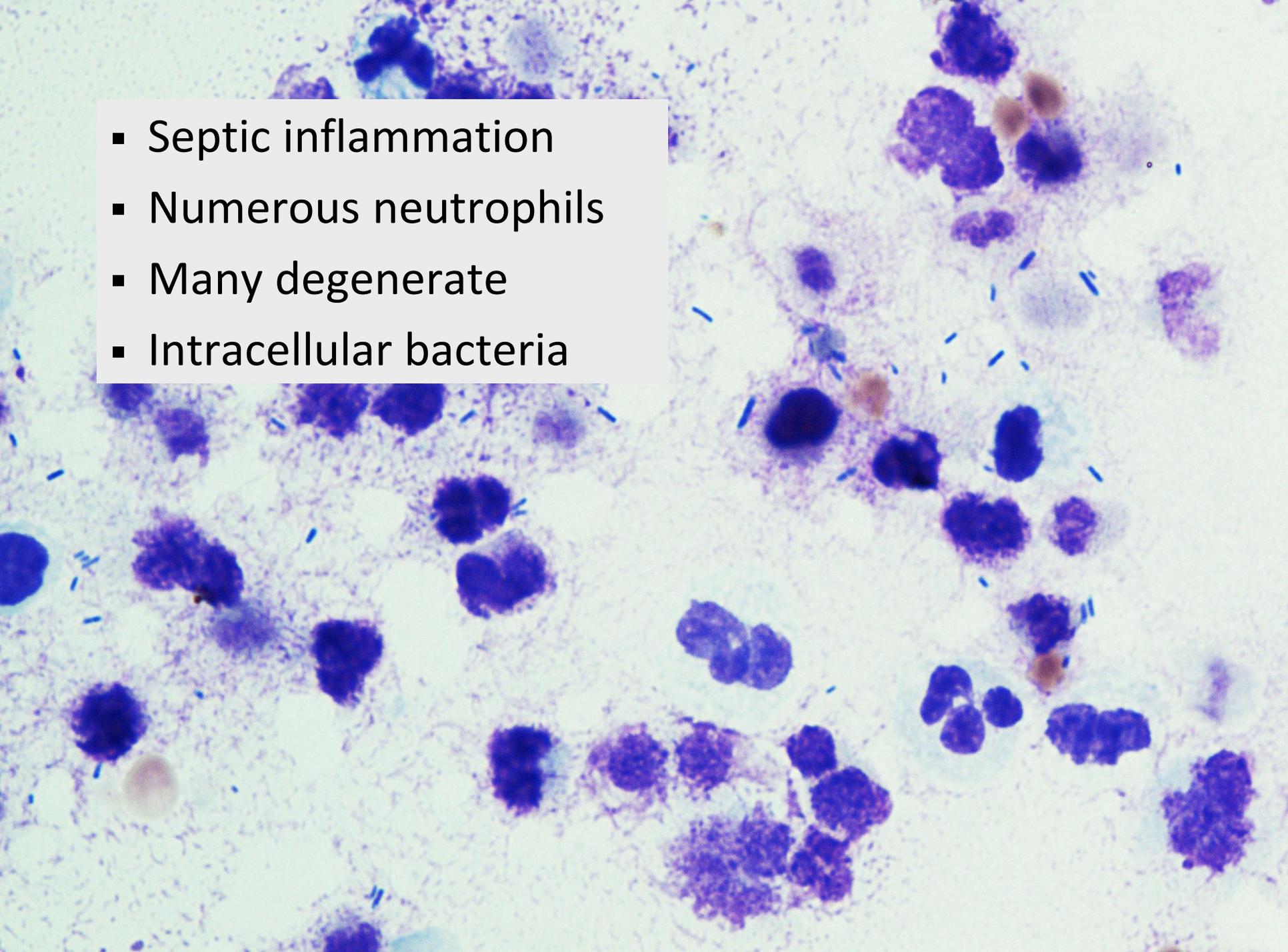
Septic
inflammation

Follicular
cyst

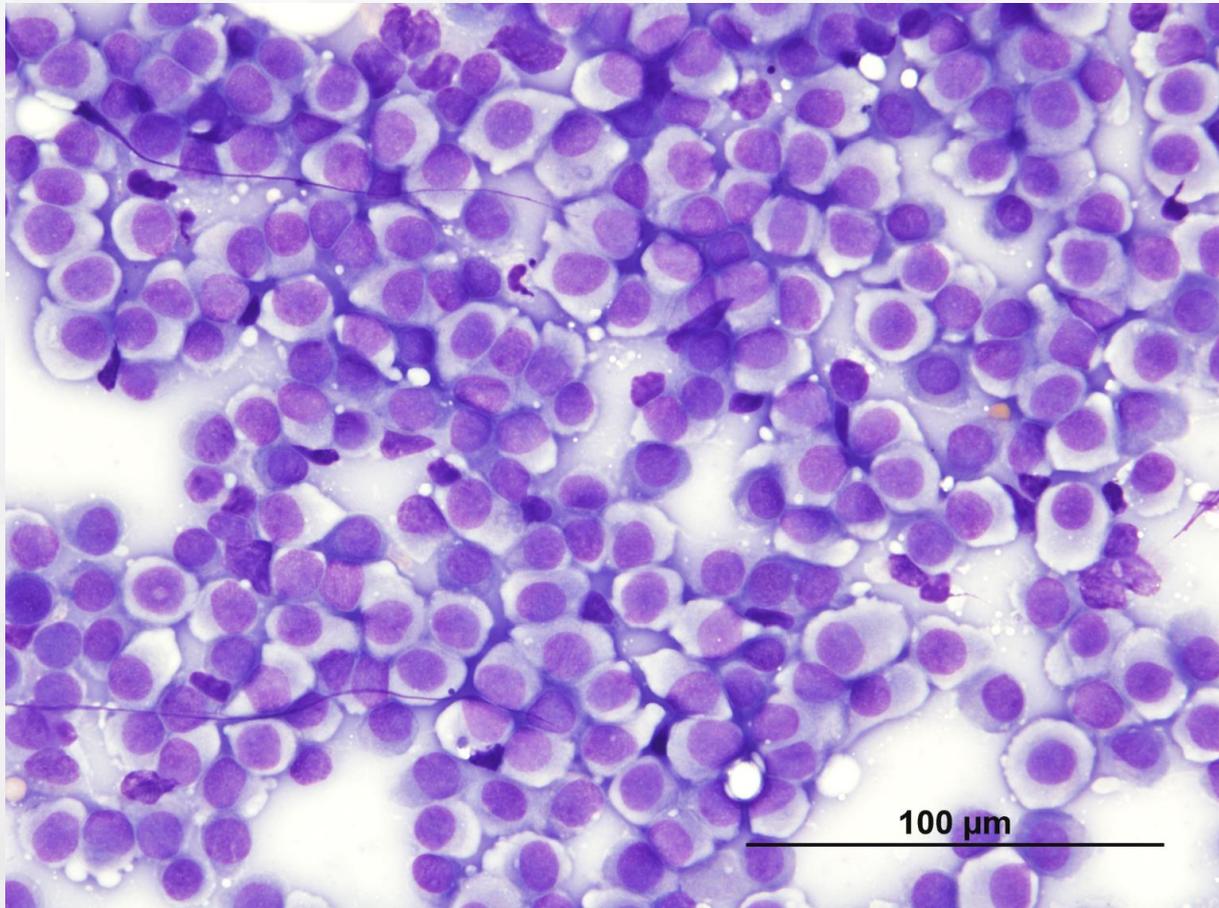
Abscess

Melanoma

- Septic inflammation
- Numerous neutrophils
- Many degenerate
- Intracellular bacteria

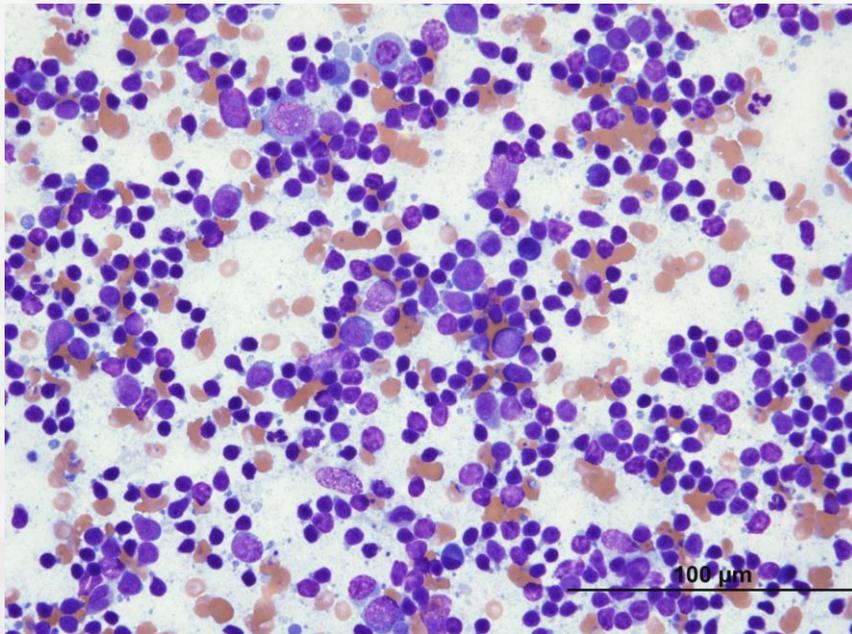


Special Considerations

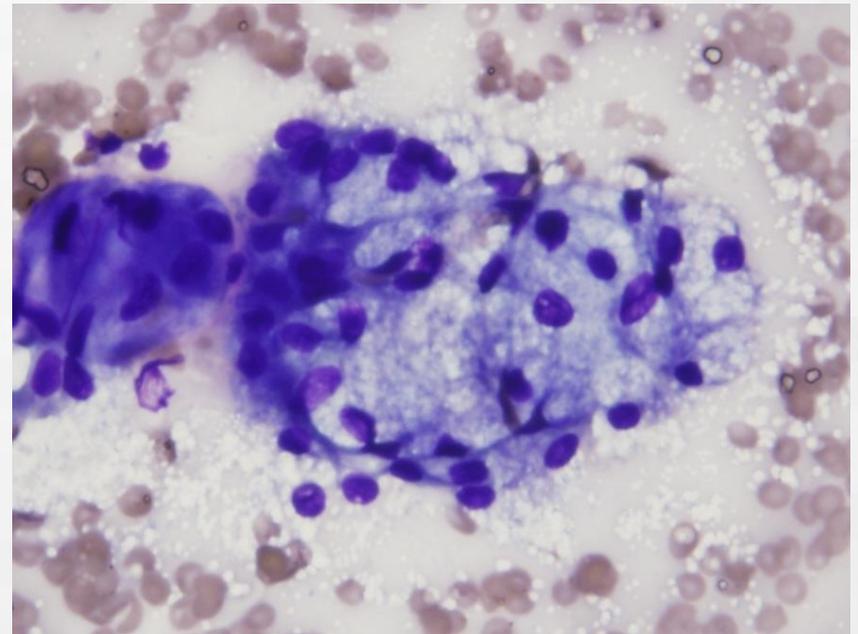


Sub-mandibular mass

Lymph node

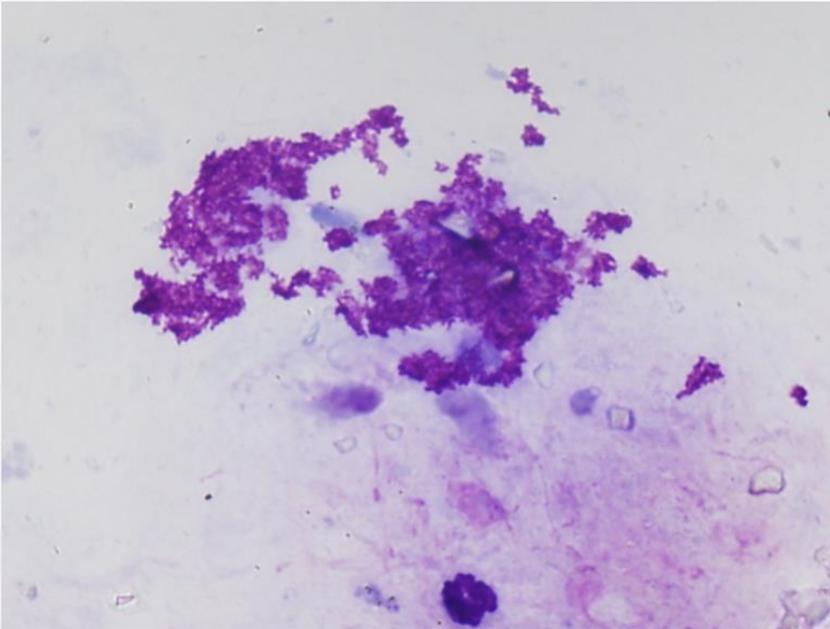


Salivary gland

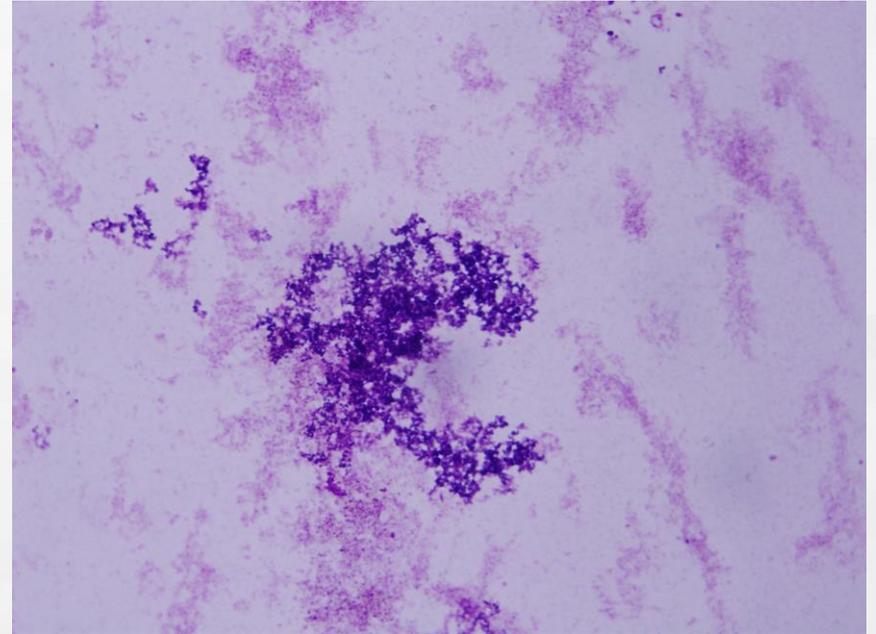


Artifacts

US gel



Stain precipitate



Questions?

Stay tuned for hematology...



Anna Lee Larimore

Part 2: Hematology

Red cells

White cells

Platelets

Part 2: Hematology

- Ask the audience:

 Respond at **PollEv.com/nicolefernan414**

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Do you have an in-house hematology analyzer?

Yes, I run all my
CBCs on it

Yes, I run presurgical
and STAT CBCs on it

No, I send CBCs to a
diagnostic lab

No

Do you routinely examine blood smears from in-house CBCs?

Yes, on almost
all patients

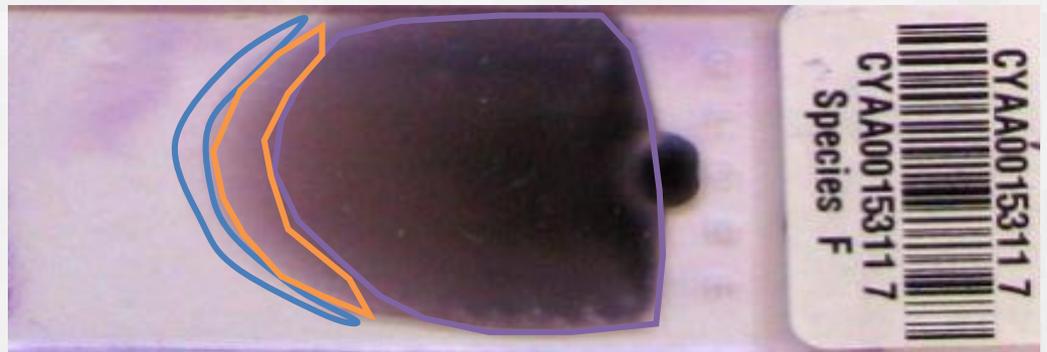
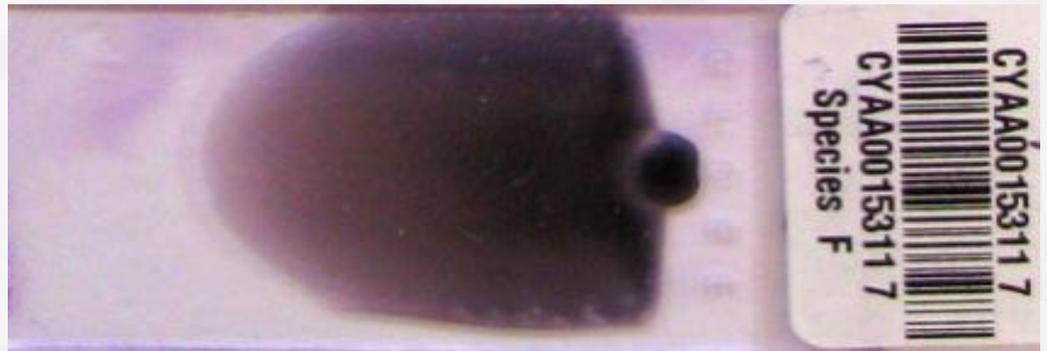
Yes, if there is
something weird
on the CBC

No

Hematology

- Your analyzer can't tell you everything
- RBC morphology
 - Regeneration
 - Causes of anemia
- WBC morphology
 - Left shift and toxic change
 - Leukemia
- Platelet clumping

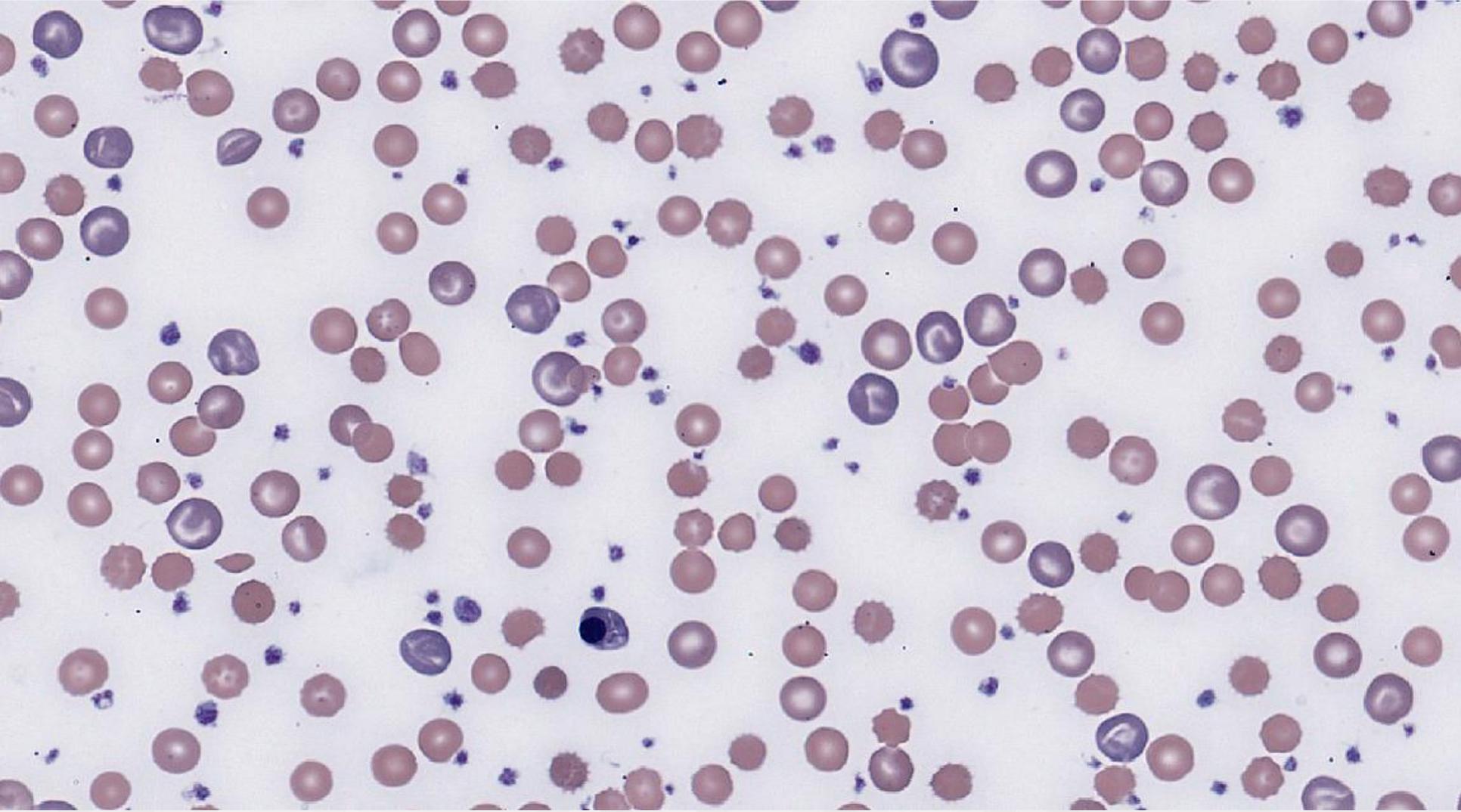
The Monolayer



Feathered Edge

Monolayer

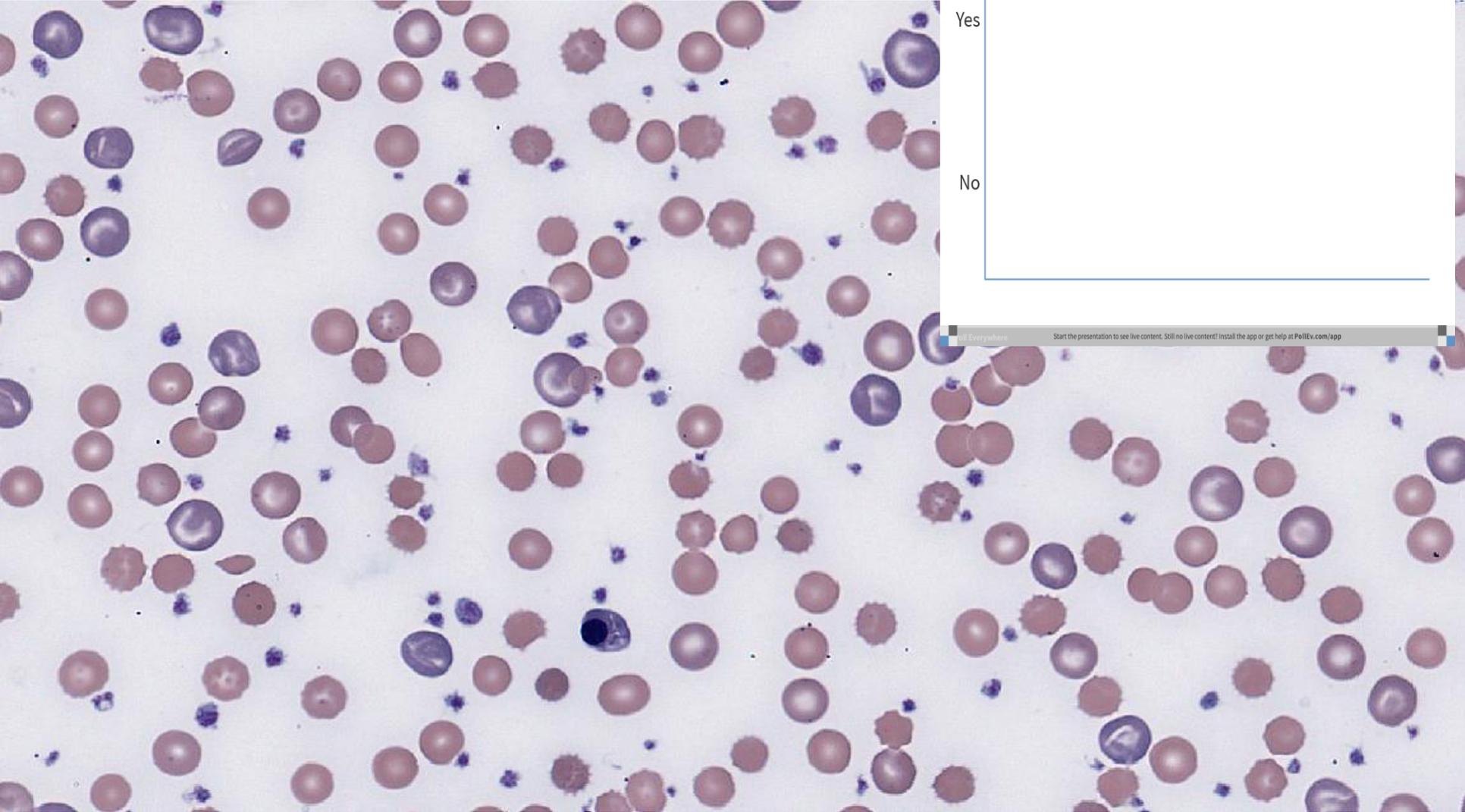
Body



Is this anemia regenerative?

Yes

No



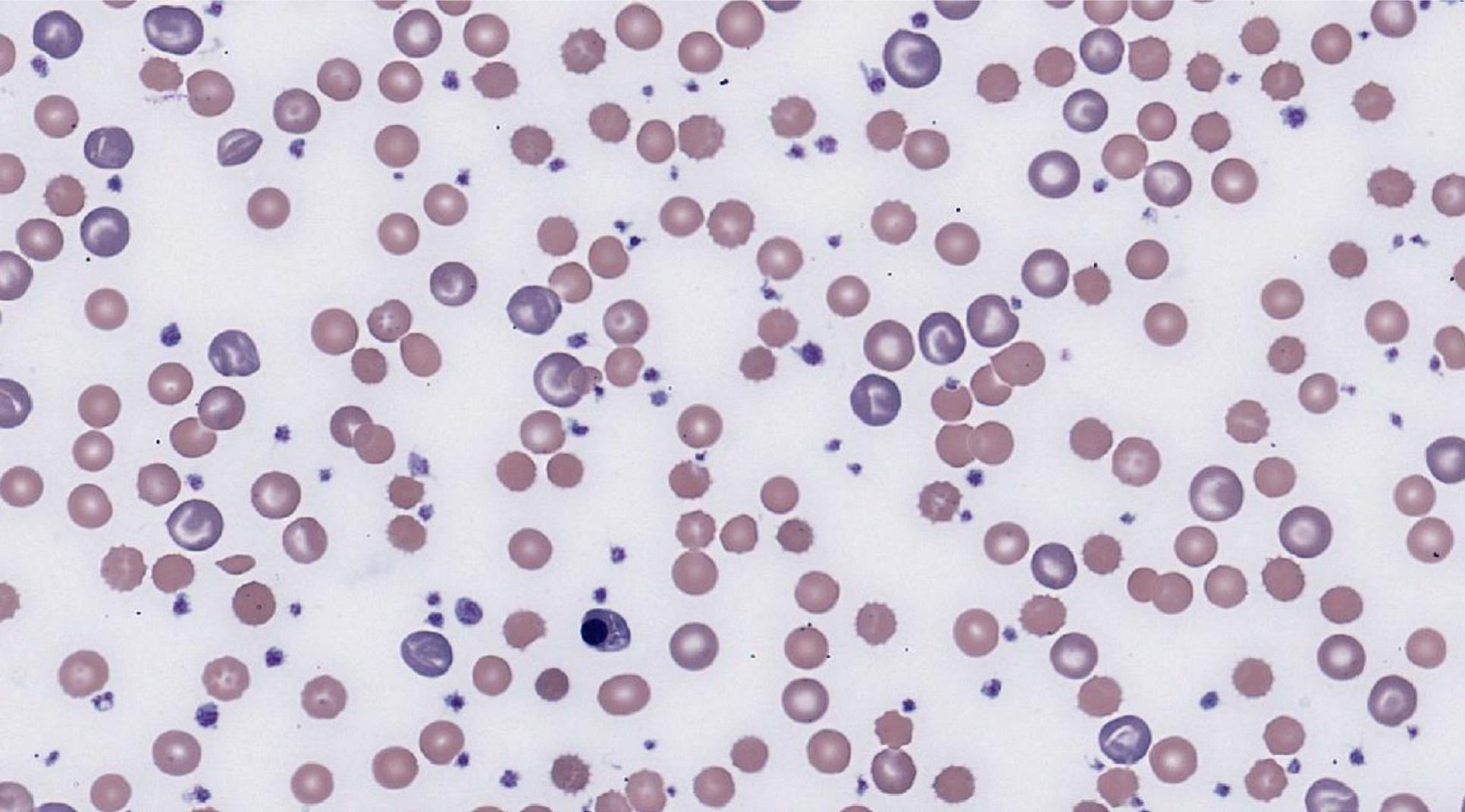
What is the evidence in support of regeneration?

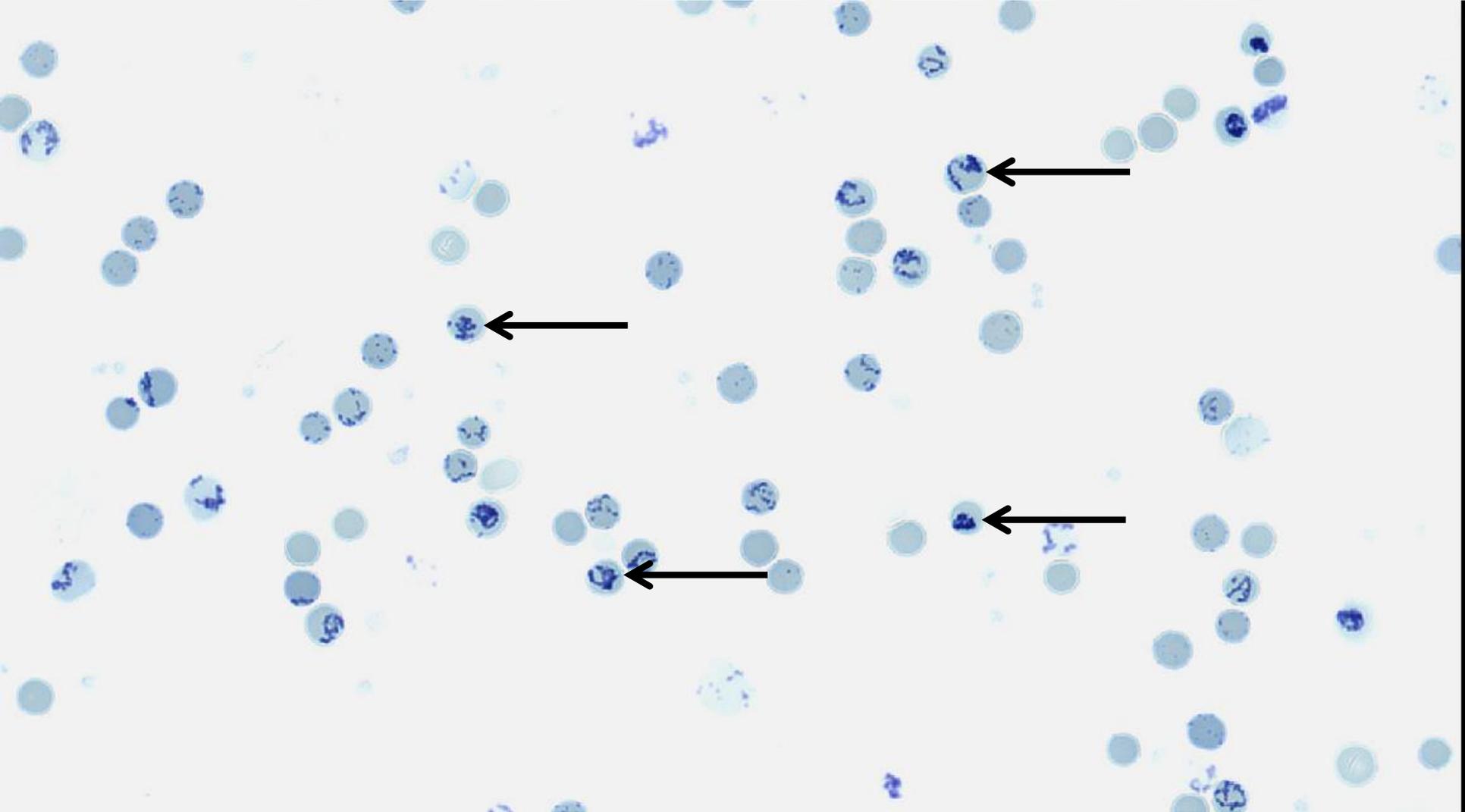


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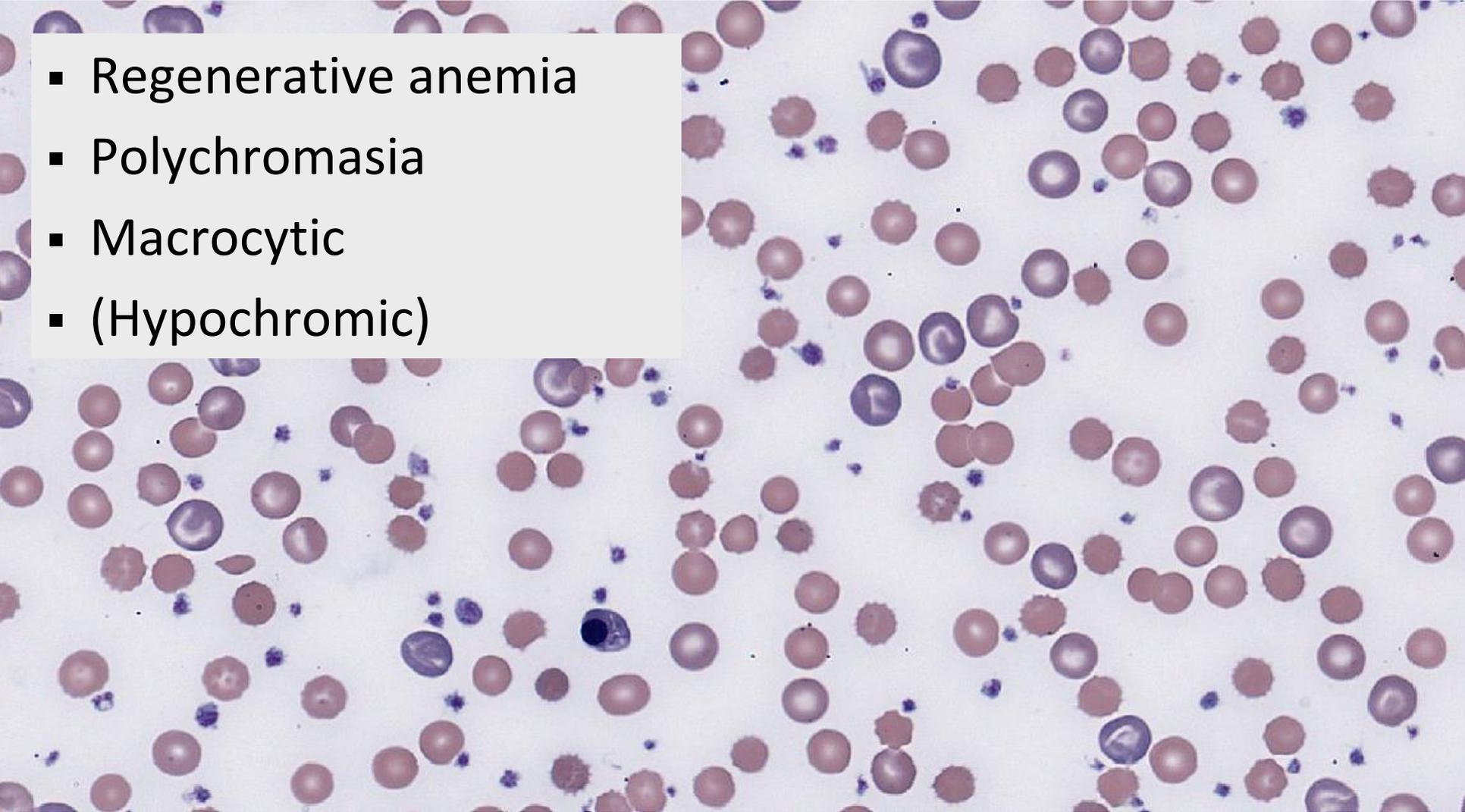


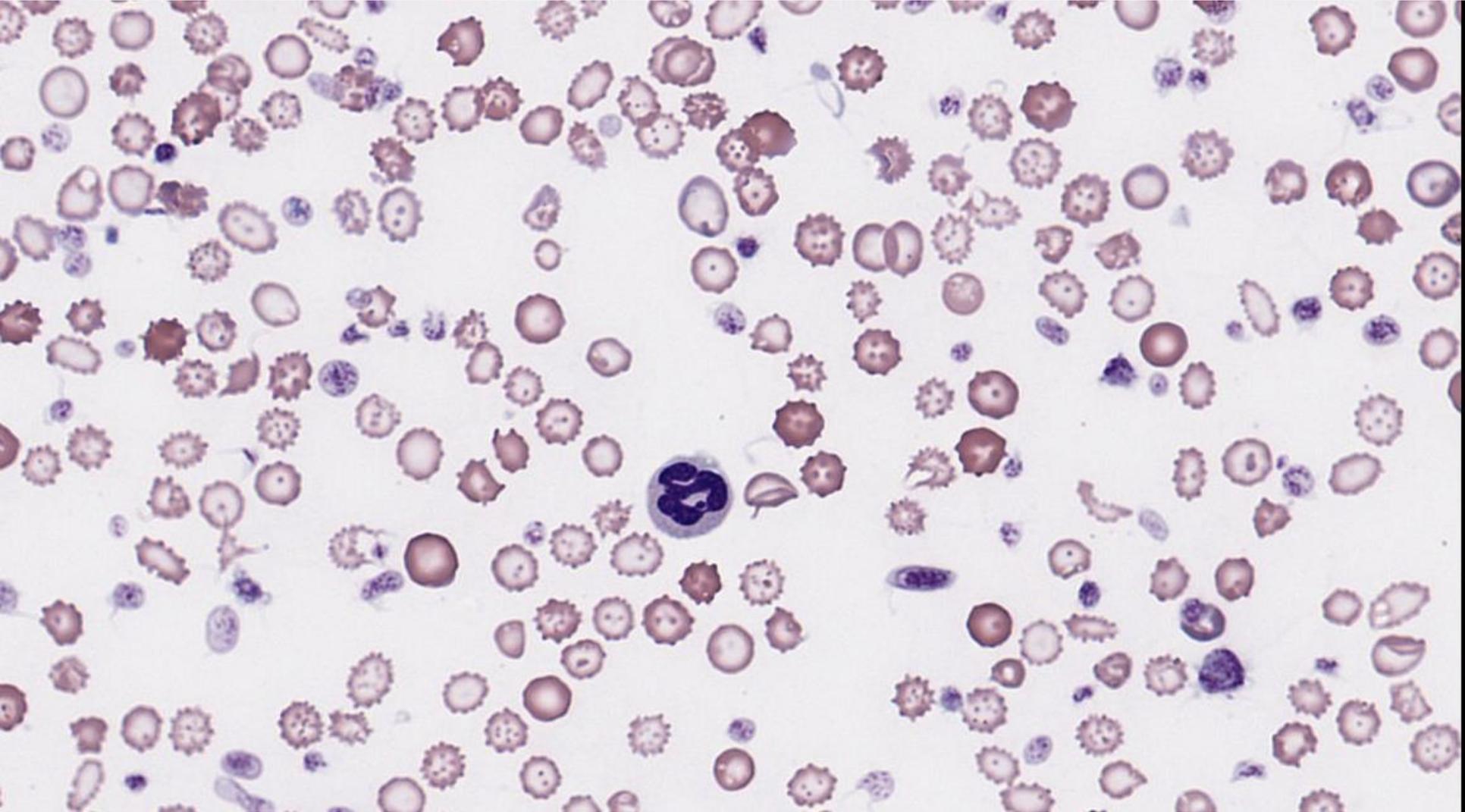
Text **NICOLEFERNAN414** to **37607** once to join, then **A, B, or C**





- Regenerative anemia
- Polychromasia
- Macrocytic
- (Hypochromic)

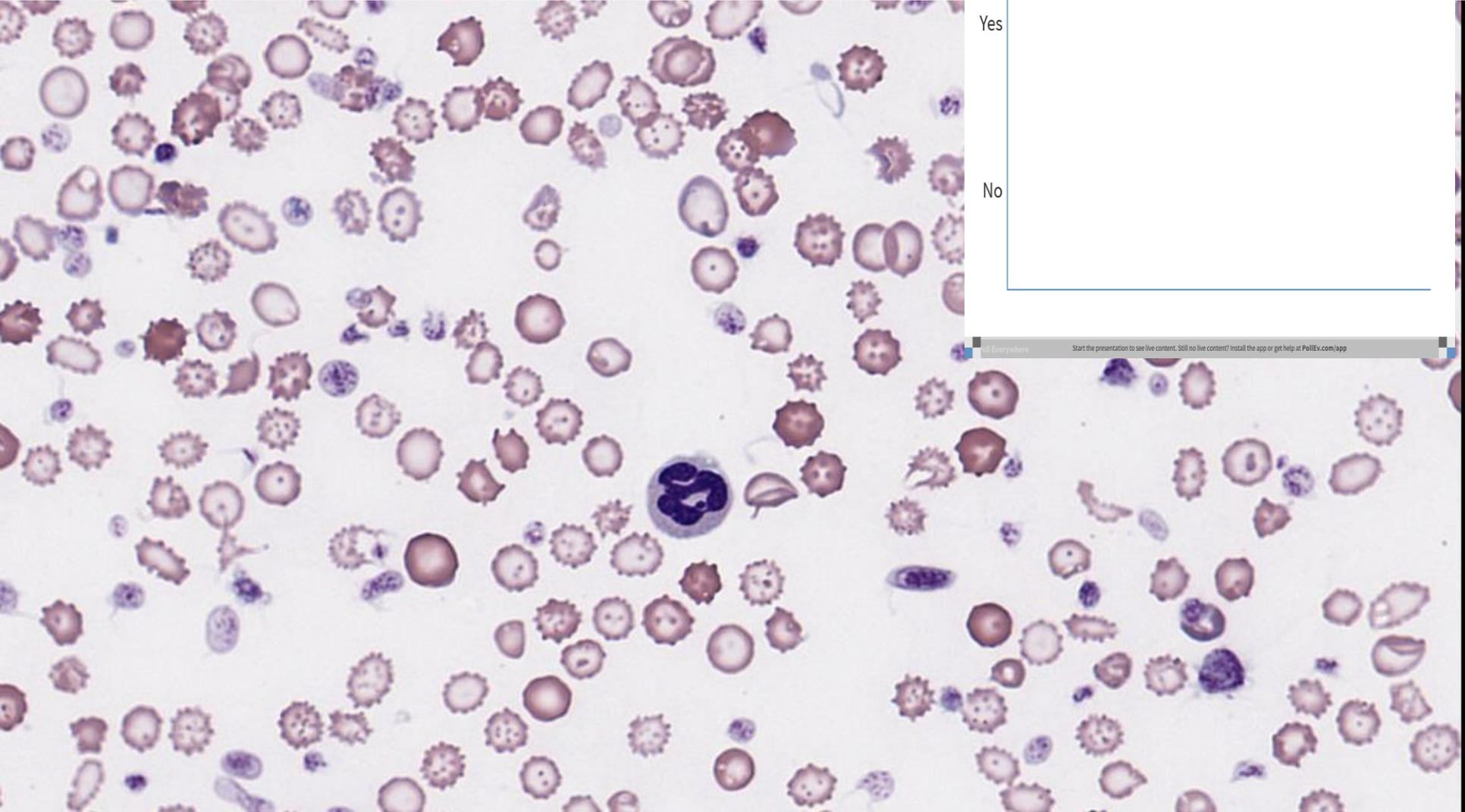


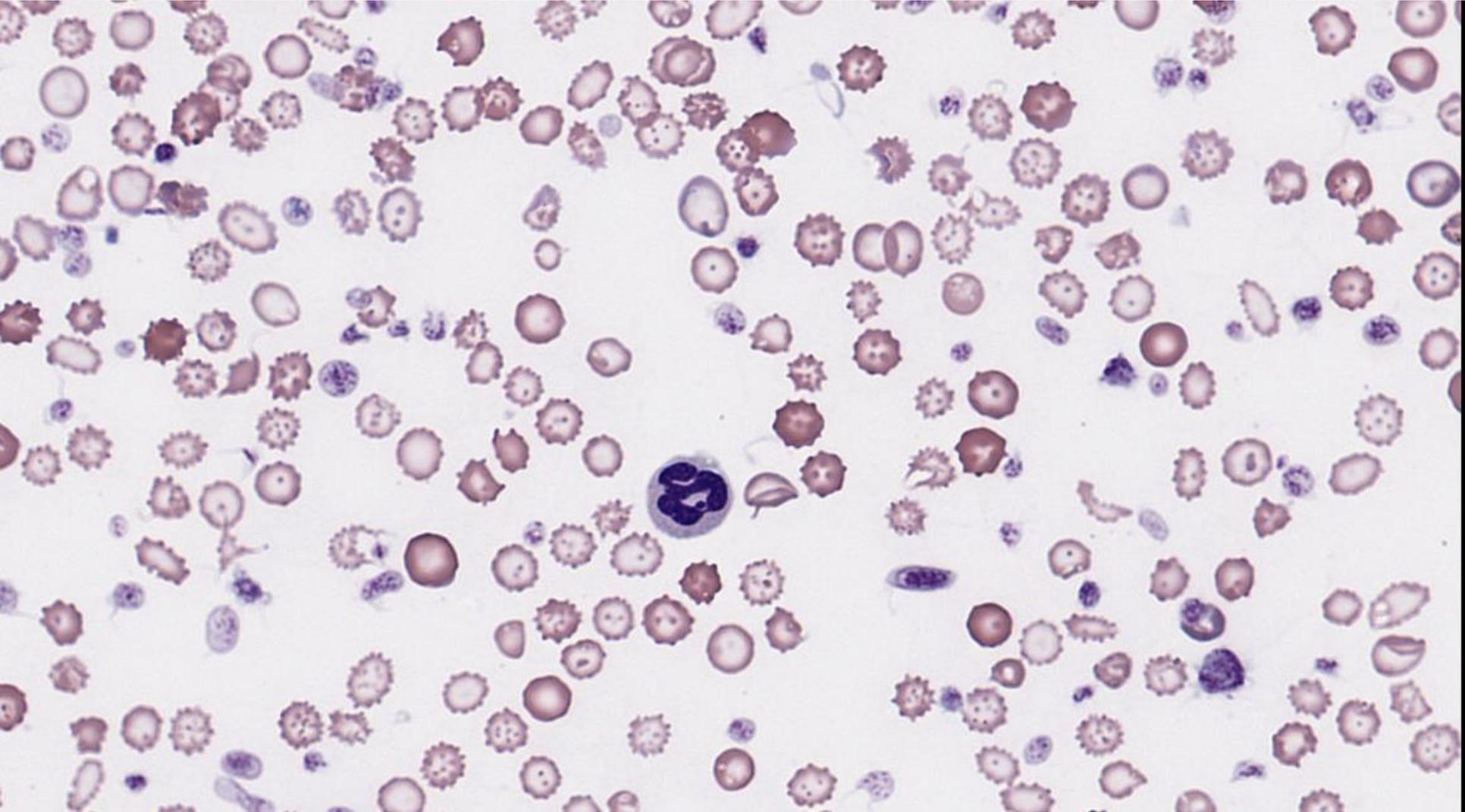


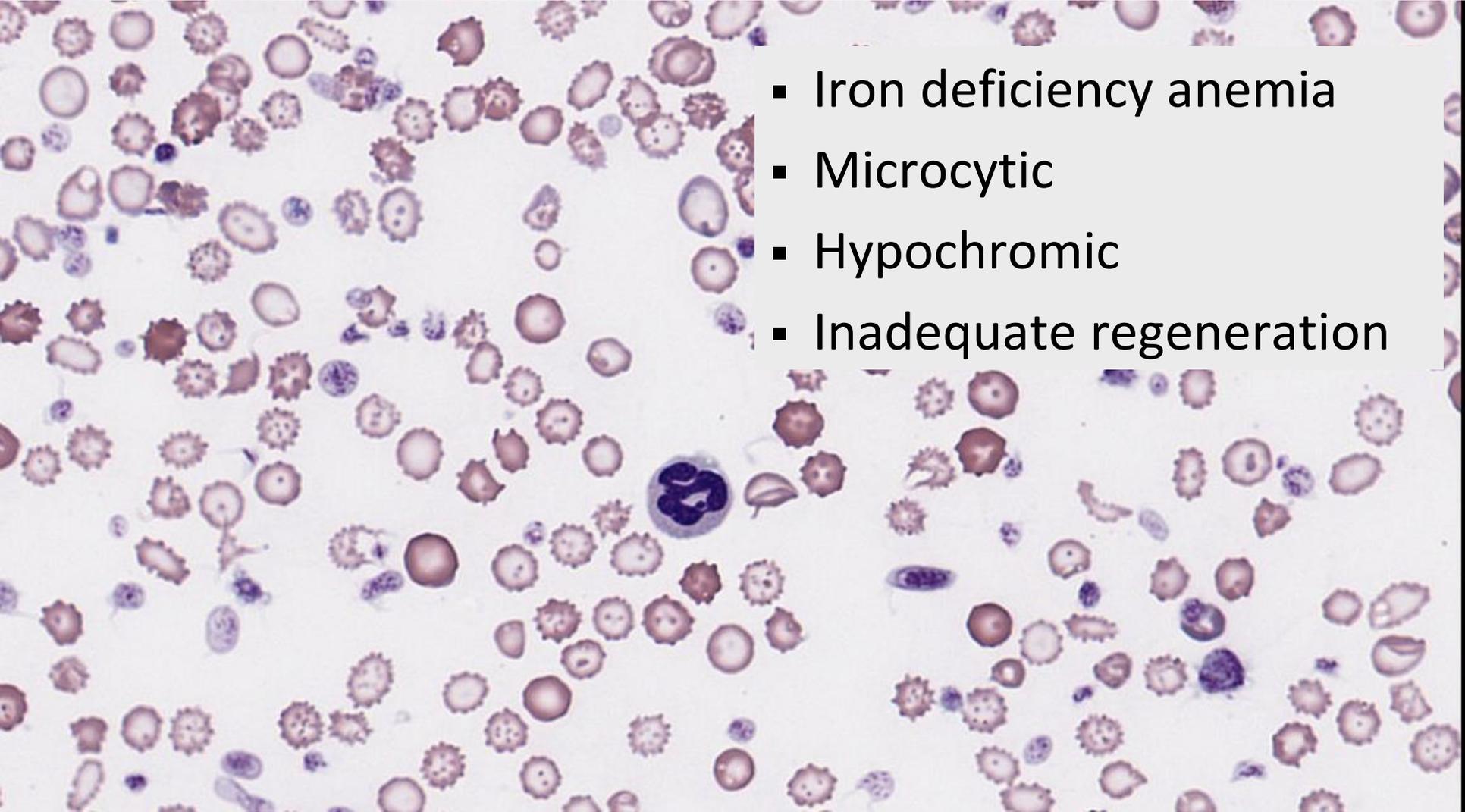
Is the anemia regenerative?

Yes

No



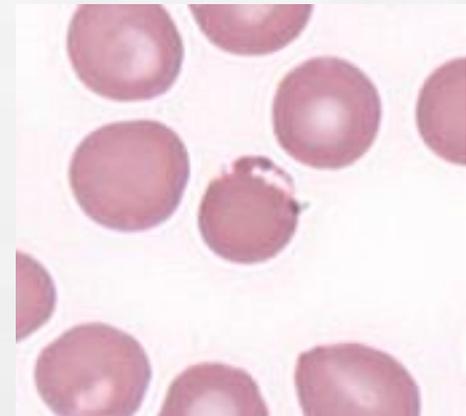
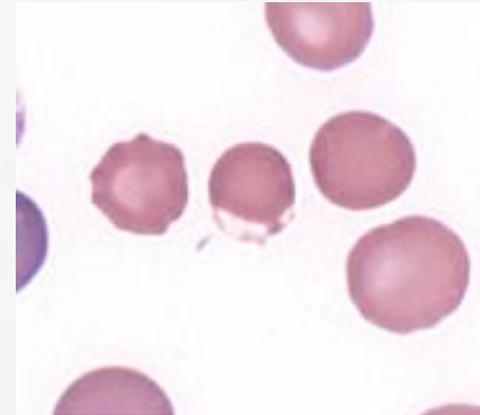
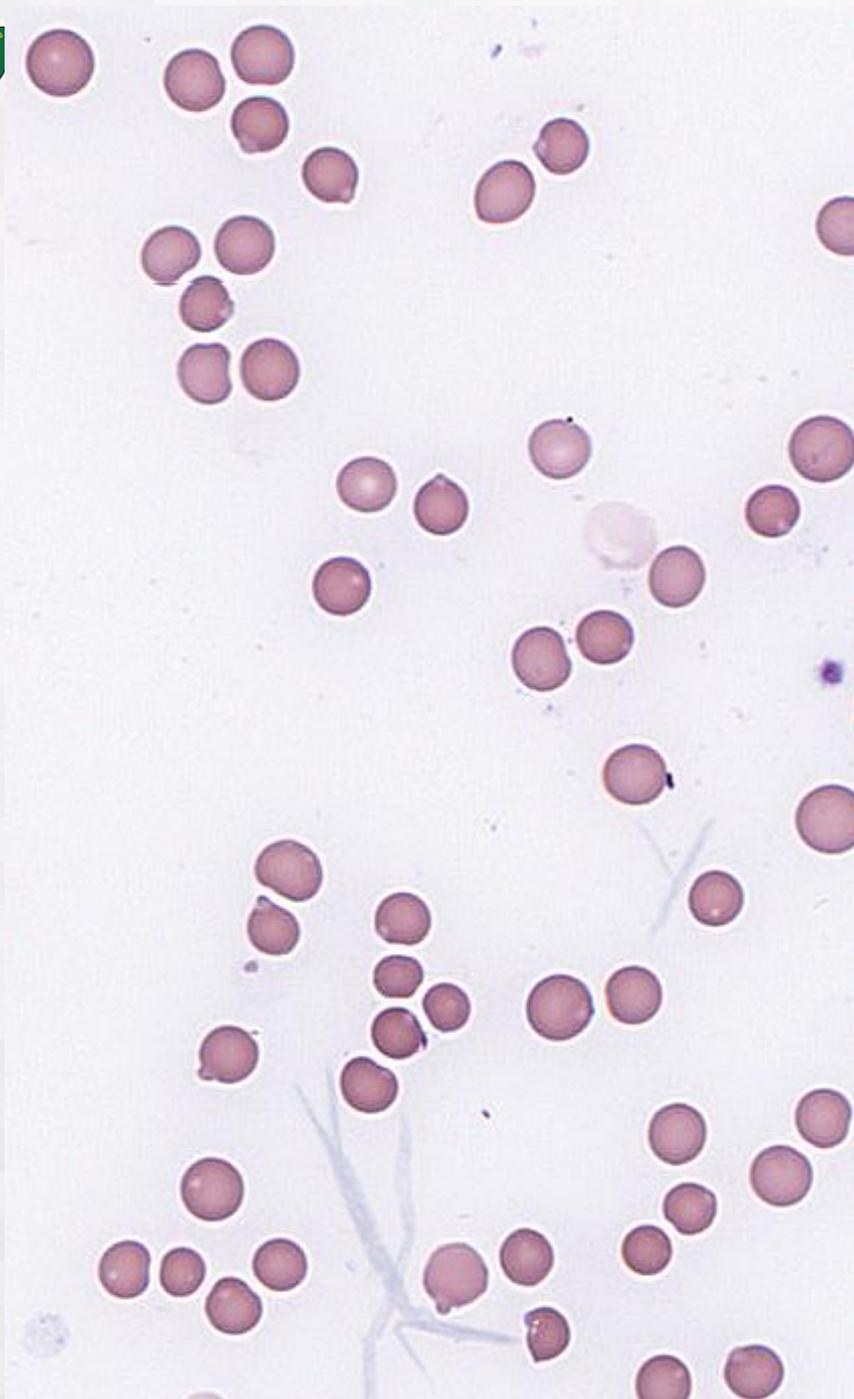




- Iron deficiency anemia
- Microcytic
- Hypochromic
- Inadequate regeneration



RBC Case 3



What is the cause of the anemia?

Oxidative
damage

Iron deficiency

Blood loss

IMHA

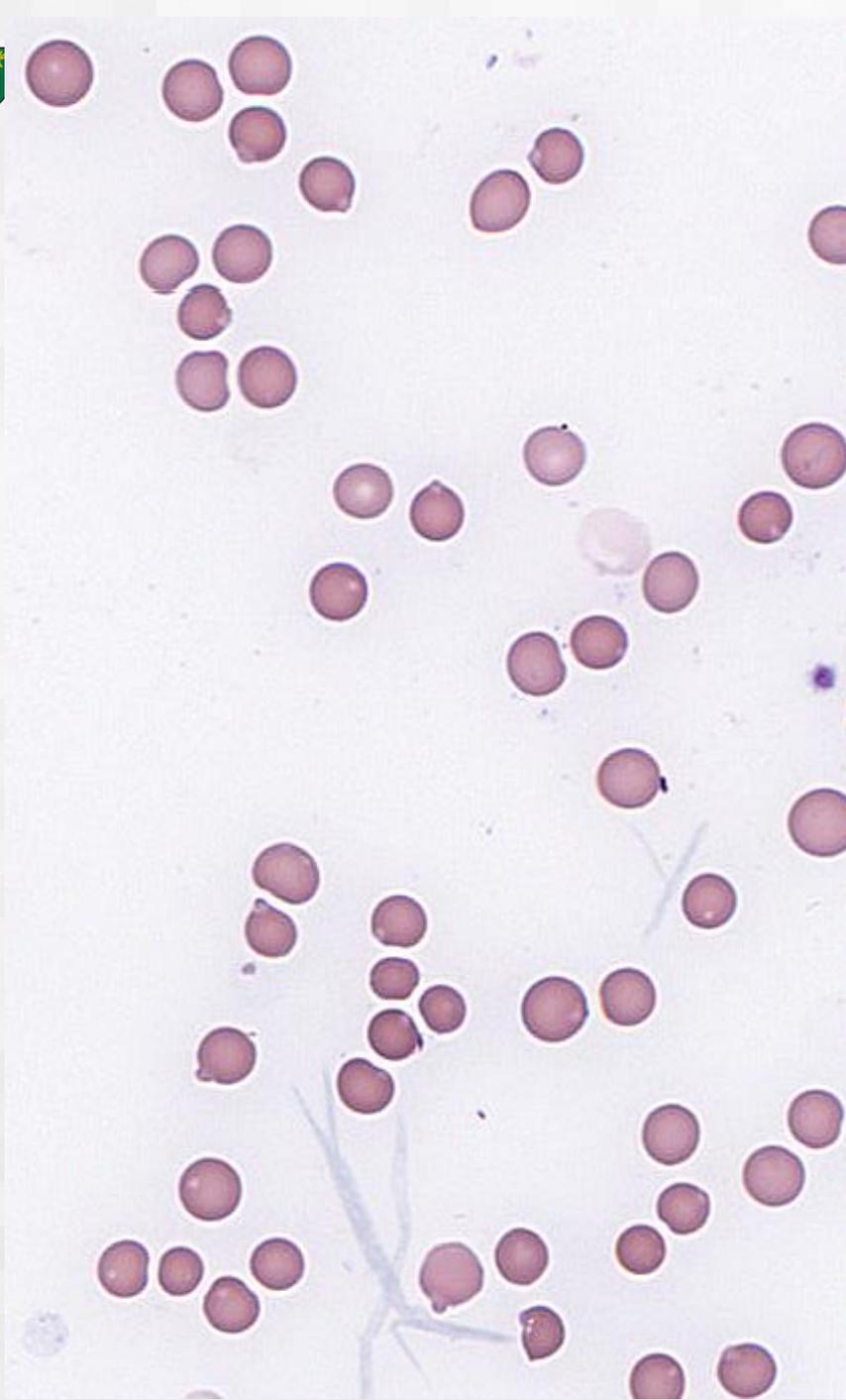
DIC



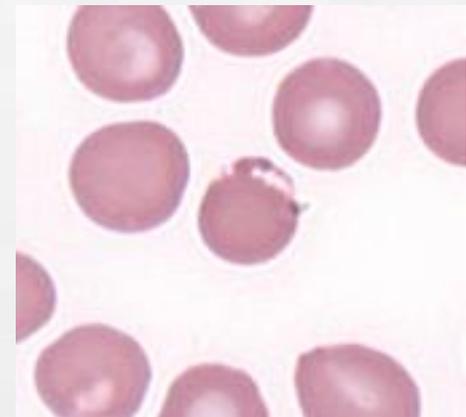
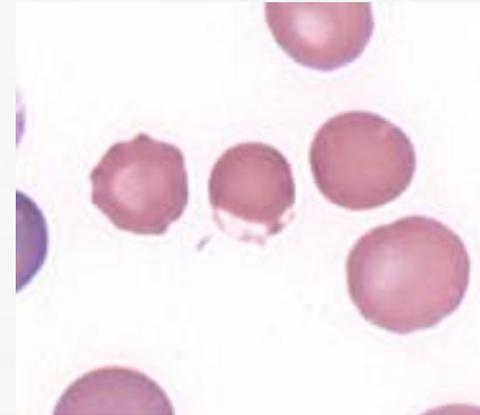
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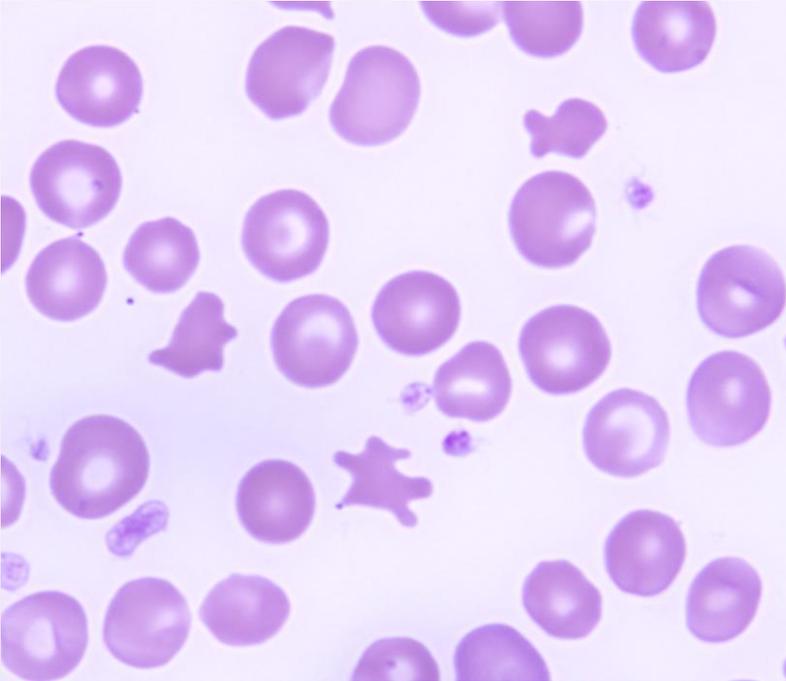
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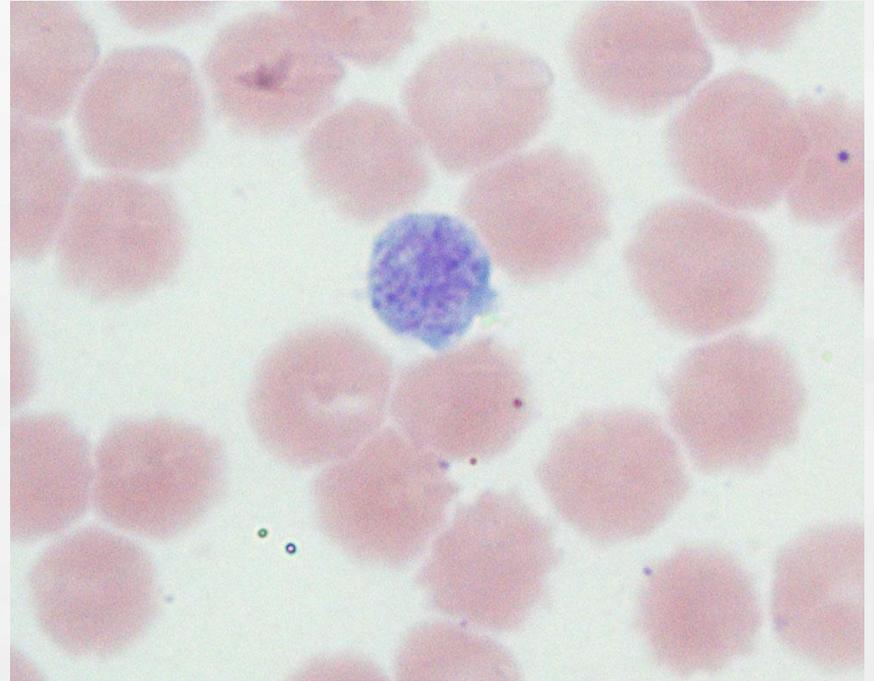
Oxidative Damage



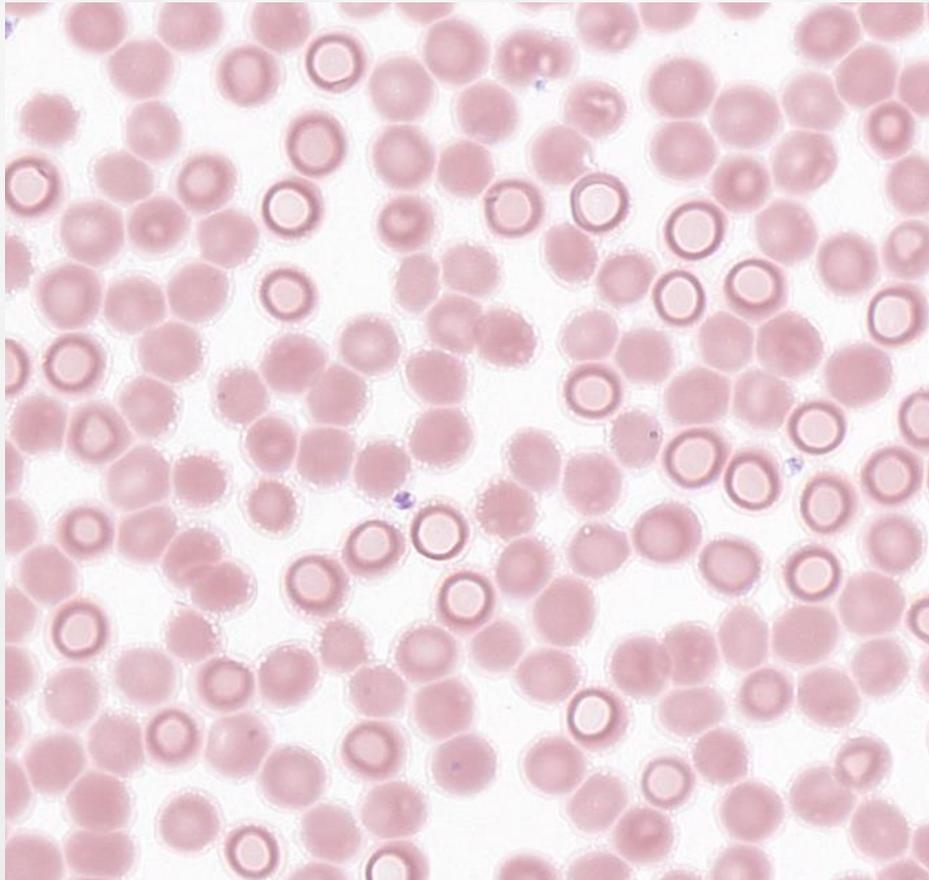
Acanthocytes



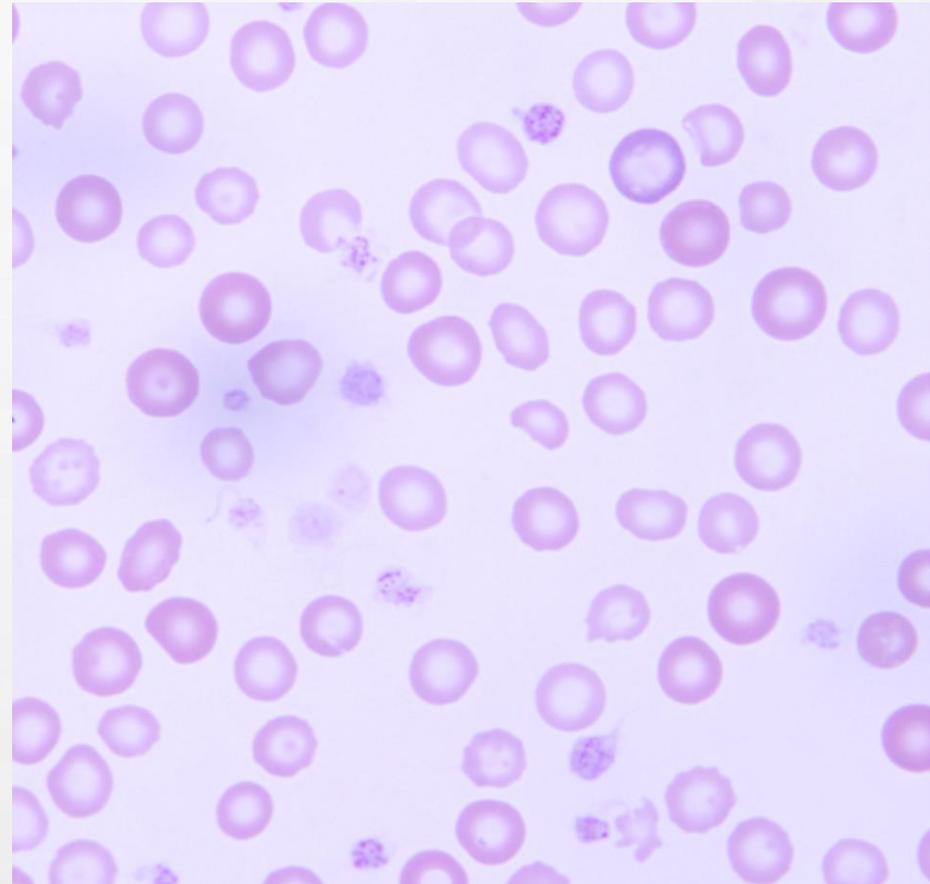
Echinocytes



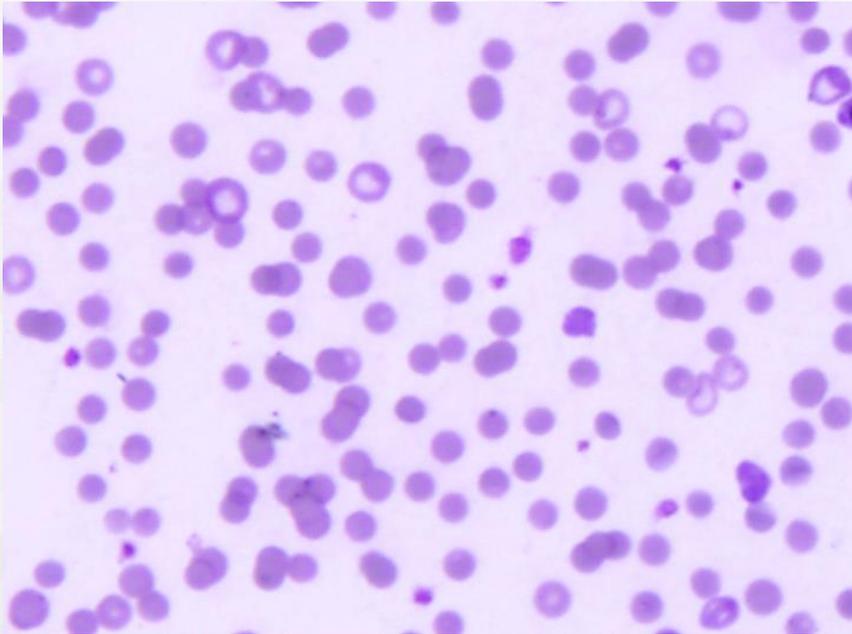
Torocytes



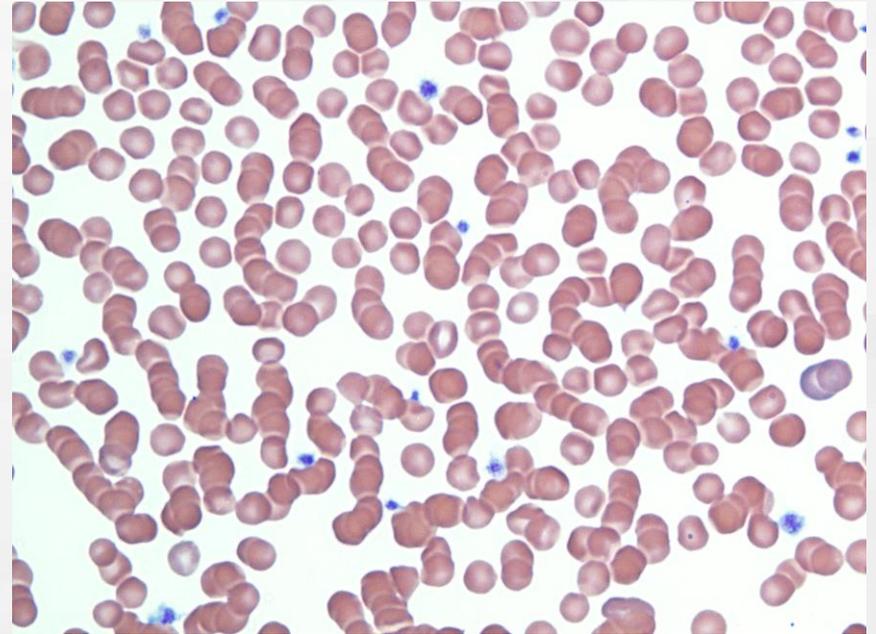
Hypochromasia



Agglutination



Rouleaux

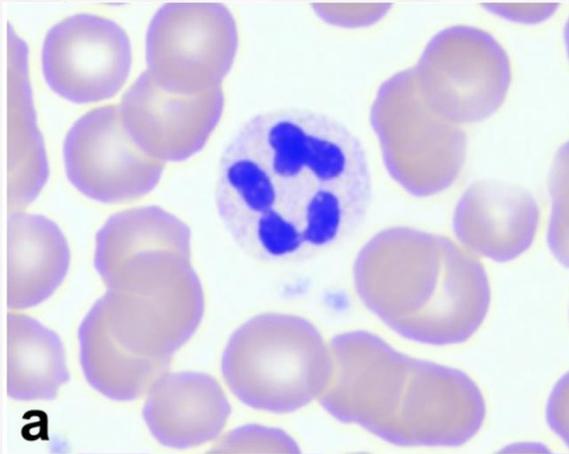


Questions?

WBC coming up...

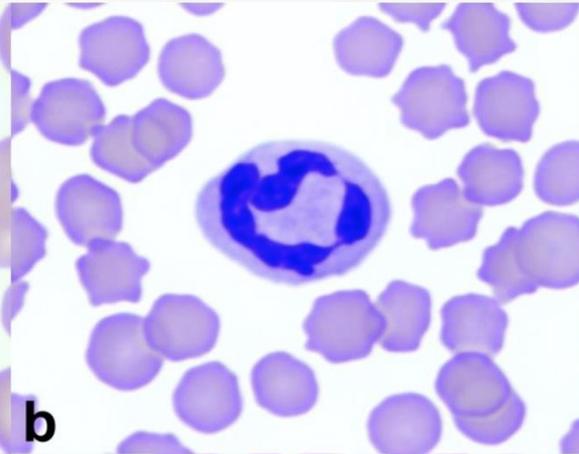
Leukocyte Review

Dog



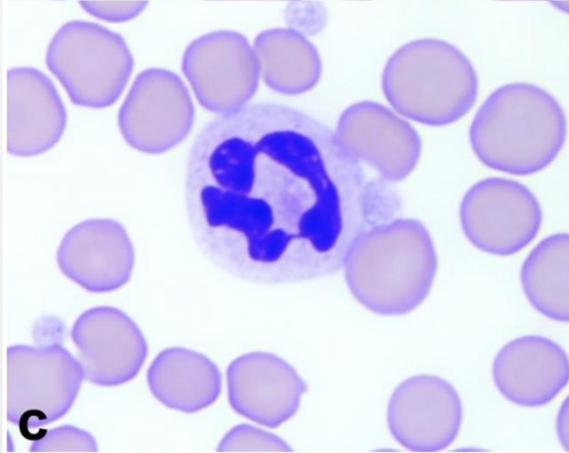
a

Cat



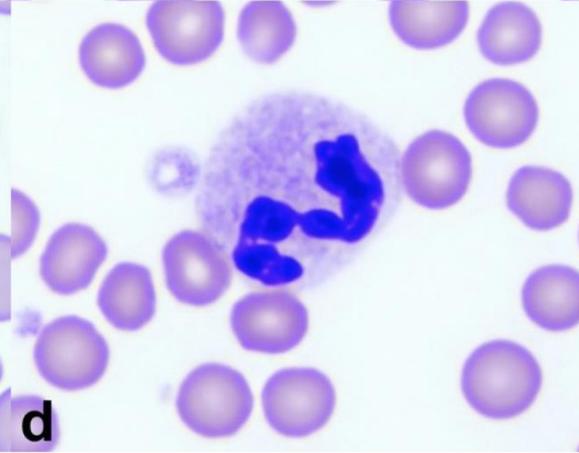
b

Horse



c

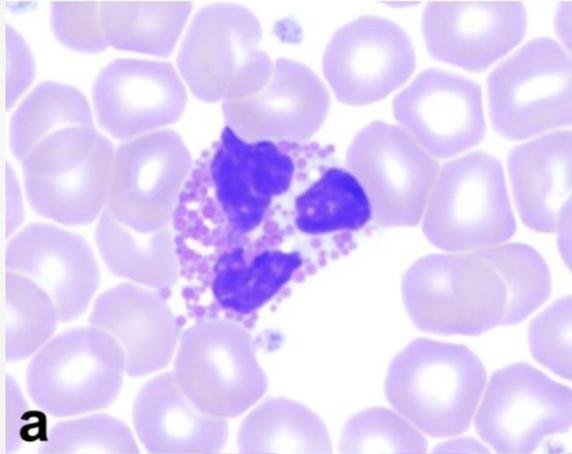
Cow



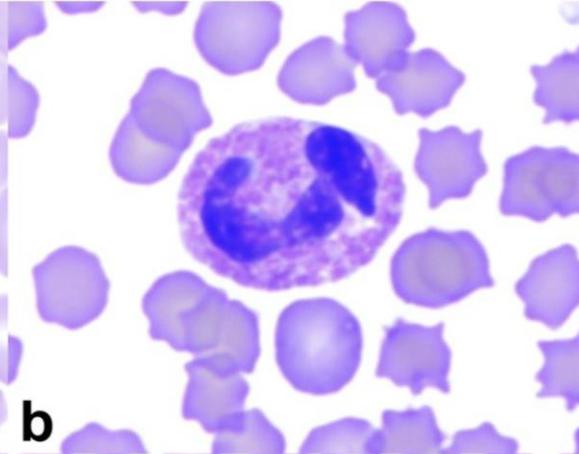
d

Leukocyte Review

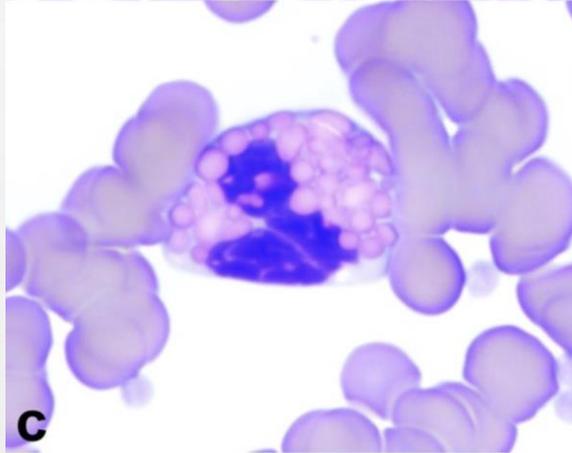
Dog



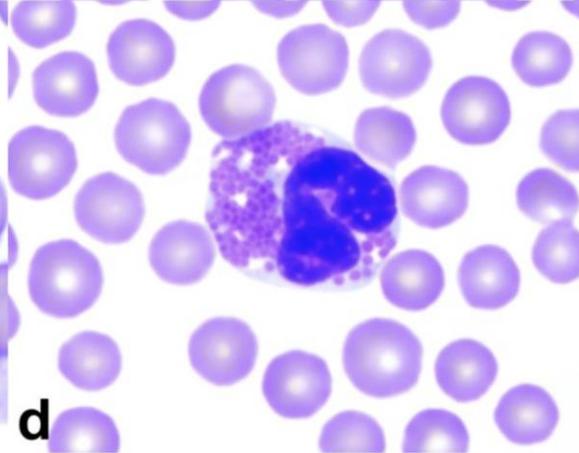
Cat



Horse

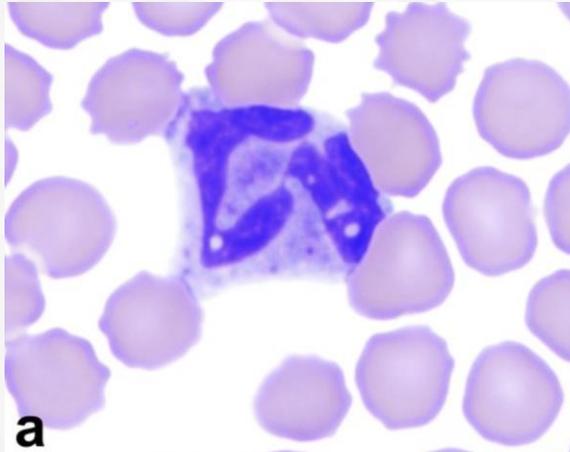


Cow



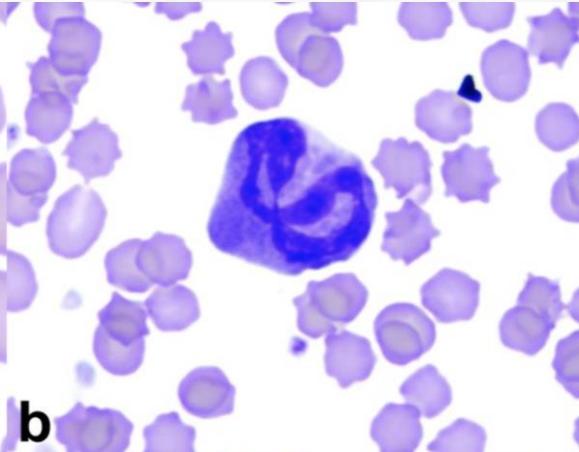
Leukocyte Review

Dog



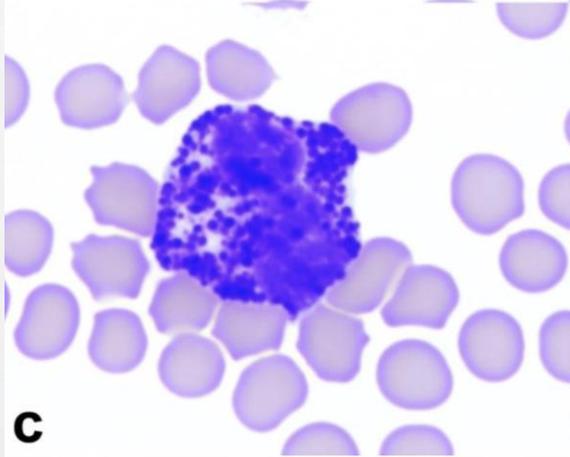
a

Cat



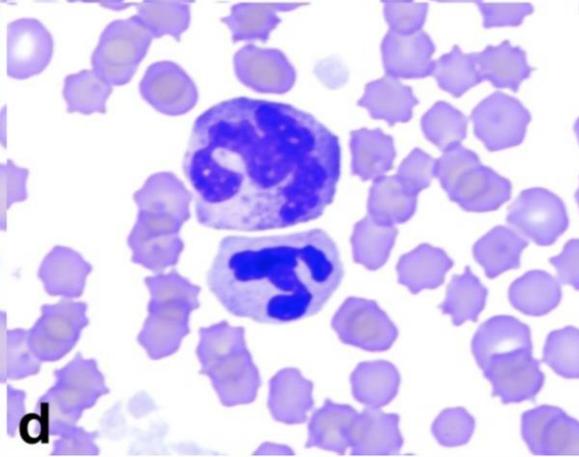
b

Horse



c

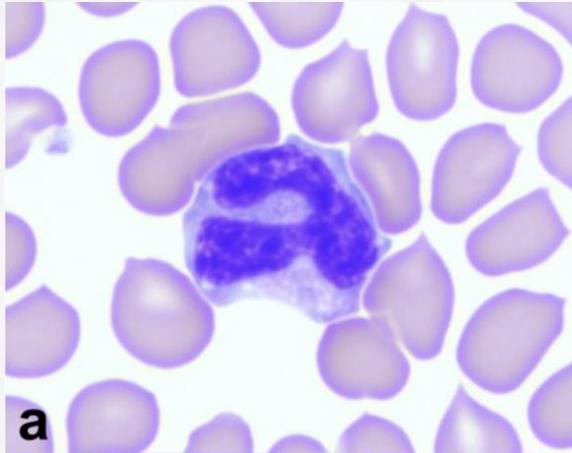
Cow



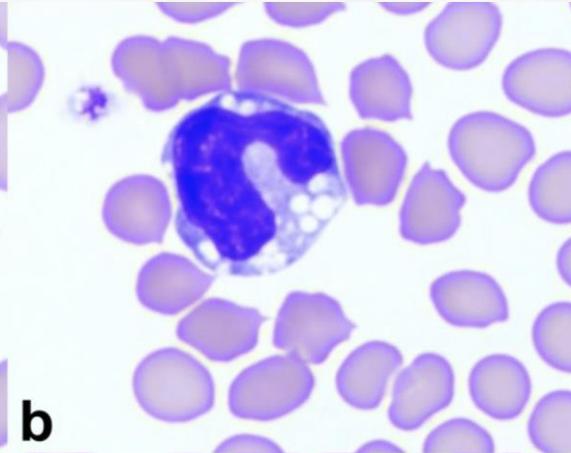
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Leukocyte Review

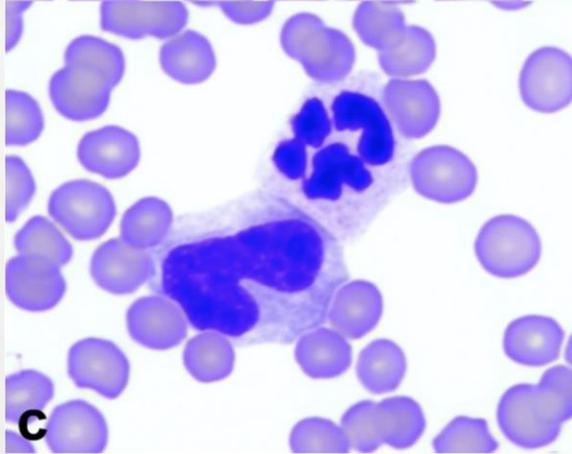
Dog



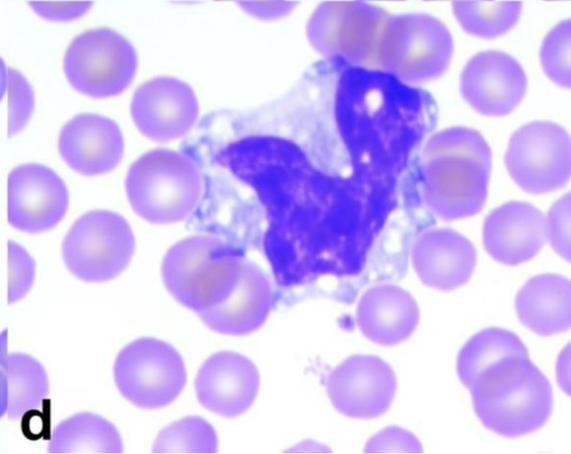
Cat



Horse

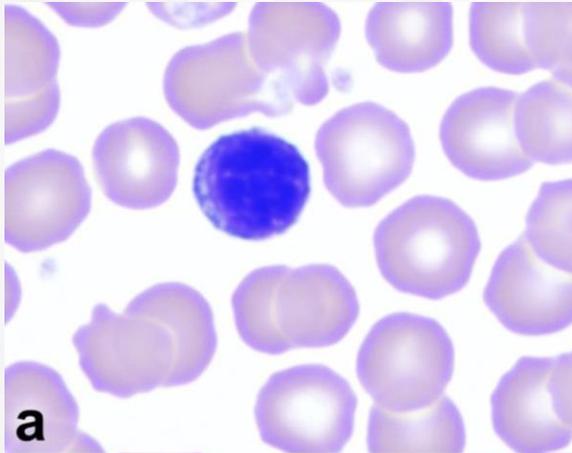


Cow

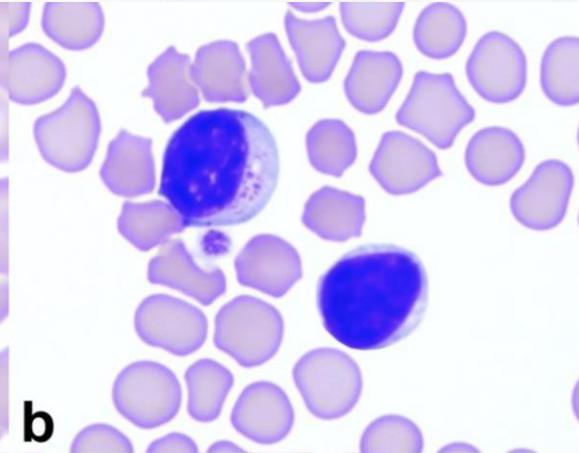


Leukocyte Review

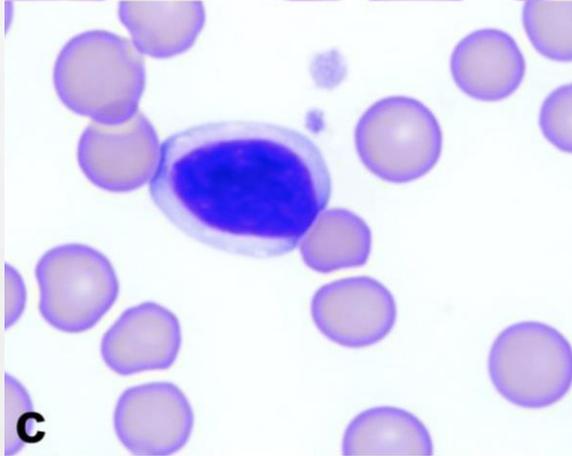
Dog



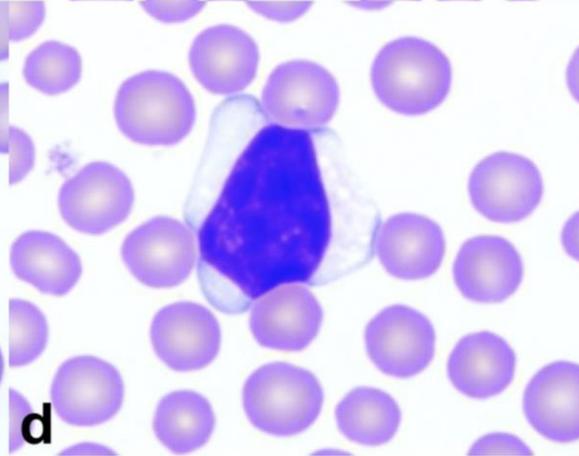
Cat

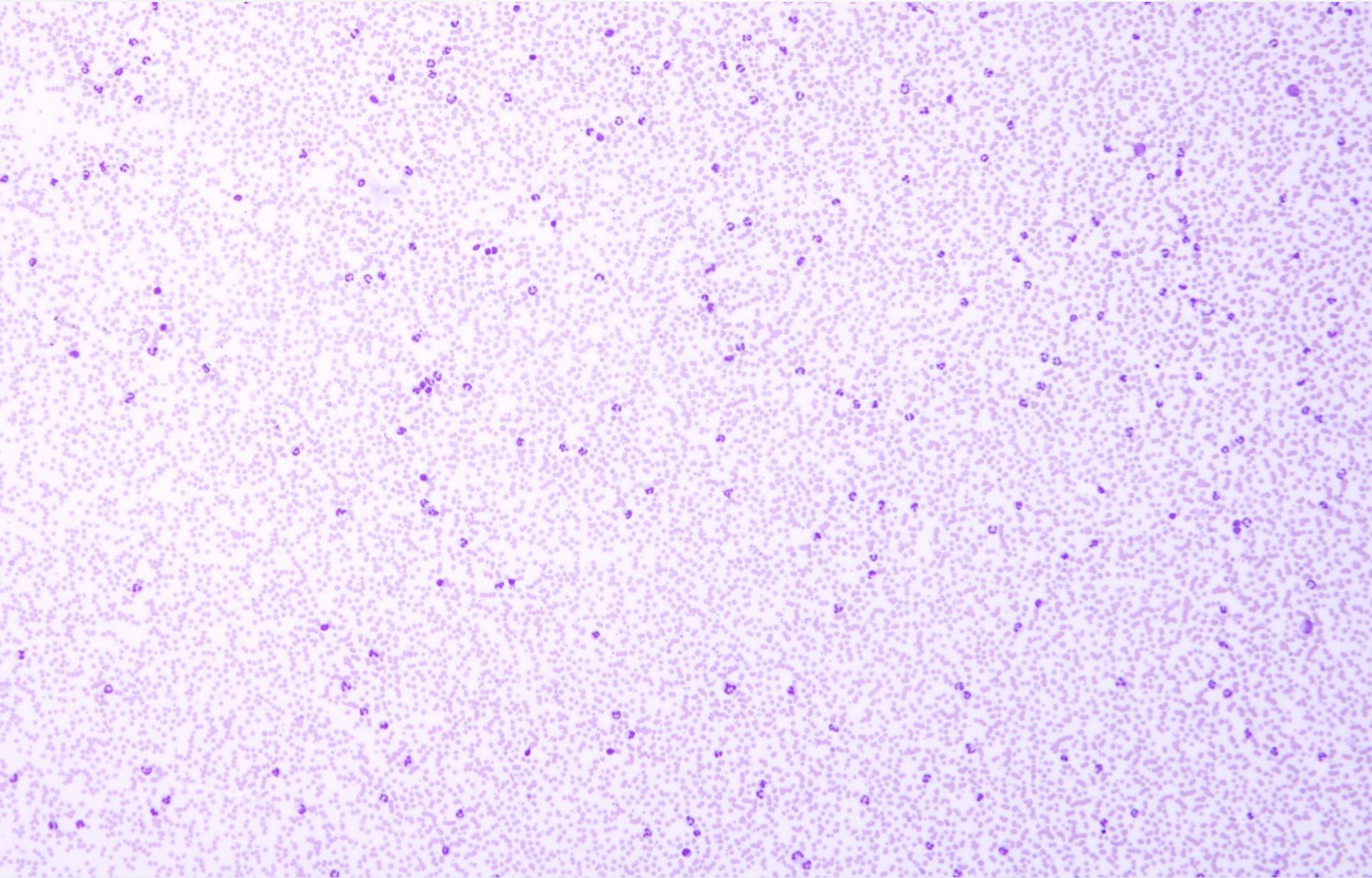


Horse



Cow





Are WBC numbers...

Decreased

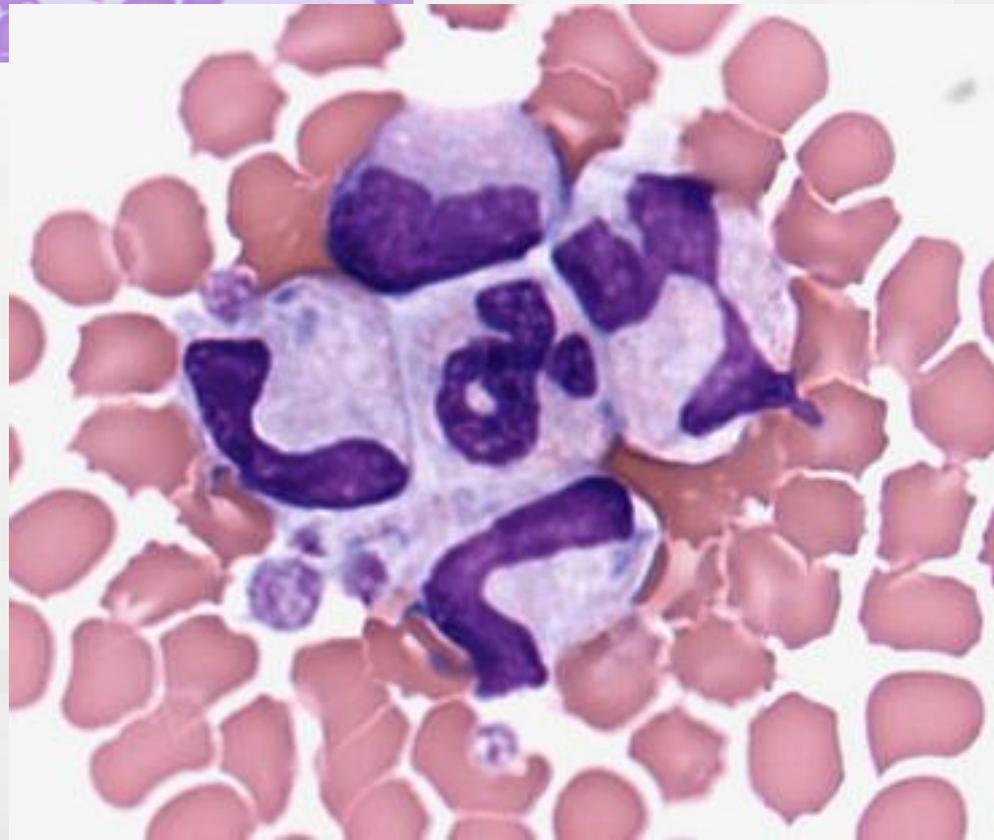
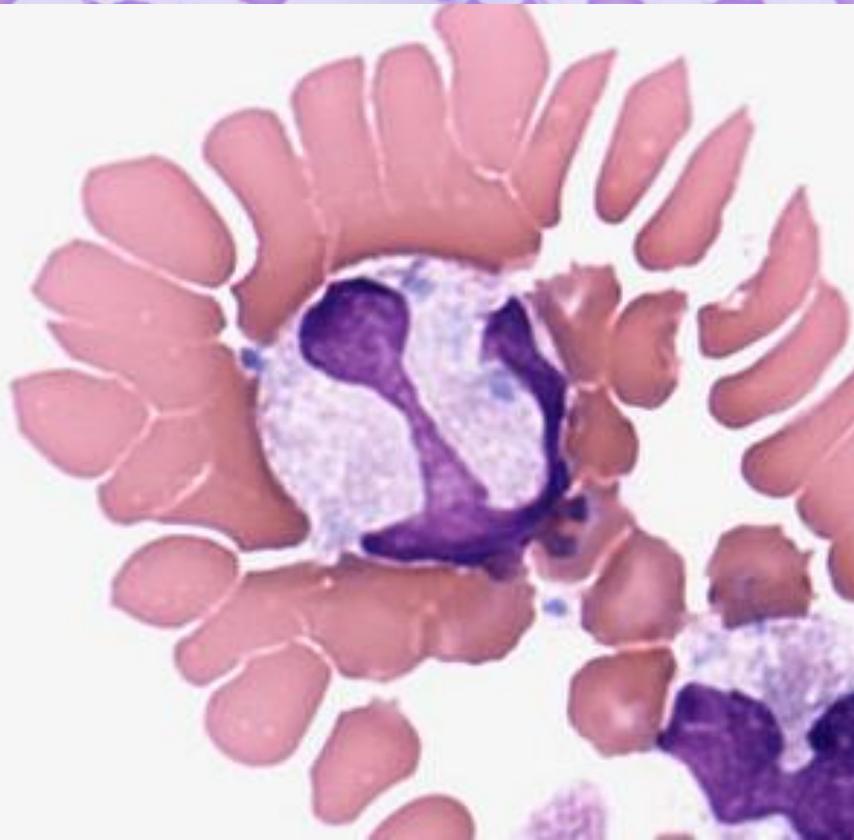
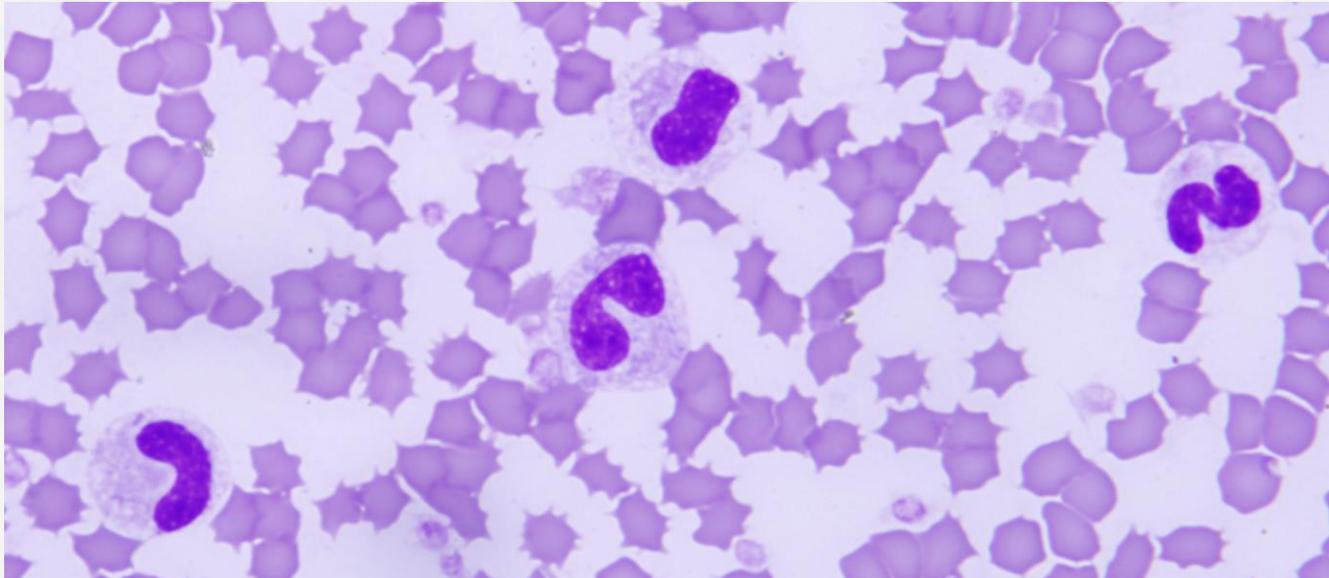
Within reference
intervals

Increased

WBC Estimate 10x

- Count the leukocytes in at least 3 fields in the **monolayer**
- Divide the total number of leukocytes counted by the number of fields to obtain the average number per field
- Divide the average number of leukocytes counted by 4 in order to approximate the number of leukocytes $\times 10^9/L$

WBC Case 1



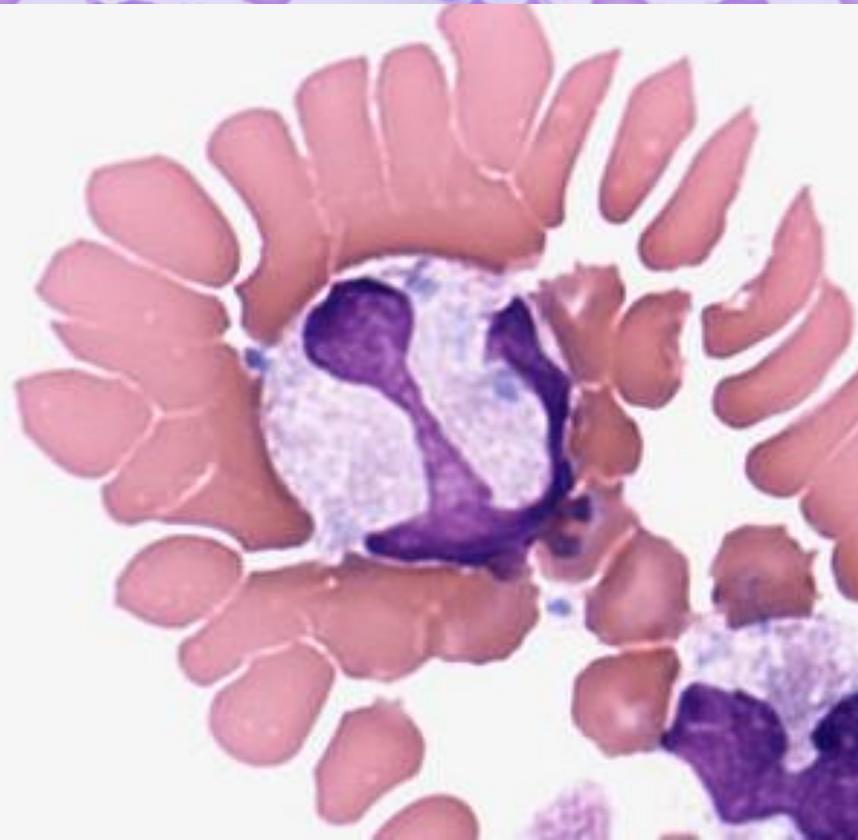
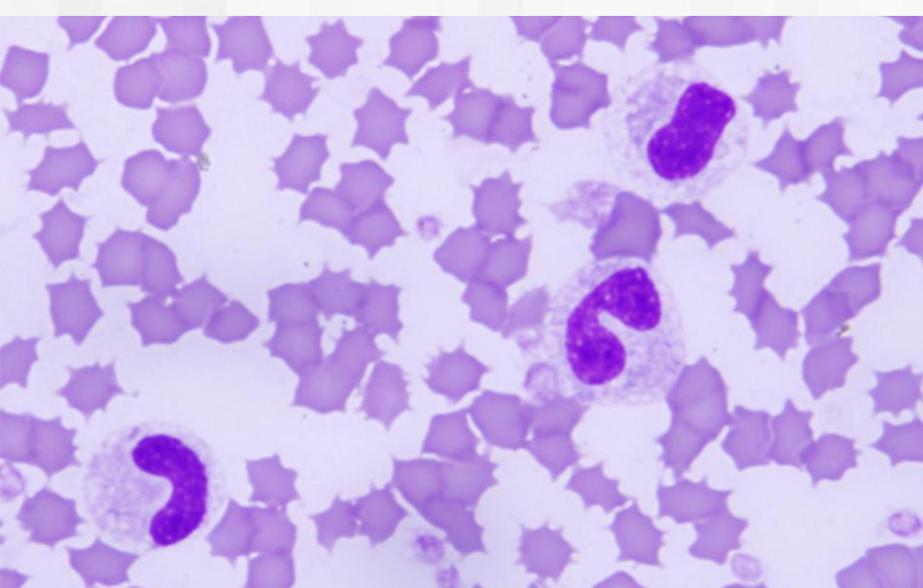
What does the WBC morphology indicate?

There is a left shift

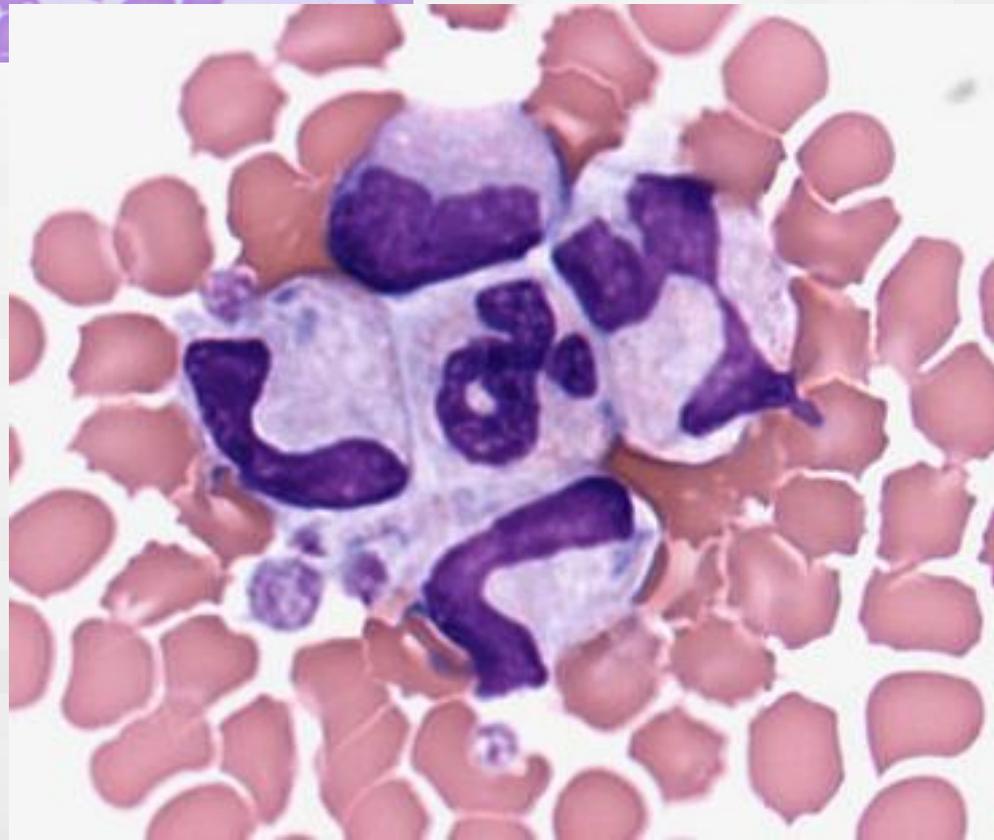
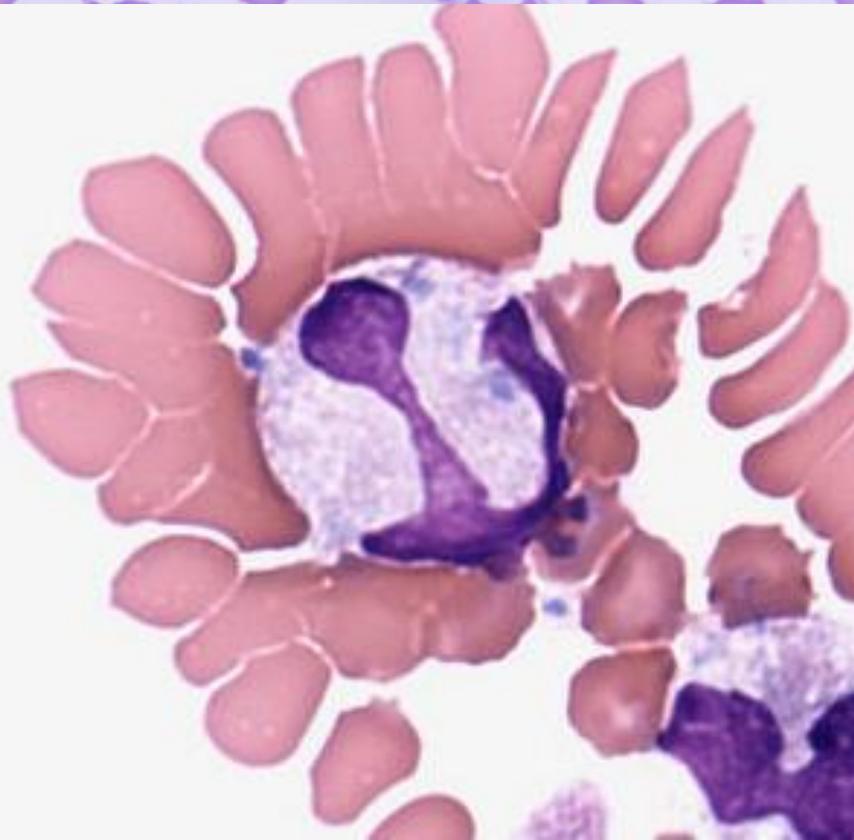
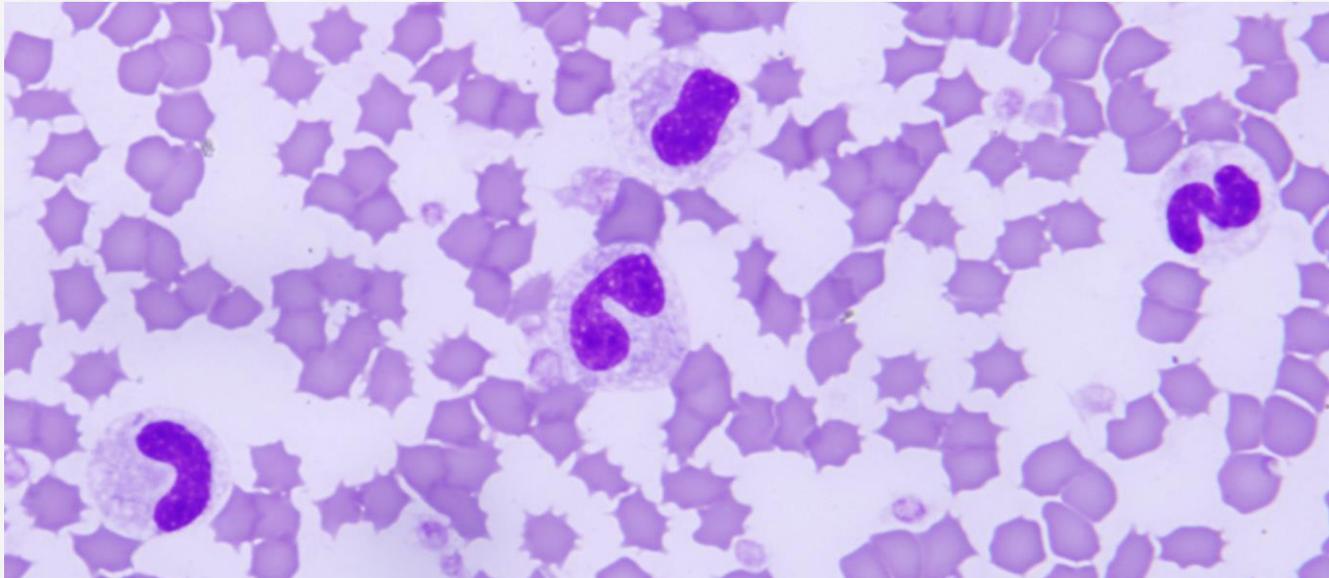
There is toxic change

There is both a left shift and toxic change

There is neither a left shift nor toxic change

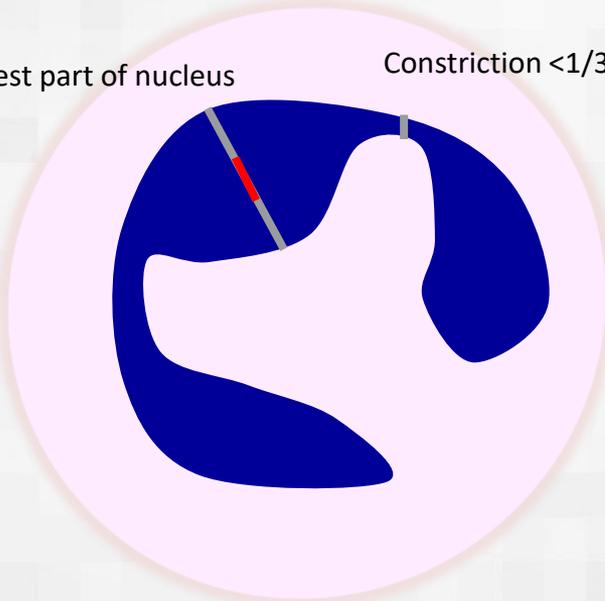


WBC Case 1 Left Shift



Widest part of nucleus

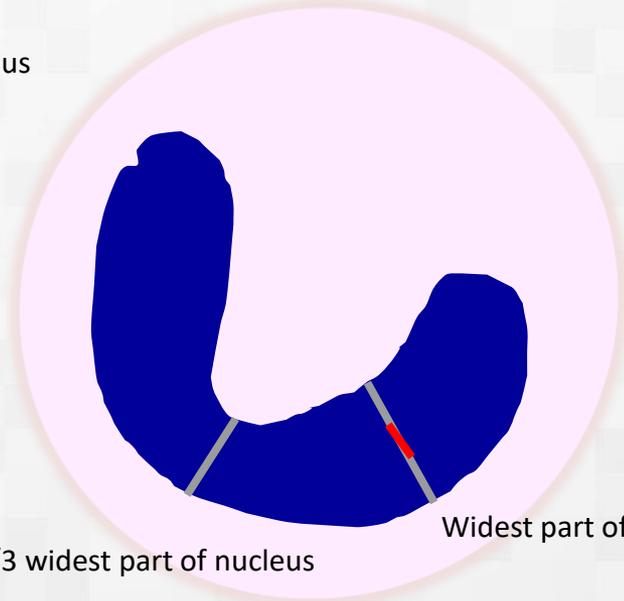
Constriction $< 1/3$ widest part of nucleus



Mature seg

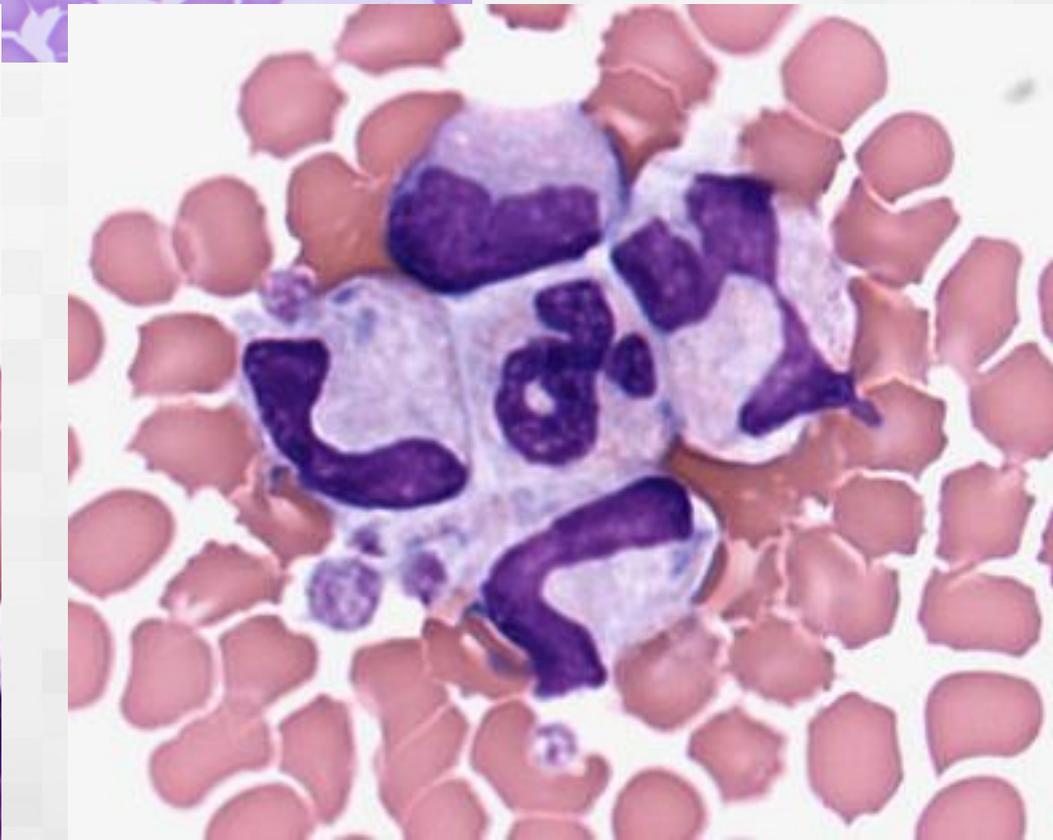
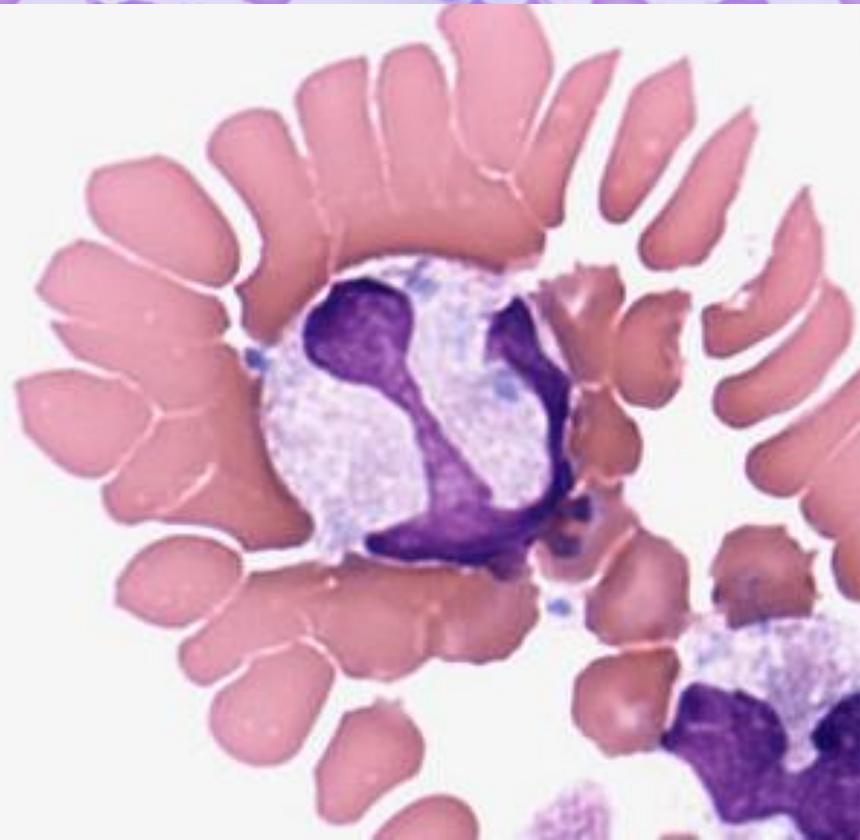
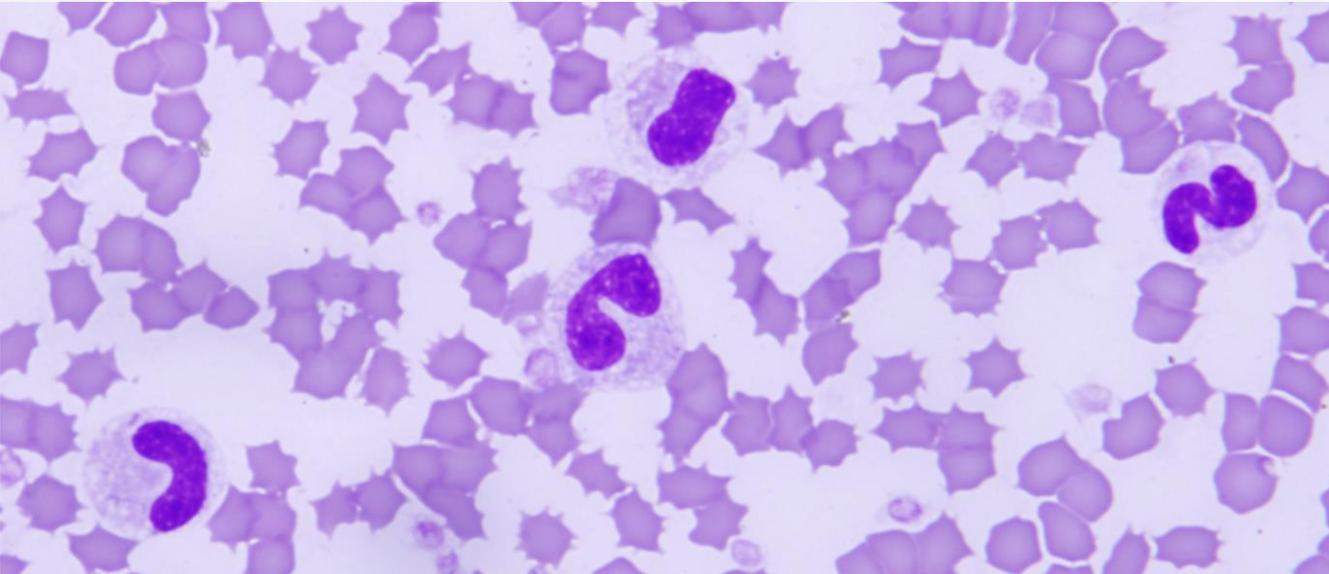
Constriction $> 1/3$ widest part of nucleus

Widest part of nucleus



Band

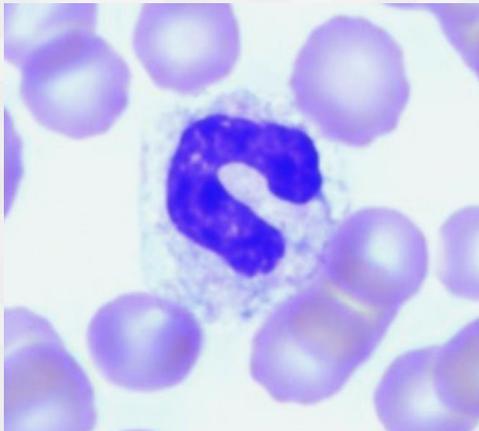
WBC Case 1 Toxic Change



Source of Confusion

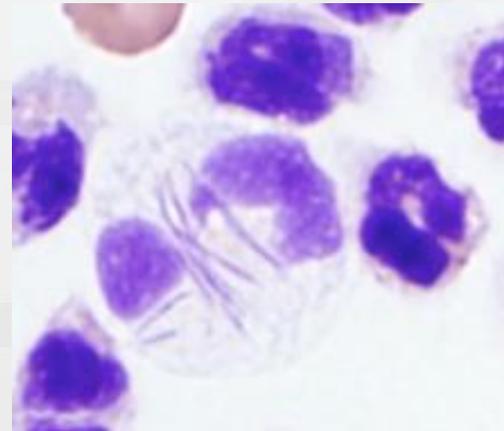
Toxic change

- In blood (blood smears)
- Cytoplasmic changes
 - Basophilia
 - Foaminess
 - Dohle bodies
 - Toxic granulation



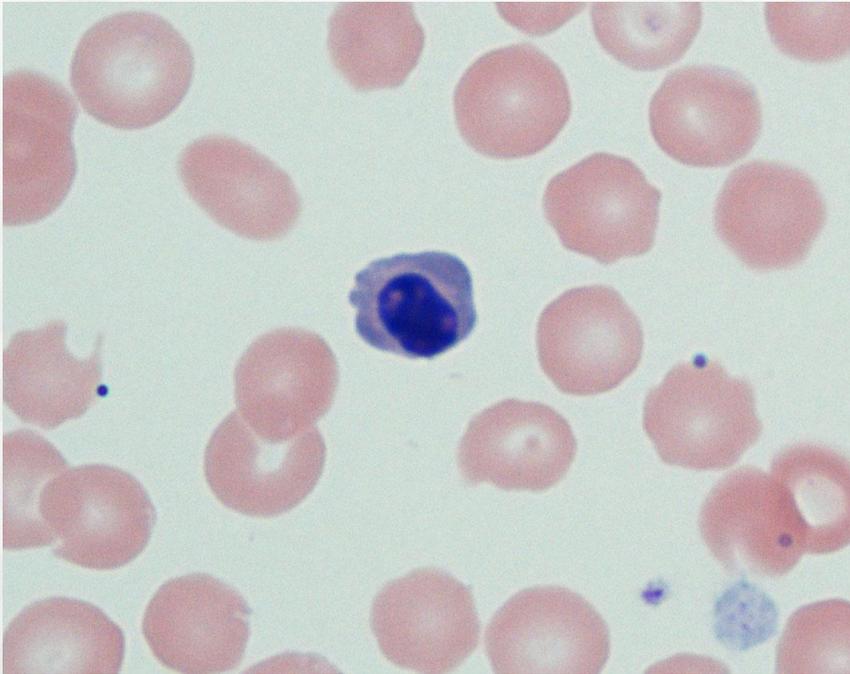
Degenerate neutrophils

- In tissues (cytology smears)
- Nuclear changes
 - Swelling
 - Pale-staining
 - Lysis
 - Hunt for bacteria

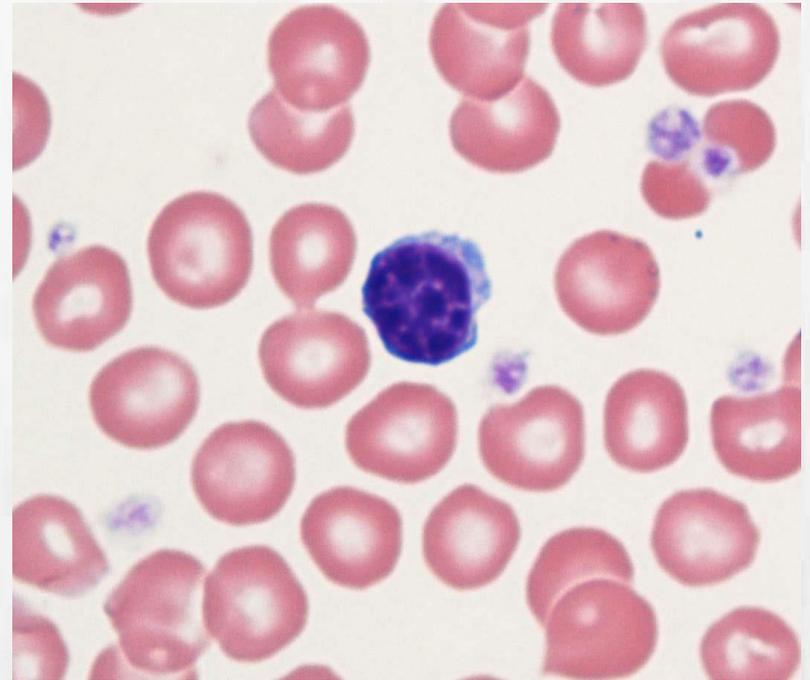


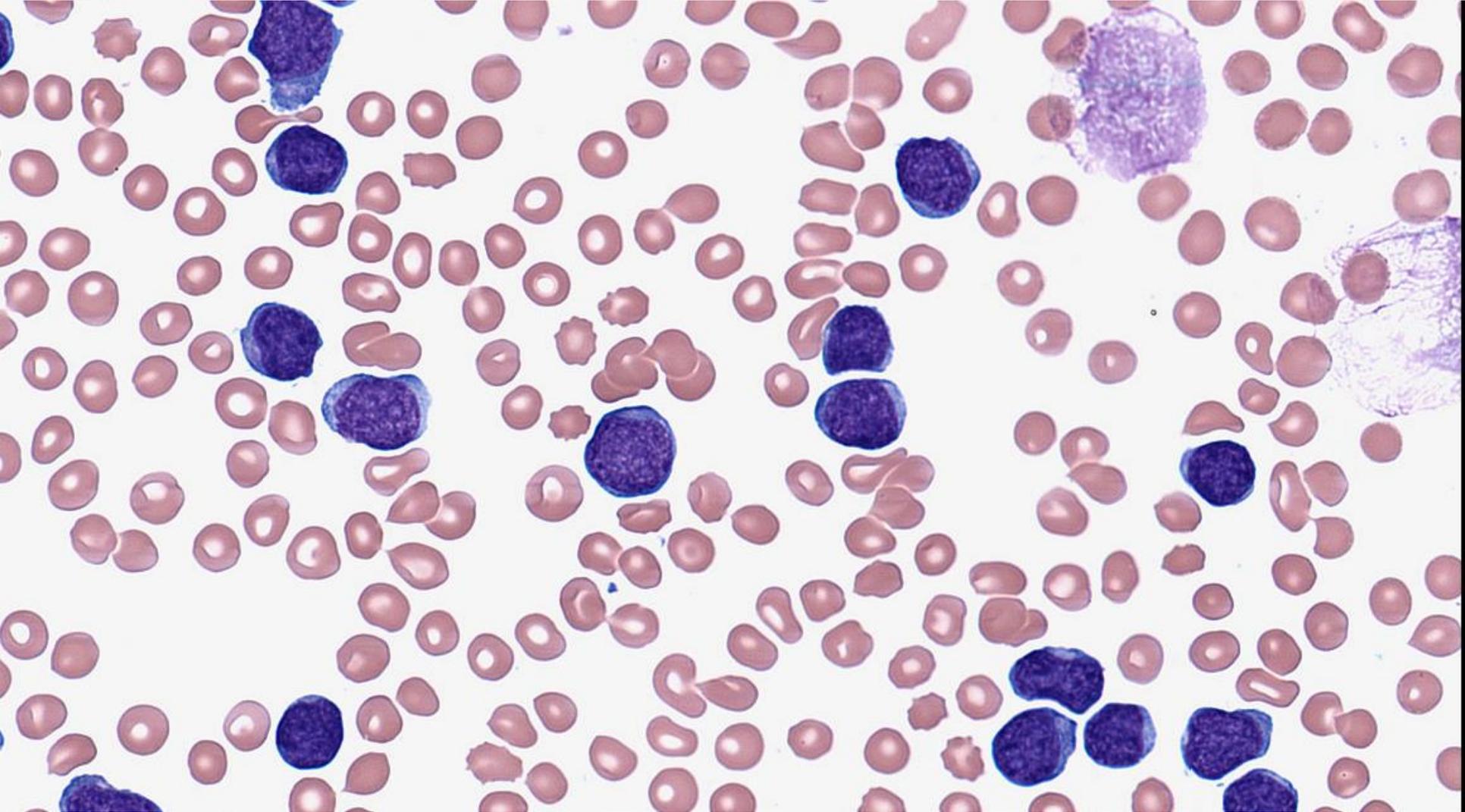
Source of Confusion

Nucleated RBC



Lymphocyte

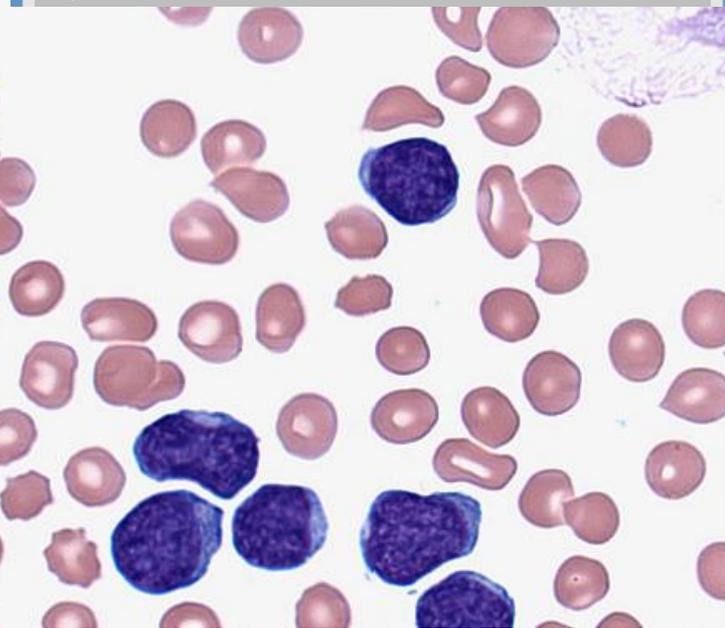
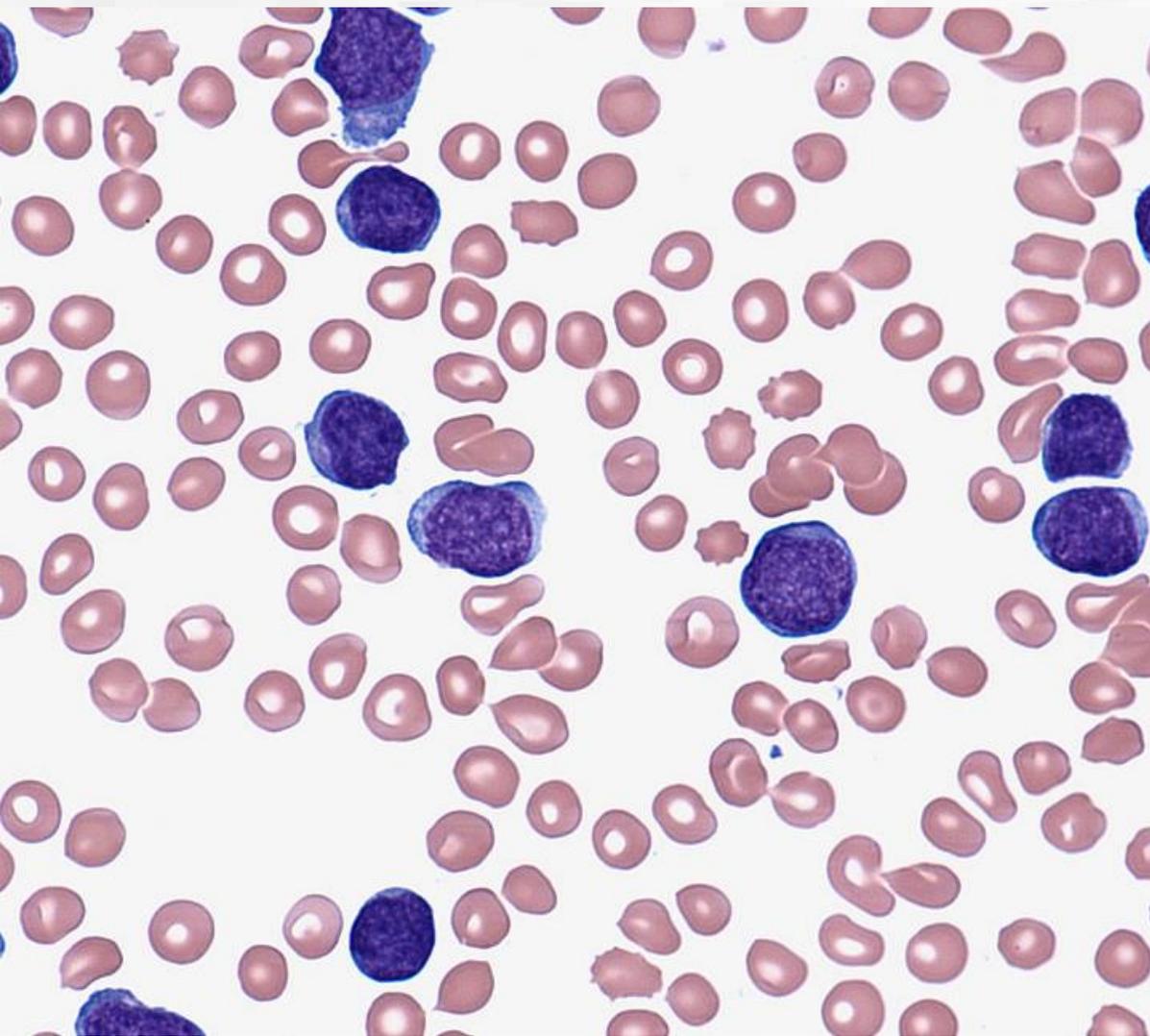


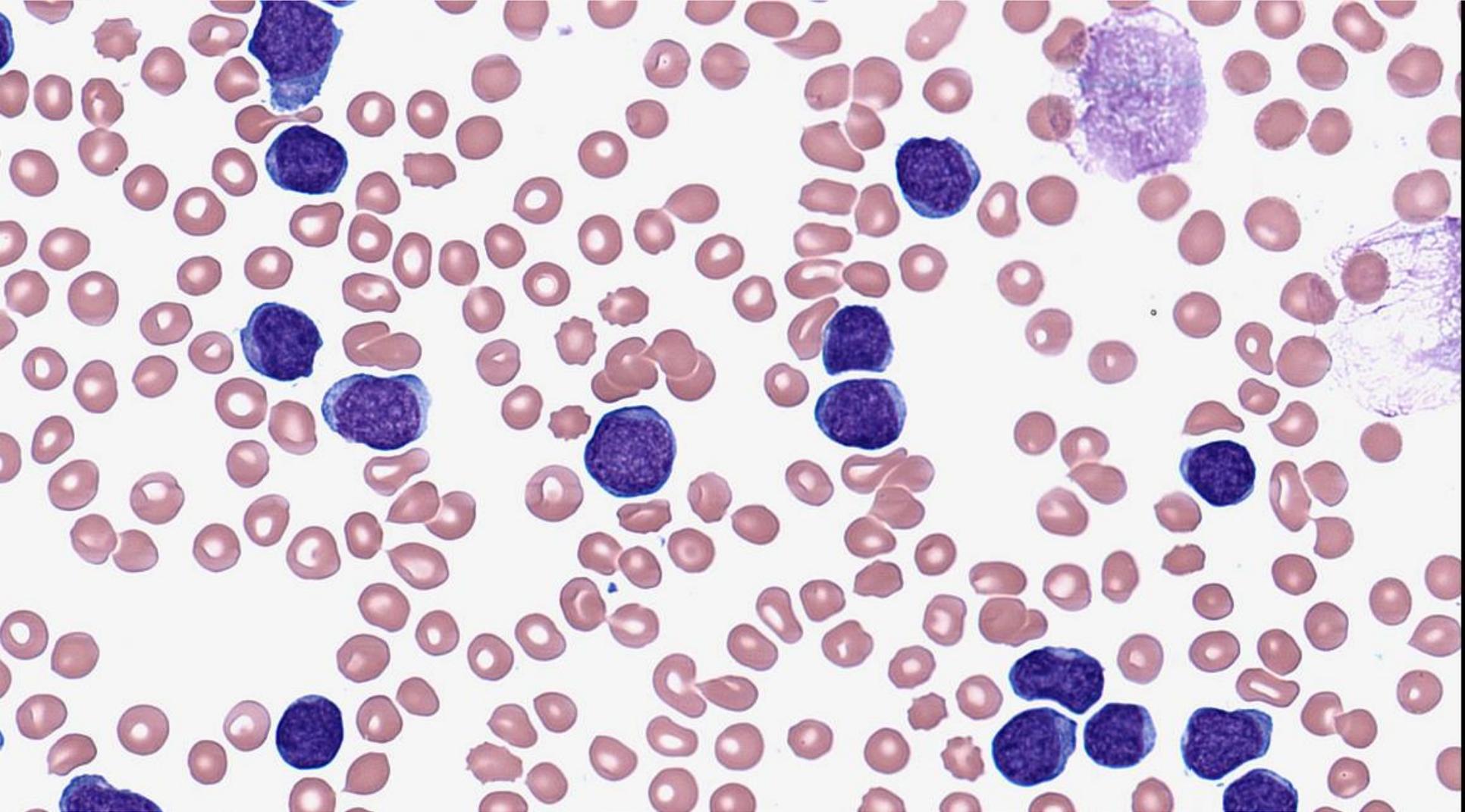


What is the most likely diagnosis?

Reactive
lymphocytosis

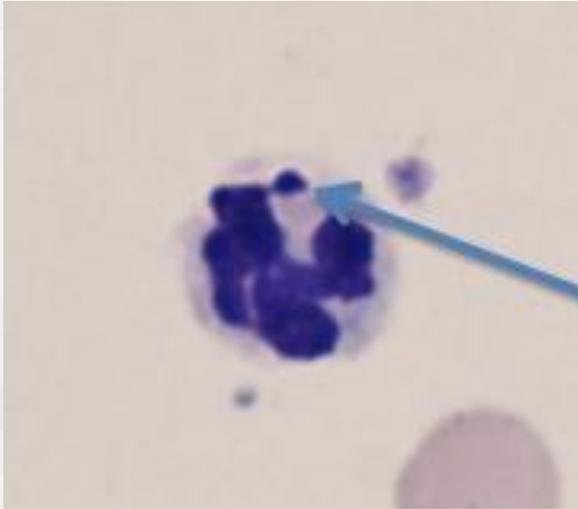
Lymphocytic
leukemia



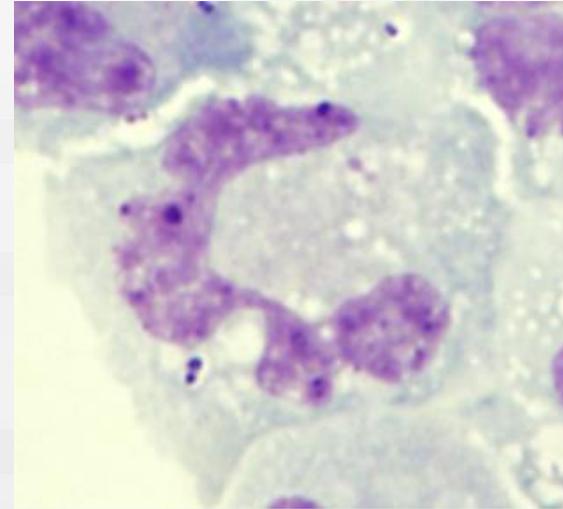


Inclusion Confusion

Barr Bodies

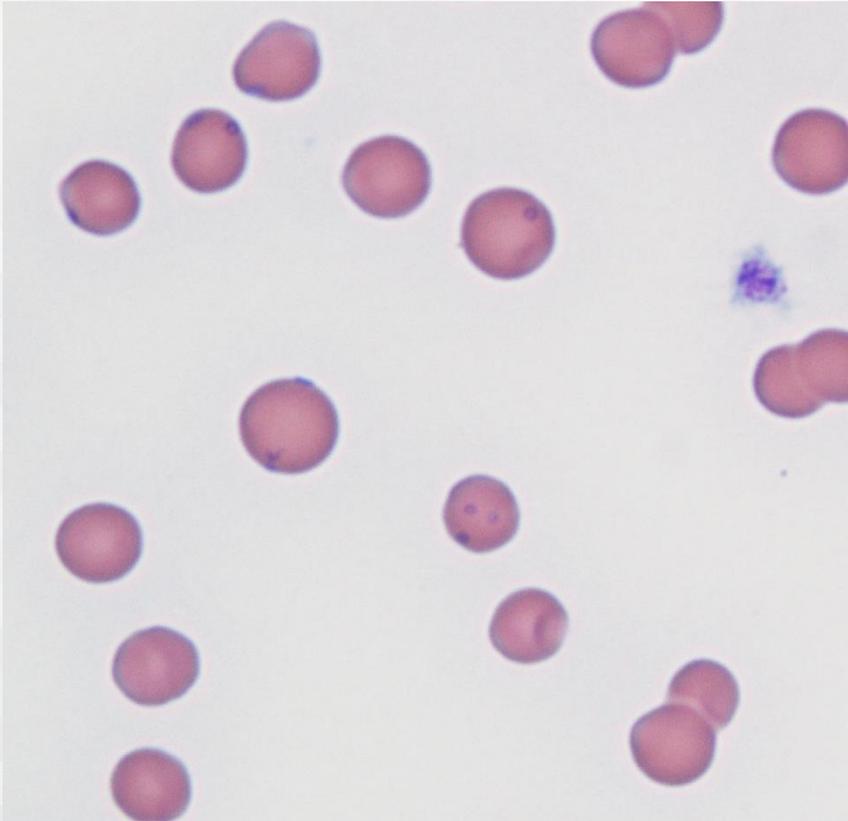


Bacteria

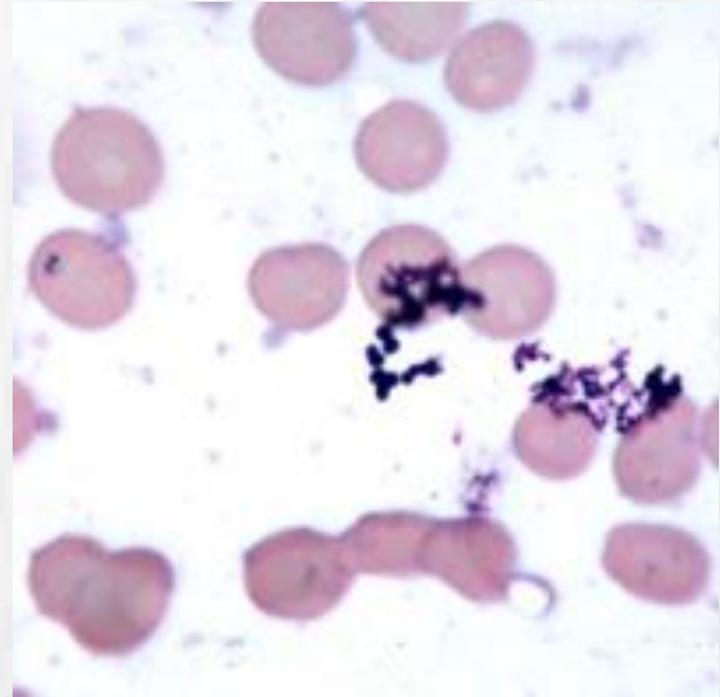


Inclusion Confusion

Mycoplasma hemofelis



Stain precipitate



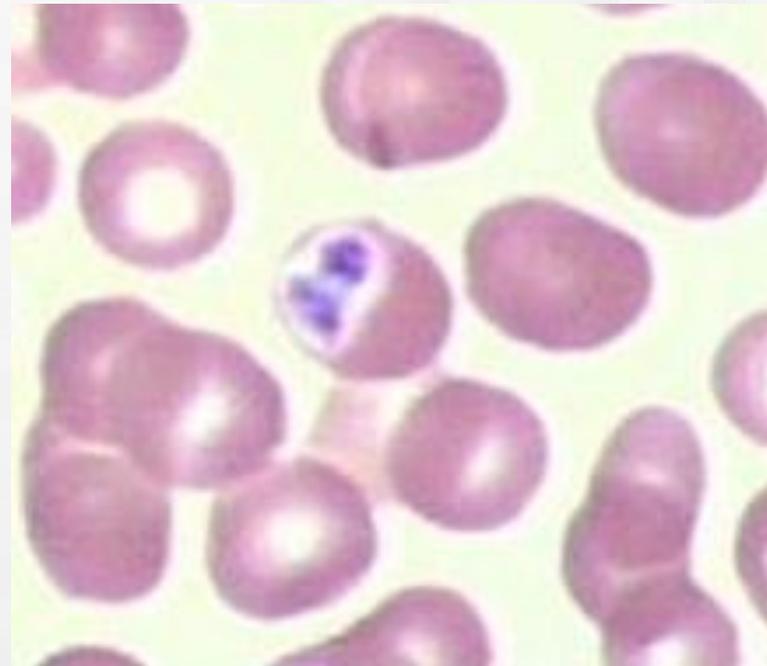
<http://www.eclinpath.com/hematology/sample-collection-heme/stain-precipitate/>

Inclusion Confusion

Anaplasma phagocytophilum



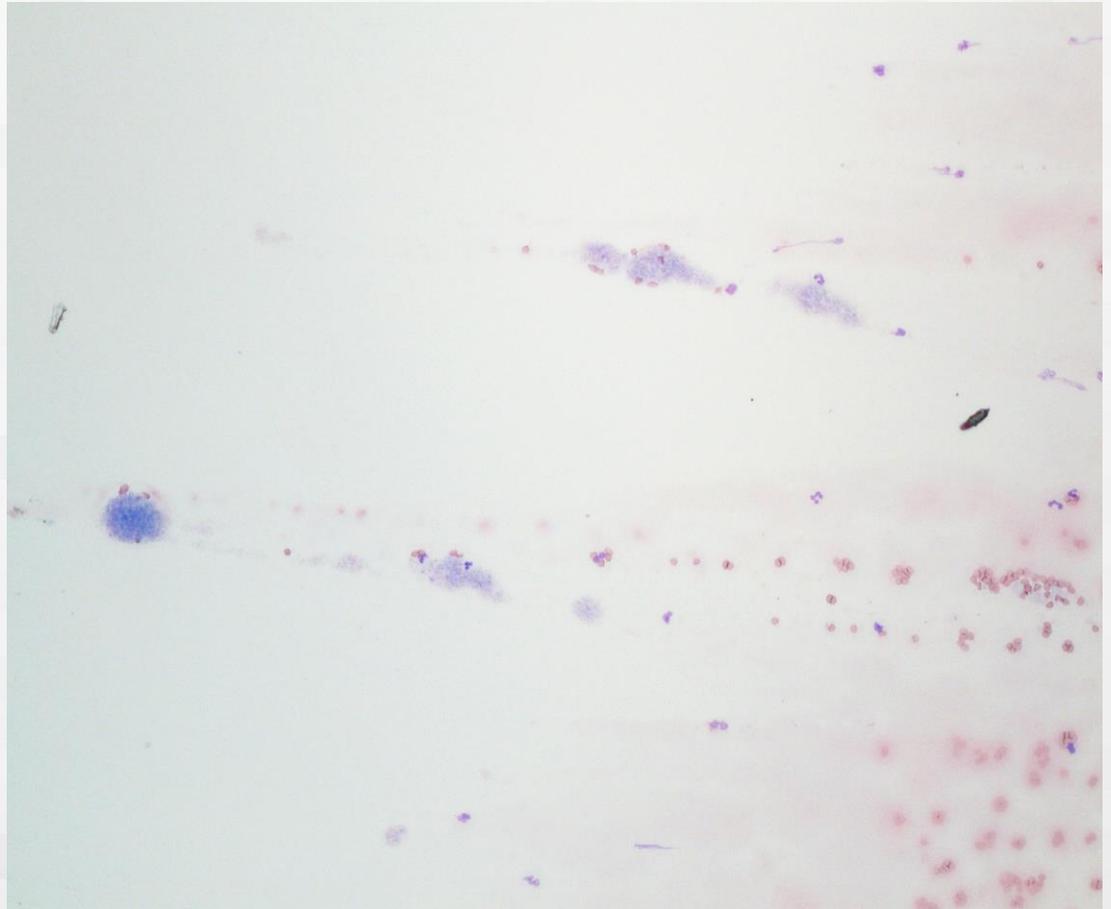
Overlapping platelet



Jacob EA. American Journal of Laboratory Medicine
Volume 1, Issue 3, November 2016, Pages: 34-57

Platelets

- Feathered edge
- 10x

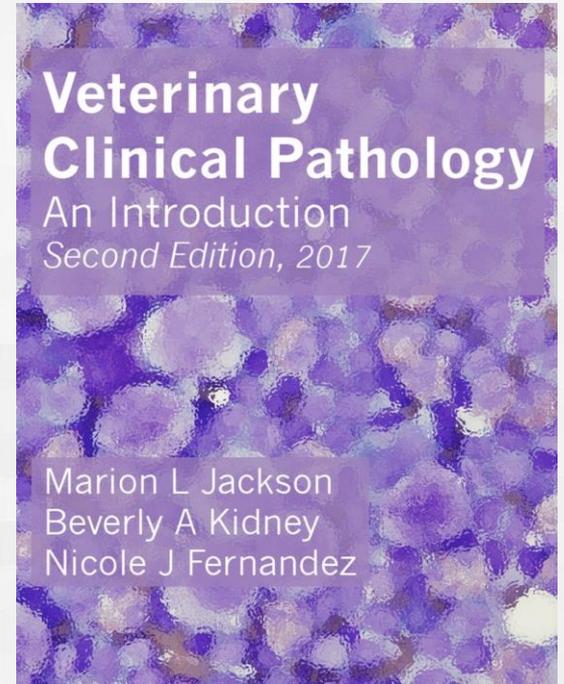


Platelet Estimate

- 10x Scan the feathered edge for platelet clumps
- 100x Count the number of platelets in at least 5 fields of the monolayer and calculate the average for 1 field. Multiply the average for 1 field by $20 \times 10^9/L$.
- If platelets are clumped the formula will **underestimate** the platelet count.
- Less than 3 - 4 per 100x field ($60 - 80 \times 10^9/L$) represents a significant thrombocytopenia

Digital Resources

- Veterinary Clinical Pathology ebook
 - iBooks
 - [Google drive link](#)
- Cornell e-ClinPath
 - <http://www.eclinpath.com/>

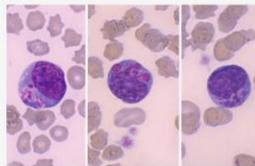


Digital Resources

Cells and Smears

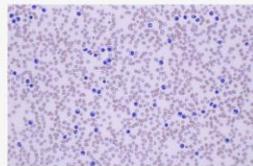
VETERINARY CLINICAL PATHOLOGY DIGITAL DATABASE

HOME HEMATOLOGY ABOUT



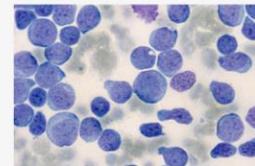
LGL

📅 2 MONTHS AGO



CLL

📅 2 MONTHS AGO



ALL

📅 2 MONTHS AGO

- <https://vetclinpathimages.com/>

Digital Resources

- PDS Protocol Manual
 - Search “pds protocol manual” for most recent version
- Videos
 - FNA & smear making: https://youtu.be/DkXDBom_3JI
 - Fine needle nonaspiration: <https://youtu.be/jgTnqCWewl4>
 - Impression smear making: <https://youtu.be/prggfKrNlbl>
 - Cytology fluid handling: https://youtu.be/Dl_Adsa7mpc
 - Cytology microscopy: <https://youtu.be/vWSD7bjkjs>
 - Urinalysis: https://www.youtube.com/playlist?list=PLm-jtvx5oGMTmnz0GJvPDzK001_AIFz5z

Digital Resources

- Hematology Videos

- Manual WBC count: <https://youtu.be/WwHI1d5pqSc>
- PCV and total protein: <https://youtu.be/cau5wWe4Uds>
- Making/staining blood smears: <https://youtu.be/nbRUiWI2Qrs>
- Blood smear evaluation: <https://youtu.be/wIZtvTGJL6M>



UNIVERSITY OF SASKATCHEWAN

Western College of Veterinary Medicine

DEPARTMENT OF VETERINARY PATHOLOGY
USASK.CA/WCVM

Nicole Fernandez

nicole.fernandez@usask.ca



Patty Voje